



POLYHYDROXYALKANOATE, METHOD FOR PRODUCTION THEREOF  
AND MICROORGANISMS FOR USE IN THE SAME

BACKGROUND OF THE INVENTION

5 Field of the Invention

The present invention relates to a novel polyhydroxyalkanoate (PHA), a method for production of such PHA and microorganisms for use in the same.

Related Background Art

10 Synthetic polymers derived from petroleum have been used as plastics etc. for a long time. Recently, the treatment of the used plastics has become one of serious social problems. These synthetic polymers have advantages of hard-to-decompose have been used in the  
15 place of metal or glass materials. On mass consumption and mass disposal, however, this feature of hard-to-decompose makes them accumulated in waste-disposal facilities, or when they are burned, it causes increased carbon dioxide exhaust, and harmful  
20 substances such as dioxin and endocrine-disruptors may be generated to cause environmental pollution. On the other hand, polyhydroxyalkanoates (PHAs) produced by microorganisms (hereinafter referred to as "microbial polyester") represented by poly-3-hydroxy butyric acid  
25 (PHB) can be used as the conventional plastics to make various kinds of products with melting processes etd., and can be decomposed by organisms unlike oil-derived

synthetic polymers. Therefore, the microbial polyester is bio-decomposed and thus incorporated in the natural material cycle when discarded, and would not remain in the natural environment to cause pollution unlike many conventional synthetic polymer compounds. Furthermore, since the microbial polyesters do not require incineration processes, they are also effective in terms of prevention of air pollution and global warming. Thus, they can be used as a plastic enabling environmental integrity. In addition, the application of the microbial polyesters to medical soft members is under consideration (Japanese Patent Application Laid-Open No. 5-159, Japanese Patent Application Laid-Open No. 6-169980, Japanese Patent Application Laid-Open No. 6-169988, Japanese Patent Application Laid-Open No. 6-225921 and the like).

Heretofore, various bacteria have been reported to produce and accumulate PHB or copolymers of other hydroxyalkanoic acids in the cells ("Biodegradable Plastics Handbook", edited by Biodegradable Plastics Society, issued by NTS Co. Ltd., P178-197, (1995)). It is known that such microbial PHAs may have a variety of compositions and structures depending on types of the producing microorganisms, the composition of culture media, culture conditions and the like, and up to now, studies regarding the control of these compositions and structures have been carried out to improve the

properties of PHA.

For example, *Alcaligenes eutropus* H16 (ATCC No. 17699) and its mutant strains reportedly produce copolymers of 3-hydroxy butyric acid (3HB) and 3-hydroxy valeric acid (3HV) at a variety of composition ratios according to the carbon source in culture (Japanese Patent Publication No. 6-15604, Japanese Patent Publication No. 7-14352, Japanese Patent Publication No. 8-19227 and the like).

10 Japanese Patent Application Laid-Open No. 5-74492 discloses a method in which the copolymer of 3HB and 3HV is produced by bringing *Methylobacterium* sp., *Paracoccus* sp., *Alcaligenes* sp. or *Pseudomonas* sp. into contact primary alcohol having 3 to 7 carbons.

15 Japanese Patent Application Laid-Open No. 5-93049 and Japanese Patent Application Laid-Open No. 7-265065 disclose that two-component copolymers of 3HB and 3-hydroxy hexanoic acid (3HHx) are produced by culturing *Aeromonas caviae* using oleic acid or olive oil as a  
20 carbon source.

Japanese Patent Application Laid-Open No. 9-191893 discloses that *Comamonas acidovorans* IFO 13852 produces polyester having 3HB and 4-hydroxy butyric acid as monomer units in culture with gluconic acid and 1,4-butandiol as a carbon source.  
25

Also, in recent years, active researches about PHA composed of 3-hydroxyalkanoate (3HA) of medium-chain-

length (abbreviated to mcl) having up to about 12 carbons. Synthetic routes can be classified broadly into two types, and their specific examples will be shown in (1) and (2) below.

5 (1) Synthesis using  $\beta$ -oxidation

Japanese Patent No. 2642937 discloses that PHA having monomer units of 3-hydroxyalkanoate having 6 to 12 carbons is produced by providing as a carbon source aliphatic hydrocarbon to *Pseudomonas oleovorans* ATCC  
10 29347. Furthermore, it is reported in Appl. Environ. Microbiol, 58(2), 746 (1992) that *Pseudomonas resinovorans* produces polyester having 3-hydroxybutyric acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid and 3-hydroxydecanoic acid at a ratio of 1:15:75:9 as  
15 monomer units, using octanoic acid as a single carbon source, and also produces polyester having 3-hydroxybutyric acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid and 3-hydroxydecanoic acid (quantitative ratio of 8:62:23:7) as units, using  
20 hexanoic acid as a single carbon source. Herein, it is assumed that 3HA monomer units having longer chain length than that of the starting fatty acid are made by way of fatty acid synthetic route that will be described next in (2).

25 (2) Synthesis using fatty acid synthetic route

It is reported in Int. J. Biol. Macromol., 16(3), 119 (1994) that *Pseudomonas* sp. 61-3 strain produces

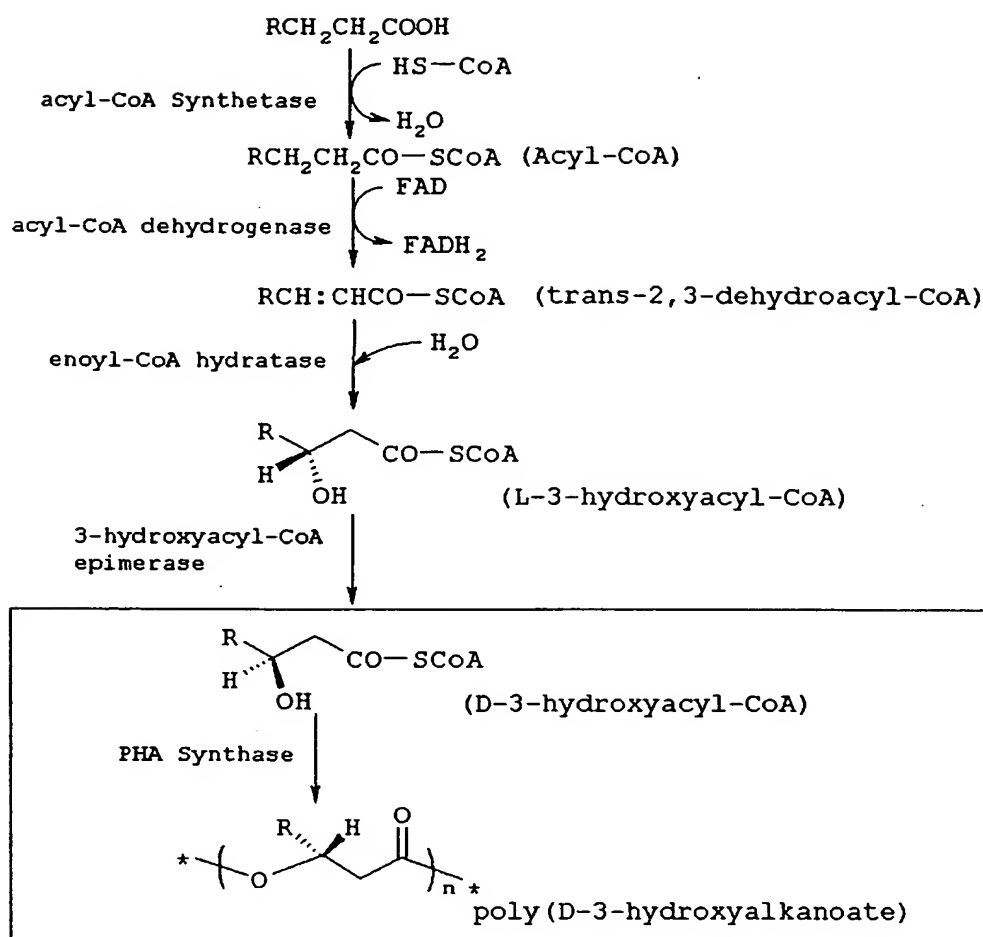


polyester made of 3-hydroxyalkanoic acids such as 3-hydroxybutyric acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid, 3-hydroxydecanoic acid and 3-hydroxydodecanoic acid and 3-hydroxyalkenoic acids such as 3-hydroxy-5-cis-decenoic acid, 3-hydroxy-5-cis-dodecenoic acid, using sodium gluconate as a single carbon source.

By the way, the biosynthesis of PHA is usually carried out by a PHA synthase using as a substrate "D-3-hydroxyacyl-CoA" occurring as an intermediate of a variety of metabolic pathways in the cell.

Herein, "CoA" means a "coenzyme A". And, as described in the prior art of the above (1), the biosynthesis of PHA is carried out with "D-3-hydroxyacyl-CoA" occurring in the " $\beta$  oxidation cycle" being a starting substance in the case where fatty acids such as octanoic acid and nonanoic acid are used as carbon sources.

Reactions through which PHA is synthesized by way of the " $\beta$  oxidation cycle" will be shown below.



On the other hand, as described in the prior art of the above (2), in the case where the PHA is biosynthesized using saccharides such as glucose and the like, the biosynthesis is carried out with "D-3-hydroxyacyl-CoA" converted from "D-3-hydroxyacyl-ACP" occurring in the "fatty acid synthesis pathway" being a starting substance.

Herein, "ACP" means a "acyl carrier protein".

By the way, as described previously, the PHA synthesized in both (1) and (2) described above is PHA

constituted by monomer units having alkyl groups in side chains. However, if a wider range of application of the microbial PHA like this, for example an application as a functional polymer is considered, it is expected that PHA having various substituents (for example phenyl groups) introduced in the side chain is significantly useful. With respect to the synthesis of such PHA, for the synthesis using  $\beta$  oxidation, a report regarding PHA having the aryl group and the like in the side chain can be found in, for example, Macromolecules, 24, p5256-5260 (1991). Specifically, it is reported that *Pseudomonas oleovorans* produces polyester having 3-hydroxy valeric acid, 3-hydroxyheptanoic acid, 3-hydroxynonanoic acid, 3-hydroxyundecanoic acid and 3-hydroxy-5-phenyl valeric acid (quantitative ratio of 0.6:16.0:41.1:1.7:40.6) as units in the amount of 160 mg for 1 L of culture solution (ratio in dry weight to the cell mass is 31.6%), using 5-phenylvaleric acid and nonanoic acid (mole ratio of 2:1, total concentration of 10 mmol/L) as a medium, and also produces polyester having 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid, 3-hydroxydecanoic acid and 3-hydroxy-5-phenyl valeric acid (quantitative ratio of 7.3:64.5:3.9:24.3) as units in the amount of 200 mg for 1 L of culture solution (ratio in dry weight to the cell mass is 39.2%), using 5-phenyl valeric acid and octanoic acid (mole ratio of

1:1, total concentration of 10 mmol/L). It is conceivable that the PHA in this report is principally synthesized by way of the  $\beta$  oxidation pathway due to that fact that nonanoic acid and octanoic acid are  
5 used.

As described above, in the microbial PHA, those with various kinds of compositions/structures are obtained by changing the type of microorganisms for use in its production, culture medium compositions, culture  
10 conditions, but if considering the application of the microbial PHA as plastics, they could not be sufficient yet in terms of properties. In order to further expand the range of the microbial PHA utility, it is important that the improvement of its properties are more widely  
15 considered, and for this purpose, the development and the search of the PHA containing monomer units of further diverse structures, its manufacturing processes and microorganisms enabling desired PHA to be produced efficiently are essential.

20 On the other hand, the PHA of a type having substituents introduced in the side chain as described previously is selected in accordance with the property for which the introduced substituent is desired, thereby making it possible to expect its development as  
25 a "functional polymer" having very useful functions and properties resulting from the property and the like of the introduced substituent, and the development and the

search of excellent PHA allowing such functionality and the biodegradability to be compatible with each other, its manufacturing processes and microorganisms enabling desired PHA to be produced efficiently are also  
5 important challenges.

Another example of such PHA having substituents introduced in the side chain includes PHA having the above described phenyl groups, and further phenoxy groups in the side chain.

10 For another example of phenyl group, it is reported in *Macromolecules*, 29, 1762-1766 (1996) that *Pseudomonas oleovorans* produces PHA including 3-hydroxy-5-(4-toryl) valeric acid as a monomer unit through the culture in a culture medium including 5-(4-  
15 toryl) valeric acid (5-(4-methylphenyl) valeric acid) as a substrate.

Furthermore, it is reported in *Macromolecules*, 32, 2889-2895 (1999) that *Pseudomonas oleovorans* produces PHA including 3-hydroxy-5-(2,4-dinitrophenyl) valeric  
20 acid and 3-hydroxy-5-(4-nitrophenyl) valeric acid as monomer units through the culture in a culture medium including 5-(2,4-dinitrophenyl) valeric acid and nonanoic acid as a substrate.

Also, for an example of the phenoxy group, it is  
25 reported in *Macromol. Chem. Phys.*, 195, 1665-1672 (1994) that *Pseudomonas oleovorans* produces PHA including 3-hydroxy-5-phenoxy valeric acid and 3-

hydroxy-9-phenoxyundecanoic acid as units from 11-phenoxyundecanoic acid.

Also, it is reported in *Macromolecules*, 29, 3432-3435 (1996) that *Pseudomonas oleovorans* is used to produce PHA including 3-hydroxy-4-phenoxybutyric acid and 3-hydroxy-6-phenoxyhexanoic acid as units from 6-phenoxyhexanoic acid, PHA including 3-hydroxy-4-phenoxybutyric acid, 3-hydroxy-6-phenoxyhexanoic acid and 3-hydroxy-8-phenoxyoctanoic acid as units from 8-phenoxyoctanoic acid, and PHA including 3-hydroxy-5-phenoxyvaleric acid and 3-hydroxy-7-phenoxyheptanoic acid as units from 11-phenoxyundecanoic acid. Excerpts of yields of polymers from this report are shown in Table 1.

Furthermore, in *Can. J. Microbiol.*, 41, 32-43 (1995), PHA including 3-hydroxy-p-cyanophenoxyhexanoic acid or 3-hydroxy-p-nitrophenoxyhexanoic acid as a monomer unit is successfully produced with octanoic acid and p-cyanophenoxyhexanoic acid or p-nitrophenoxyhexanoic acid being a substrate, using *Pseudomonas oleovorans* ATCC 29347 and *Pseudomonas putida* KT 2442.

In Japanese Patent No. 2989175, a homopolymer constituted by 3-hydroxy-5-(monofluorophenoxy)pentanoate(3H5(MFP)P) units or 3-hydroxy-5-(difluorophenoxy)pentanoate(3H5(DFP)P) units and a copolymer containing at least 3H5(MFP)P units or

3H5(DFP)P units; *Pseudomonas putida* for synthesizing these polymers; and a method of producing the aforesaid polymers using *Pseudomonas* species are described.

5        These productions are carried out through "two-stage culture" as described below.

Time of culture: First stage, 24 hours ; Second stage, 96 hours

A substrate and a resulting polymer at each stage will be shown below.

10       (1) Resulting polymer : 3H5 (MFP) P homopolymer

Substrate at the first stage : Citric acid, Yeast extract

Substrate at the second stage :

Monofluorophenoxyundecanoic acid

15       (2) Resulting polymer : 3H5 (DFP) P homopolymer

Substrate at the first stage : Citric acid, Yeast extract

Substrate at the second stage :

Difluorophenoxyundecanoic acid

20       (3) Resulting polymer : 3H5 (MFP) P copolymer

Substrate at the first stage : Octanoic acid or Nonanoic acid, Yeast extract

Substrate at the second stage :

Monofluorophenoxyundecanoic acid

25       (4) Resulting polymer : 3H5 (DFP) P copolymer

Substrate at the first stage : Octanoic acid or Nonanoic acid, Yeast extract

Substrate at the second stage :

Difluorophenoxyundecanoic acid

As its effect, a medium-chain-length fatty acid having substituents may be materialized to synthesize a polymer having phenoxy groups with ends of the side chain replaced by one to two fluorine atoms, and stereoregularity and water repellency can be provided while maintaining a high melting point and good processability.

Also, PHA including cyclohexyl groups in monomer units is expected to show polymeric properties different from those of PHA including normal aliphatic hydroxyalkanoic acid as a unit, and an example of production using *Pseudomonas oleovorans* has been reported (Macromolecules, 30, 1611-1615 (1997)).

According to this report, when *Pseudomonas oleovorans* was cultured in a culture medium where nonanoic acid (hereinafter described as NA) and cyclohexylbutyric acid (hereinafter described as CHBA) or cyclohexyl valeric acid (hereinafter described as CHVA) coexisted, PHA including units containing cyclohexyl groups and units originating from nonanoic acid were obtained (each ratio unknown)

About the yields, it is reported that quantitative ratios of CHBA and NA are varied with substrate concentration total of 20 mmol/L and results as shown in Table 2 were obtained.



However, in this example, the yield of polymers per culture solution is not sufficient, and the obtained PHA itself has aliphatic hydroxyalkanoic acid coexist in its monomer unit.

5           In this way, in the case where PHA with a variety of substituents introduced in the side chain is produced, as seen in the reported example of *Pseudomonas oleovorans* described previously and the like, a method is used in which alkanoate having a  
10           substituent to be introduced is used not only as a stock for the polymer but also as a carbon source for growth.

          However, for the method in which alkanoate having a substituent to be introduced is used not only as a  
15           stock for the polymer but also as a carbon source for growth, the supply of an energy source based on the production of the acetyl-CoA by  $\beta$  oxidation from such alkanoate is expected, and in this method, only a  
20           substrate having a certain degree of chain length is capable of producing acetyl-CoA by  $\beta$  oxidation, thus limiting alkanoate that can be used as a substrate of PHA, which is a major problem. Also, generally, since  
25           substrates with the chain length decreased by two methylene chains an after another are newly produced by the  $\beta$  oxidation, and these are captured as monomer units of PHA, the PHA that is synthesized is often a copolymer constituted by monomer units that are

different in the chain length by two methylene chains one after another. In the reported example described above, a copolymer constituted by three types of monomer units, that is 3-hydroxy-8-phenoxyoctanoic acid  
5 originating from 8-phenoxyoctanoic acid which is a substrate, 3-hydroxy-6-phenoxyhexanoic acid and 3-hydroxy-4-phenoxybutyric acid which are by-products originating from metabolites is produced. In this respect, if PHA constituted by single monomer units is  
10 to be obtained, it is quite difficult to use this method. Furthermore, for a method premised on the supply of an energy source based on the production of acetyl-CoA by the  $\beta$  oxidation, the growth of microorganisms is slow and the synthesis of PHA  
15 requires lots of time, and the yield of the synthesized PHA is often low, which is also a major problem.

For this reason, a method in which, in addition to the alkanoate having substituents to be introduced, microorganisms are cultured in the culture medium in  
20 which fatty acids of medium-chain-length and the like such as octanoic acid and nonanoic acid as the carbon source for growth, followed by extracting PHA is considered to be effective and is generally used.

However, according to the study by the inventors,  
25 the PHA synthesized by way of the  $\beta$  oxidation pathway using fatty acids of medium-chain-length such as octanoic acid and nonanoic acid as the carbon source

for growth has poor purity, and 50% or more of the polymers are made of mcl-3HA monomer units originating from the carbon source (for example, 3-hydroxyoctanoic acid, 3-hydroxynonanoic acid and the like). These mcl-3HA units make polymers adhesive at room temperature when they are sole components, and if they coexist in the PHA of the present invention in large quantity, the glass transition temperature (Tg) of the polymer is significantly lowered. Thus, to obtain hard polymers at room temperature, the coexistence of mcl-3HA monomer units is not desired. Also, it is known that such a hetero-side chain structure interferes intra-molecular or inter-molecular interaction originating from the side chain structure, and has significant influence on crystallinity and orientation. For achieving the improvement of polymer properties and the addition of functionality, the coexistence of these mcl-3HA monomer units raises a major problem. Means for solving this problem includes providing a refinement process to separate/remove "undesired" monomer units such as mcl-3HA monomer units originating from the carbon source for growth, in order to acquire PHA constituted by monomer units having only specified substituents. However, the problem is that operations are complicated and a significant decrease in the yield can not be avoided. A more serious problem is that if desired monomer units and undesired monomer units form a

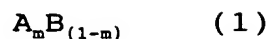
copolymer, it is quite difficult to remove only undesired monomer units. Particularly, in the case where the purpose is to synthesize PHA including monomer units having groups obtained from unsaturated hydrocarbons, ester groups, aryl groups, cyan groups, nitro groups, groups obtained from halogenated hydrocarbons, groups having epoxide and the like introduced therein as side chain structures, the mcl-3HA monomer unit often forms a copolymer with a desired monomer unit, and it is extremely difficult to remove the mcl-3HA monomer unit after PHA is synthesized.

#### SUMMARY OF THE INVENTION

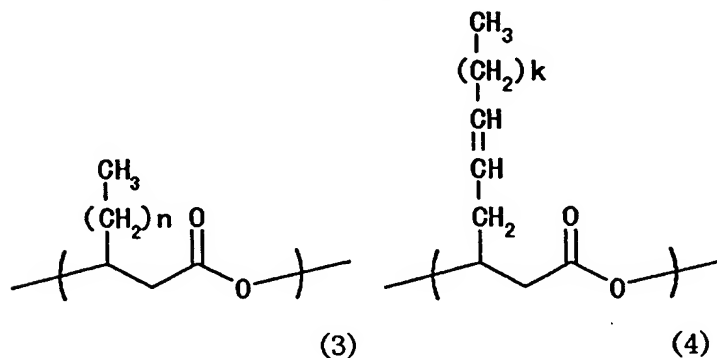
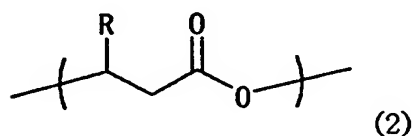
The present invention solves the above described problems, and provides PHA including monomer units of diverse structures having substituents in the side chain, which is useful as device materials, medical materials and the like, and a method of producing such PHA using microorganisms, and particularly a production method in which the coexistence of undesired monomer units is reduced, desired PHA can be obtained in high purity and also in high yields. The present invention is also intended to provide strains enabling such PHA to be synthesized in high purity and with efficiency.

According to one aspect of the present invention, there is provided a polyhydroxyalkanoate having a monomer unit composition represented by a general

formula (1):

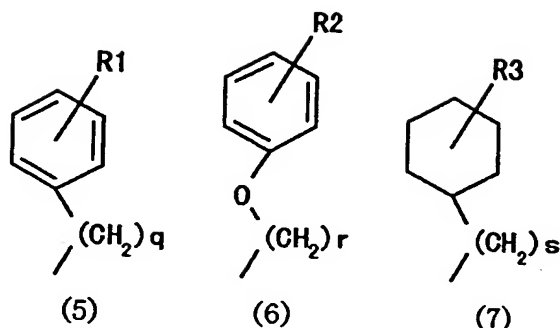


wherein A is represented by General Formula (2), B is at least one selected from the group consisting of monomer units represented by General Formula (3) or (4), and m has a value of 0.01 or larger and smaller than 1:



wherein

n has a value of 0 to 10, k has a value of 3 or 5, and R is at least one group selected from the group consisting of groups represented by General Formulae (5) to (7):



in Formula (5)

R1 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and q is an integer selected from 1 to 8;

in Formula (6)

R2 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and r is an integer selected from 1 to 8;

in Formula (7)

R3 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and s is an integer selected from 1 to 8;

provided that following R are excluded from the choice:

when selecting one type of group as R in the general formula (2):

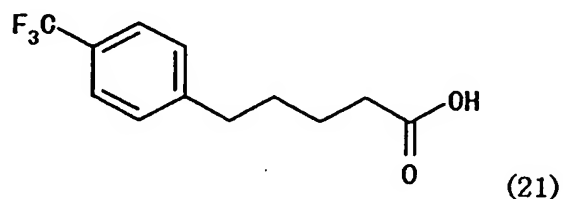
groups of Formula (5) in which R1 is H and q = 2, R1 is H and q = 3, and R1 is -NO<sub>2</sub> and q = 2;

groups of Formula (6) in which R2 is a halogen atom and r = 2, provided that two components are

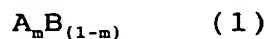
selected as B from General Formula (3) or (4), R<sub>2</sub> is -CN and r = 3, and R<sub>2</sub> is -NO<sub>2</sub> and r = 3; and the groups of Formula (7) in which R<sub>3</sub> is H and s = 1, and R<sub>3</sub> is H and s = 2; and

when selecting two types of groups as R in General Formula (2), the combinations of two types of groups of Formula (6) in which R<sub>2</sub> is a halogen atom and r = 2, and R<sub>2</sub> is a halogen atom and r = 4, provided that one component is selected as B from General Formula (3) or (4).

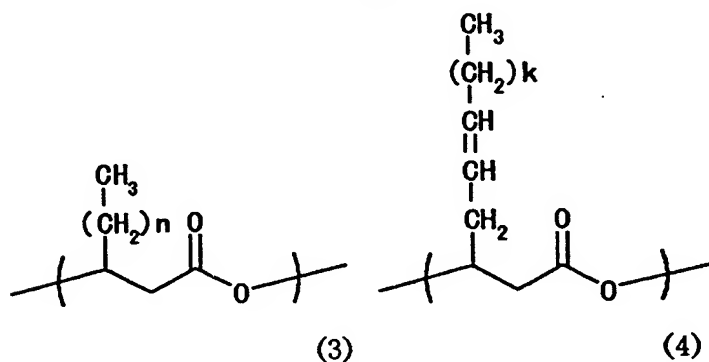
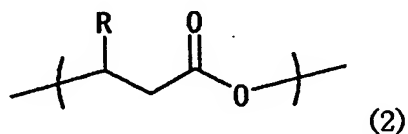
According to one aspect of the present invention, there is provided 5-(4-trifluoromethylphenyl)valeric acid of Formula (21).



According to another aspect of the present invention, there is provided a method of producing a polyhydroxyalkanoate, comprising a step of culturing a microorganism capable of synthesizing a polyhydroxyalkanoate of which monomer unit is represented by Formula (1) from an alkanoate in a medium containing the alkanoate:

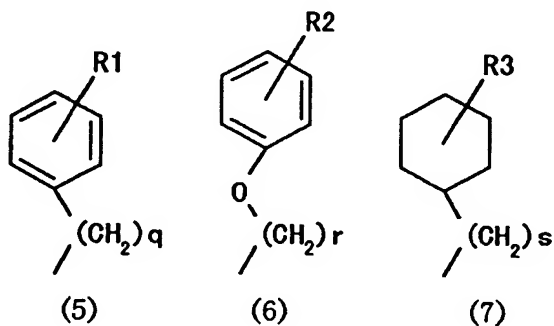


wherein A is represented by General Formula (2), B is at least one selected from the group consisting of monomer units represented by General Formula (3) or (4), and m is 0.01 or larger and smaller than 1,



wherein

n is an integer selected from 0 to 10, k is 3 or 5, and R is at least one group selected from the group consisting of the groups represented by General Formulae (5) to (7):





in Formula (5)

R1 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and q is an integer selected from 1 to 8;

5 in Formula (6)

R2 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and r is an integer selected from 1 to 8;

in Formula (7)

10 R3 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and s is an integer selected from 1 to 8;

provided that following R are excluded from the choice:

15 when selecting one type of group as R in General Formula (2):

groups of Formula (5) in which R1 is H and q = 2, R1 is H and q = 3, and R1 is -NO<sub>2</sub> and q = 2; the groups of Formula (6) in which R2 is a halogen atom and r = 2, R2 is -CN and r = 3, and R2 is -NO<sub>2</sub> and r = 3; and

groups of Formula (7) in which R3 is H and s = 1, and R3 is H and s = 2; and

when selecting two types of groups as R in General Formula (2),

25 groups of Formula (6) in which R2 is a halogen atom and r = 2.

According to a further aspect of the present invention, there is provided a process of producing polyhydroxyalkanoate, comprising a step of culturing a microorganism capable of producing the  
5 polyhydroxyalkanoate utilizing alkanoate in a medium containing the alkanoate and a saccharide.

According to a further aspect of the present invention, there is provided a process of producing polyhydroxyalkanoate, comprising a step of culturing a  
10 microorganism capable of producing a polyhydroxyalkanoate utilizing an alkanoate in a medium containing the alkanoate and a polypeptone.

According to a further aspect of the present invention, there is provided a process of producing  
15 polyhydroxyalkanoate comprising a step of culturing a microorganism capable of producing a polyhydroxyalkanoate utilizing an alkanoate in a medium containing the alkanoate and an organic acid participating in TCA cycle.

20 According to a further aspect of the present invention, there is provided a process of producing polyhydroxyalkanoate, wherein the a microorganism is cultured in at least two steps: one is in a medium containing an alkanoate and a polypeptone and the  
25 subsequent one is in a medium containing the alkanoate and pyruvic acid or salt thereof with nitrogen source limitation.

According to a further aspect of the present invention, there is provided *Pseudomonas cichorii* H45, FERM BP-7374.

5 According to a further aspect of the present invention, there is provided a novel bacterial strain *Pseudomonas cichorii* YN2, FERM BP-7375.

According to a further aspect of the present invention, there is provided a novel bacterial strain *Pseudomonas putida* P91, FERM BP-7373.

10 According to a further aspect of the present invention, there is provided a novel bacterial strain *Pseudomonas jessenii* P161, FERM BP-7376.

As already stated above, the present invention provides novel polyhydroxyalkanoates and novel  
15 substituted alkanoic acids to be a raw material therefor and novel microorganisms which have ability to produce and accumulate in the cell the novel polyhydroxyalkanoates, and provides methods for producing the polyhydroxyalkanoates using such  
20 microorganism. According to them, the polyhydroxyalkanoates useful as functional polymers in which different functional groups are introduced can be manufactured very efficiently and in high purity, therefore it may be expected to be applied to each  
25 field such as device and medical materials.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a chart which shows the measurement of nuclear magnetic resonance spectrum of 3HFPV synthesized in Example 1;

5        FIG. 2 is a chart which shows the measurement of  $^1\text{H}$  nuclear magnetic resonance spectrum of PHA obtained in Example 6;

10        FIG. 3 is a chart which shows the measurement of  $^{13}\text{C}$  nuclear magnetic resonance spectrum of PHA obtained in Example 6;

FIG. 4 is a chart which shows the measurement of nuclear magnetic resonance spectrum of 5-(4-fluorophenoxy)valeric acid obtained in Example 7;

15        FIG. 5 is a chart which shows the Total Ion Chromatography (TIC) of GC-MS for a methyl esterification compound of a monomer unit constituting PHA obtained in Example 8;

20        FIGS. 6A and 6B are charts which show the mass spectra of the peaks on TIC for a methyl esterification compound of a monomer unit constituting PHA obtained in Example 8;

25        FIGS. 7A and 7B are charts which show the mass spectra of the peaks on TIC for a methyl esterification compound of a monomer unit constituting PHA obtained in Example 8;

FIG. 8 is a chart which shows TIC for a methyl esterification compound of a monomer unit constituting

PHA obtained in Example 9;

FIG. 9 is a chart which shows TIC for a methyl esterification compound of a monomer unit constituting PHA obtained in Example 10;

5        FIG. 10 is a chart which shows TIC for a methyl esterification compound of a monomer unit constituting PHA obtained in Example 11;

      FIGS. 11A and 11B are charts which show the Total Ion Chromatography (TIC) of 5-(4-  
10        trifluoromethylphenyl)valeric acid (FIG. 11A) and its mass spectrum (FIG. 11B);

      FIGS. 12A and 12B are charts which show the analytical results of the methylated compounds of PHA copolymers obtained in Example 14. FIG. 12A is the  
15        Total Ion Chromatography (TIC) of the methylated compounds of PHA copolymers and FIG. 12B is the mass spectrum for the peak (around 36.5 min.) containing 3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid which is an objective unit on the TIC;

20        FIGS. 13A and 13B are charts which show the analytical results of the methylated compounds of PHA copolymers obtained in Example 15. FIG. 13A is the Total Ion Chromatography (TIC) of the methylated compounds of PHA copolymers and FIG. 13B is the mass  
25        spectrum for the peak (around 36.5 min.) containing 3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid which is an objective unit on the TIC;

FIG. 14 is a chart which shows the  $^1\text{H}$ -NMR spectrum of PHA obtained in Example 30;

FIG. 15 is a chart which shows the  $^1\text{H}$ -NMR spectrum of PHA obtained in Example 34;

5        FIG. 16 is a chart which shows the  $^1\text{H}$ -NMR spectrum of 4-(4-fluorophenoxy)butyric acid;

FIG. 17 is a chart which shows the  $^{13}\text{C}$ -NMR spectrum of 4-(4-fluorophenoxy)butyric acid;

10        FIG. 18 are charts which show the GC-MS spectrum data measured after methanolysis of PHA recovered from the cultured cells of strain YN2 in Example 41;

FIG. 19 is a chart which shows the  $^1\text{H}$ -NMR spectrum of PHA recovered from the cultured microbial cells of strain YN2 in Example 41;

15        FIG. 20 is a chart which shows the  $^1\text{H}$ -NMR spectrum of 4-(3-fluorophenoxy)butyric acid;

FIG. 21 is a chart which shows the  $^{13}\text{C}$ -NMR spectrum of 4-(3-fluorophenoxy)butyric acid;

20        FIG. 22 are charts which show the GC-MS spectrum data measured after methanolysis of PHA recovered from the cultured microbial cells of strain YN2 in Example 47;

25        FIG. 23 is a chart which shows the  $^1\text{H}$ -NMR spectrum of PHA recovered from the cultured microbial cells of strain YN2 in Example 47;

FIG. 24 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS

measurement in Example 53;

FIG. 25 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-(4-fluorophenoxy)valerate obtained from the GC-MS measurement in Example 53;

5        FIG. 26 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 54;

FIG. 27 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-(4-fluorophenyl)valerate obtained from the GC-MS measurement in Example 54;

10

FIG. 28 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 55;

FIG. 29 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 55;

15

FIG. 30 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-(4-fluorophenoxy)valerate (3HFPxV) obtained from the GC-MS measurement in Example 55;

20

FIG. 31 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 56;

FIG. 32 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 56;

25

FIG. 33 is a chart which shows the mass spectrum

of methyl 3-hydroxy-5-(4-fluorophenyl)valerate (3HFPV) obtained from the GC-MS measurement in Example 56;

FIG. 34 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 57;

FIG. 35 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 57;

FIG. 36 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-(4-fluorophenyl)valerate (3HFPV) obtained from the GC-MS measurement in Example 57;

FIG. 37 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-(4-fluorophenoxy)valerate (3HFPxV) obtained from the GC-MS measurement in Example 57;

FIG. 38 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 58;

FIG. 39 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 58;

FIG. 40 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 58;

FIG. 41 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 58;



FIG. 42 is a chart which shows the mass spectrum of methyl 3-hydroxy-7-phenoxyheptanoate (3HPxHp) obtained from the GC-MS measurement in Example 58;

5        FIG. 43 is a chart which shows the mass spectrum of methyl 3-hydroxy-9-phoxynonanoate (3HPxN) obtained from the GC-MS measurement in Example 58;

FIG. 44 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 59;

10        FIG. 45 is a chart which shows the mass spectrum of methyl 3-hydroxyhexanoate obtained from the GC-MS measurement in Example 59;

FIG. 46 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 59.

FIG. 47 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 59;

20        FIG. 48 is a chart which shows the mass spectrum of methyl 3-hydroxydodecanoate obtained from the GC-MS measurement in Example 59;

FIG. 49 is a chart which shows the mass spectrum of methyl 3-hydroxydodecenoate obtained from the GC-MS measurement in Example 59;

25        FIG. 50 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 59;

FIG. 51 is a chart which shows the mass spectrum of methyl 3-hydroxy-7-phenoxyheptanoate (3HPxHp) obtained from the GC-MS measurement in Example 59;

5      FIG. 52 is a chart which shows the mass spectrum of methyl 3-hydroxy-9-phenoxynonanoate (3HPxN) obtained from the GC-MS measurement in Example 59;

FIG. 53 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 60;

10      FIG. 54 is a chart which shows the mass spectrum of methyl 3-hydroxy-hexanoate obtained from the GC-MS measurement in Example 60;

FIG. 55 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 60;

FIG. 56 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 60;

20      FIG. 57 is a chart which shows the mass spectrum of methyl 3-hydroxydodecanoate obtained from the GC-MS measurement in Example 60;

FIG. 58 is a chart which shows the mass spectrum of methyl 3-hydroxydodecenoate obtained from the GC-MS measurement in Example 60;

25      FIG. 59 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 60;

FIG. 60 is a chart which shows the mass spectrum of methyl 3-hydroxy-7-phenoxyheptanoate (3HPxHp) obtained from the GC-MS measurement in Example 60;

FIG. 61 is a chart which shows the mass spectrum  
5 of methyl 3-hydroxy-9-phoxynonanoate (3HPxN) obtained from the GC-MS measurement in Example 60;

FIG. 62 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 61;

10 FIG. 63 is a chart which shows the mass spectrum of methyl 3-hydroxy-4-phenoxybutyrate (3HPxB) obtained from the GC-MS measurement in Example 61;

FIG. 64 is a chart which shows the mass spectrum of methyl 3-hydroxy-6-phoxyhexanoate (3HPxHx)  
15 obtained from the GC-MS measurement in Example 61;

FIG. 65 is a chart which shows the mass spectrum of methyl 3-hydroxy-8-phoxyoctanoate (3HPxO) obtained from the GC-MS measurement in Example 61;

FIG. 66 is a chart which shows the mass spectrum  
20 of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 62;

FIG. 67 is a chart which shows the mass spectrum of methyl 3-hydroxy-4-phenoxybutyrate (3HPxB) obtained from the GC-MS measurement in Example 62;

25 FIG. 68 is a chart which shows the mass spectrum of methyl 3-hydroxy-6-phoxyhexanoate (3HPxHx) obtained from the GC-MS measurement in Example 62;

FIG. 69 is a chart which shows the mass spectrum of methyl 3-hydroxy-8-phenoxyoctanoate (3HPxO) obtained from the GC-MS measurement in Example 62;

FIG. 70 is a chart which shows the mass spectrum  
5 of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 63;

FIG. 71 is a chart which shows the mass spectrum of methyl 3-hydroxy-octanoate obtained from the GC-MS measurement in Example 63;

10 FIG. 72 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 63;

FIG. 73 is a chart which shows the mass spectrum of methyl 3-hydroxy-4-phenoxybutyrate (3HPxB) obtained  
15 from the GC-MS measurement in Example 63;

FIG. 74 is a chart which shows the mass spectrum of methyl 3-hydroxy-6-phenoxyhexanoate (3HPxHx) obtained from the GC-MS measurement in Example 63;

FIG. 75 is a chart which shows the mass spectrum  
20 of methyl 3-hydroxy-8-phenoxyoctanoate (3HPxO) obtained from the GC-MS measurement in Example 63;

FIG. 76 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 64;

25 FIG. 77 is a chart which shows the mass spectrum of methyl 3-hydroxyhexanoate obtained from the GC-MS measurement in Example 64;

FIG. 78 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 64;

5      FIG. 79 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 64;

FIG. 80 is a chart which shows the mass spectrum of methyl 3-hydroxydodecanoate obtained from the GC-MS measurement in Example 64;

10      FIG. 81 is a chart which shows the mass spectrum of methyl 3-hydroxydodecenoate obtained from the GC-MS measurement in Example 64;

FIG. 82 is a chart which shows the mass spectrum of methyl 3-hydroxy-4-phenoxybutyrate (3HPxB) obtained from the GC-MS measurement in Example 64;

15      FIG. 83 is a chart which shows the mass spectrum of methyl 3-hydroxy-6-phenoxyhexanoate (3HPxHx) obtained from the GC-MS measurement in Example 64;

FIG. 84 is a chart which shows the mass spectrum of methyl 3-hydroxy-8-phenoxyoctanoate (3HPxO) obtained from the GC-MS measurement in Example 64;

20      FIG. 85 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 65;

25      FIG. 86 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 65;

FIG. 87 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 65;

5      FIG. 88 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 65;

FIG. 89 is a chart which shows the mass spectrum of methyl 3-hydroxy-7-phenoxyheptanoate (3HPxHp) obtained from the GC-MS measurement in Example 65;

10      FIG. 90 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 66;.

FIG. 91 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 66;

15      FIG. 92 is a chart which shows the mass spectrum of methyl 3-hydroxy-7-phenoxyheptanoate (3HPxHp) obtained from the GC-MS measurement in Example 66;

FIG. 93 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 67;

20      FIG. 94 is a chart which shows the mass spectrum of methyl 3-hydroxyhexanoate obtained from the GC-MS measurement in Example 67;

25      FIG. 95 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 67;

FIG. 96 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 67;

5        FIG. 97 is a chart which shows the mass spectrum of methyl 3-hydroxydodecanoate obtained from the GC-MS measurement in Example 67;

FIG. 98 is a chart which shows the mass spectrum of methyl 3-hydroxydodecenoate obtained from the GC-MS measurement in Example 67;

10       FIG. 99 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 67;

FIG. 100 is a chart which shows the mass spectrum of methyl 3-hydroxy-7-phenoxyheptanoate (3HPxHp)  
15       obtained from the GC-MS measurement in Example 67;

FIG. 101 is a chart which shows the mass spectrum of methyl 3-hydroxyhexanoate obtained from the GC-MS measurement in Example 68;

FIG. 102 is a chart which shows the mass spectrum  
20       of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 68;

FIG. 103 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 68;

25       FIG. 104 is a chart which shows the mass spectrum of methyl 3-hydroxydodecanoate obtained from the GC-MS measurement in Example 68;

FIG. 105 is a chart which shows the mass spectrum of methyl 3-hydroxydodecenoate obtained from the GC-MS measurement in Example 68;

5        FIG. 106 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 68;

FIG. 107 is a chart which shows the mass spectrum of methyl 3-hydroxy-7-phenoxyheptanoate (3HPxHp) obtained from the GC-MS measurement in Example 68;

10       FIG. 108 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 69;

FIG. 109 is a chart which shows the mass spectrum of methyl 3-hydroxyhexanoate obtained from the GC-MS measurement in Example 69;

15       FIG. 110 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 69;

FIG. 111 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 69;

FIG. 112 is a chart which shows the mass spectrum of methyl 3-hydroxydodecanoate obtained from the GC-MS measurement in Example 69;

25       FIG. 113 is a chart which shows the mass spectrum of methyl 3-hydroxydodecenoate obtained from the GC-MS measurement in Example 69;



FIG. 114 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 69;

5      FIG. 115 is a chart which shows the mass spectrum of methyl 3-hydroxyhexanoate obtained from the GC-MS measurement in Example 70;

FIG. 116 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 70;

10      FIG. 117 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 70;

FIG. 118 is a chart which shows the mass spectrum of methyl 3-hydroxydodecanoate obtained from the GC-MS measurement in Example 70;

15      FIG. 119 is a chart which shows the mass spectrum of methyl 3-hydroxydodecenoate obtained from the GC-MS measurement in Example 70;

FIG. 120 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenoxyvalerate (3HPxV) obtained from the GC-MS measurement in Example 70;

FIG. 121 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS measurement in Example 71;

25      FIG. 122 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 71;

FIG. 123 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenylvalerate (3HPV) obtained from the GC-MS measurement in Example 71;

5 FIG. 124 is a chart which shows the mass spectrum of methyl 3-hydroxy-octanoate obtained from the GC-MS measurement in Example 72;

FIG. 125 is a chart which shows the mass spectrum of methyl 3-hydroxy-5-phenylvalerate (3HPV) obtained from the GC-MS measurement in Example 72;

10 FIG. 126 is a chart which shows the mass spectrum of methyl 3-hydroxybutyrate obtained from the GC-MS measurement in Example 73;

FIG. 127 is a chart which shows the mass spectrum of methyl 3-hydroxyoctanoate obtained from the GC-MS  
15 measurement in Example 73;

FIG. 128 is a chart which shows the mass spectrum of methyl 3-hydroxydecanoate obtained from the GC-MS measurement in Example 73; and

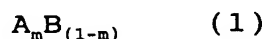
FIG. 129 is a chart which shows the mass spectrum  
20 of methyl 3-hydroxy-5-phenylvalerate (3HPV) obtained from the GC-MS measurement in Example 73.

#### DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

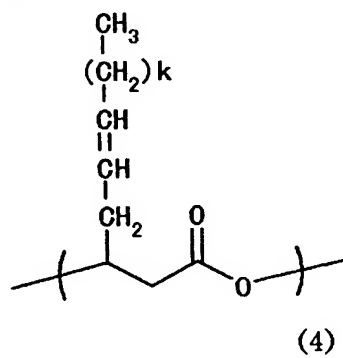
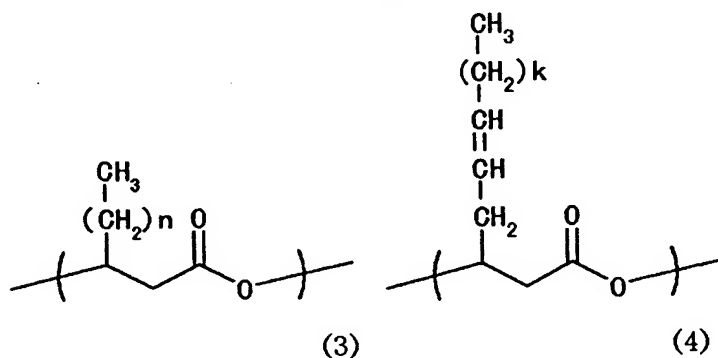
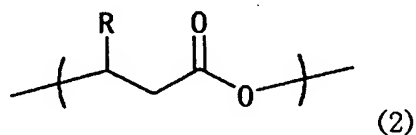
For solving the above described problems, the  
25 inventor et al. strenuously carried out researches for search of innovative microorganisms capable of producing PHA and accumulating the same in the cells

and a method of manufacturing desired PHA using the innovative microorganism, particularly with the aim of developing PHA having substituted or unsubstituted phenoxy groups, phenyl groups and cyclohexyl groups on the side chain, which is useful as device materials and medical materials, and completed the invention.

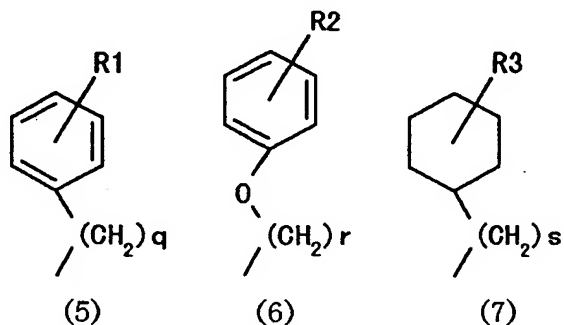
The polyhydroxyalkanoate of the present invention is characterized by having a monomer unit composition represented by formula (1).



(wherein A is represented by formula (2), B is at least one or more selected from monomer units represented by formula (3) or (4), and m is 0.01 or more and less than 1).



(In formulae, n is 0 to 10, k is 3 or 5, and R is at least one or more groups selected from groups represented by formulae (5) to (7)).



(In formula (5), R1 is a group selected from hydrogen atom (H), halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>, and q is selected from integers of 1 to 8; In formula (6), R2 is a group selected from hydrogen atom (H), halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>, and r is selected from integers of 1 to 8; In formula (7), R3 is a group selected from hydrogen atom (H), halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>, and s is selected from integers of 1 to 8;

wherein, when one kind of group is selected, as R in formula (2),

the group of R1 = H and q = 2, the group of R1 = H and q = 3, and the group of R1 = -NO<sub>2</sub> and q = 2 in formula (5),

the group of R2 = halogen atom and r = 2 [however, only when two components are selected from formula (3) or (4) as the above described B], the group of R2 = -CN and r = 3, and the group of R2 = -NO<sub>2</sub> and r = 3 in formula (6), and

the group of R3 = H and s = 1 and the group of R3

= H and s = 2 in formula (7)

are excluded from alternatives, and

when two kinds of groups are selected,

in formula (6), a combination of two kinds of

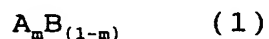
5 groups of R<sub>2</sub> = halogen atom and r = 2 and 4 [however,  
only when one component is selected from formula (3) or  
(4) as the above described B]

are excluded from alternatives).

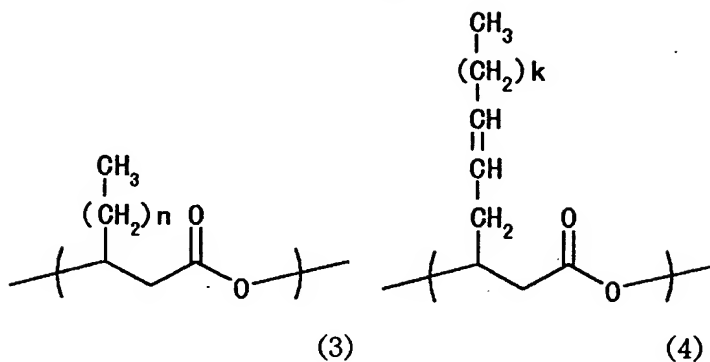
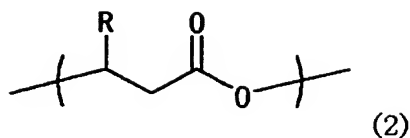
Herein, the polyhydroxyalkanoate of the present  
10 invention may include more than two kinds of monomer  
units represented by formula (2), but it is preferably  
designed so that the appropriate number of monomer  
units are included, considering the needed polymer's  
functionality and properties. Generally, when  
15 alkanoates up to about five kinds are used as a raw  
material of desired monomer units, "secondary"  
substrates with the chain length sequentially decreased  
by two methylene units are newly produced by  $\beta$   
oxidation from part of the alkanoates, as described  
20 previously, and captured as monomer units of the PHA.  
Thus, about ten kinds or less of monomer units  
represented by formula (2) are included in PHA and it  
is expected that the object of the present invention be  
sufficiently achieved. Furthermore, if fine control of  
25 the functionality and the property is desired,  
configuration with more species of monomer units is  
also possible.

Also, with respect to substituted positions of R1, R2 and R3, for any of a ortho, meta or para position, and for a first position in the case of the cyclohexyl group of R3, polyhydroxyalkanoate containing  
5 corresponding monomer units can be configured, but if there is no significant differences in functionality and properties for any isomer, it is advantageously configured with constituents at the meta or para position in terms of yields or ease with which it is  
10 captured in the polymer.

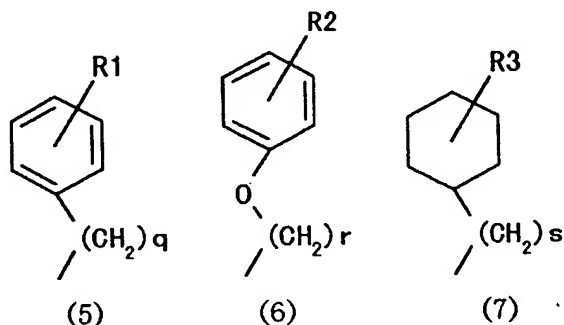
Also, the method of manufacturing polyhydroxyalkanoate of the present invention is a method of manufacturing polyhydroxyalkanoate having monomer unit composition represented by formula (1)  
15 using microorganisms, characterized in that microorganisms are cultured together with the alkanoate and the polyhydroxyalkanoate is extracted from the cells of organisms to obtain the polyhydroxyalkanoate having monomer unit composition represented by formula  
20 (1).



(wherein A is represented by formula (2), B is at least one or more selected from monomer units represented by formula (3) or (4), and m is 0.01 or  
25 more and less than 1).



(In formulae,  $n$  is 0 to 10,  $k$  is 3 or 5, and  $R$  is at least one or more groups selected from groups represented by formulae (5) to (7)).



(In formula (5),  $R_1$  is a group selected from hydrogen atom (H), halogen atom,  $-\text{CN}$ ,  $-\text{NO}_2$ ,  $-\text{CF}_3$ ,  $-\text{C}_2\text{F}_5$  and  $-\text{C}_3\text{F}_7$ , and  $q$  is selected from integers of 1 to 8;

In formula (6),  $R_2$  is a group selected from hydrogen atom (H), halogen atom,  $-\text{CN}$ ,  $-\text{NO}_2$ ,  $-\text{CF}_3$ ,  $-\text{C}_2\text{F}_5$  and  $-\text{C}_3\text{F}_7$ , and  $r$  is selected from integers of 1 to 8;

In formula (7),  $R_3$  is a group selected from

hydrogen atom (H), halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>, and s is selected from integers of 1 to 8;

wherein, when one kind of group is selected, as R in formula (2),

5        the group of R1 = H and q = 2, the group of R1 = H and q = 3, and the group of R1 = -NO<sub>2</sub> and q = 2 in formula (5),

         the group of R2 = halogen atom and r = 2, the group of R2 = -CN and r = 3, and the group of R2 = -NO<sub>2</sub> and r = 3 in formula (6), and

10        the group of R3 = H and s = 1 and the group of R3 = H and s = 2 in formula (7)

are excluded from alternatives, and

when two kinds of groups are selected,

15        in formula (6), a combination of two kinds of groups of R2 = halogen atom and r = 2 are excluded from alternatives).

         Herein, the polyhydroxyalkanoate of the present invention may more than two kinds of monomer units represented by formula (2), but it is preferably synthesized so that the appropriate number of monomer units are included, considering the needed polymer's functionality and properties. Generally, when alkanoates of up to about five kinds are used as a raw material for desired substrates, "secondary" substrates with the chain length shortened by two methylene units are newly produced by  $\beta$  oxidation from the alkanoates



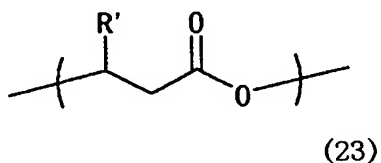
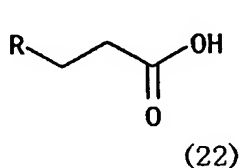
as described previously, and captured as monomer units in PHA. Thus, monomer units of up to ten kinds represented by formula (2) are included in PHA, and it is expected that the object of the present invention is  
5 sufficiently achieved. Furthermore, if fine control of the functionality and the property is desired, culture may be performed to include more species of monomer units.

Also, with respect to substituted positions of R1,  
10 R2 and R3, for any of a ortho, meta or para position, and for a first position in the case of the cyclohexyl group of R3, polyhydroxyalkanoate containing corresponding monomer units can be configured, but if there is no significant differences in functionality  
15 and properties for any isomer, it is advantageously configured with constituents at the meta or para position in terms of yields or ease with which it is captured in the polymer.

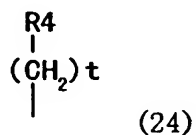
Furthermore, the inventors did strenuous research  
20 to develop a method for obtaining desired PHA having little or no undesired monomer units coexisting therein, and as a result, found that when the microorganism is cultured in a culture medium containing as the sole carbon source the alkanoate to  
25 be the raw material for PHA and a saccharide, it is possible to produce PHA having little or no undesired monomer units coexisting therein, leading to the

completion of the invention.

That is, the method of manufacturing polyhydroxyalkanoate (PHA) of the present invention is a method of manufacturing polyhydroxyalkanoate containing monomer units represented by formula (23) using microorganisms, characterized by having a process in which a microorganism capable of synthesizing polyhydroxyalkanoate containing monomer units represented by formula (23) from the alkanoate represented by general formula (22) are cultured in a culture medium containing as the sole carbon source the alkanoate of formula (22) and a saccharide.



(In above formulae, R is at least one or more groups represented by formula (24), and R' is one or more groups selected from the groups selected in formula (22), the group of t-2 in the selected groups, the group of t-4 in the selected groups and the group of t-6 in the selected groups. Herein, t-2, t-4 and t-6 can be only integers of 1 or more.)



(In the above formula, R4 represents a saturated

or unsaturated phenyl group, a saturated or unsaturated phenoxy group, and a saturated or unsaturated cyclohexyl group, and t represents an integer in the range of 1 to 8 independently.)

5           Herein, more than one kinds of alkanoate represented by formula (22) may be used when culture is carried out, but the appropriate number thereof are preferably used, considering the needed polymer's functionality and properties. Generally, when  
10   alkanoates of up to five kinds represented by formula (22) are used as raw materials for the desired monomer units, "secondary" substrates with the chain length shortened by two methylene units are newly produced by  $\beta$  oxidation from part of the alkanoates as described  
15   previously, and captured as monomer units of PHA. Thus, monomer units of up to ten kinds represented by formula (2), for example, are included in PHA, and it is expected that the above described object be sufficiently achieved. Furthermore, if fine control of  
20   the functionality and the property is desired, more kinds of alkanoates can be used.

          Substituents at the group of R4 described above include halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub>, -C<sub>3</sub>F<sub>7</sub> and the like. With respect to substituted positions of R4,  
25   in any of a ortho, meta or para position, and in a first position in the case of the cyclohexyl group, polyhydroxyalkanoate constituted by corresponding

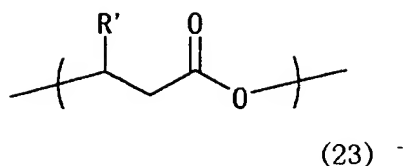
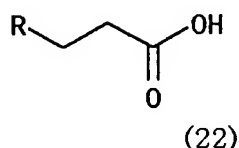
monomer units can be obtained, but if there is no significant differences in functionality and properties for any isomer, constituents at the meta or para position can be suitably used in terms of yields or  
5 ease with which it is captured in the polymer.

Also, for saccharides, for example, glucose, fructose, mannose and the like may be suitably used.

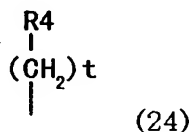
Furthermore, the inventors did strenuous research to develop a method for producing desired PHA having  
10 little or no undesired monomer units coexisting therein, and as a result, found that when the microorganism is cultured in a culture medium containing as the sole carbon source an alkanoate to be the raw material for PHA and polypeptone only, it is  
15 possible to produce PHA having little or no undesired monomer units coexisting therein, leading to the completion of the invention.

That is, the method of manufacturing polyhydroxyalkanoate (PHA) of the present invention is  
20 a method of manufacturing polyhydroxyalkanoate containing monomer units represented by formula (23) using microorganisms, characterized by having a process in which microorganisms capable of synthesizing polyhydroxyalkanoate containing monomer units  
25 represented by formula (23) from the alkanoate represented by formula (22) are cultured in a culture medium containing the alkanoate and polypeptone as the

only carbon source.



(In the above described formulae, R is at least one or more groups represented by formula (24), and R' is one or more groups selected from the groups selected in formula (22), the group of t-2 in the selected groups, the group of t-4 in the selected groups and the group of t-6 in the selected groups. Herein, t-2, t-4 and t-6 can be only integers of 1 or more.)



(In the above described formula, R4 represents a saturated or unsaturated phenyl group, a saturated or unsaturated phenoxy group, and a saturated or unsaturated cyclohexyl group, and t represents an integer in the range of 1 to 8 independently.)

Herein, more than one kinds of alkanoate represented by formula (22) may be used when culture is carried out, but the appropriate number thereof are preferably used, considering the needed polymer's functionality and properties. Generally, when up to five kinds of alkanoates represented by formula (22) are used as raw materials for the desired monomer

units, secondary" substrates with the chain length shortened by two methylene units are newly produced by  $\beta$  oxidation from a part of the alkanoates as described previously, and captured as the monomer units in PHA.

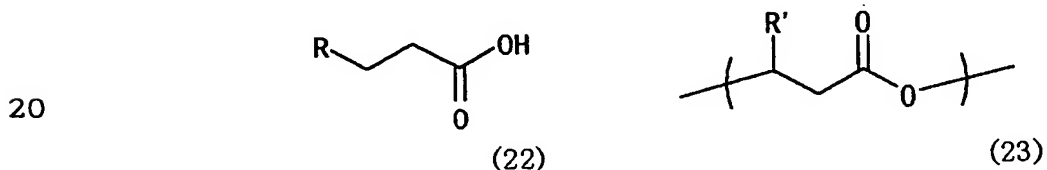
5 Thus, up to about ten kinds of monomer units represented by formula (2), for example, are included in PHA, and it is expected that the above described object be sufficiently achieved. Furthermore, if fine control of the functionality and the property is  
10 desired, more kinds of alkanoate can be used.

Substituents at the group of R4 described above include halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub>, -C<sub>3</sub>F<sub>7</sub> and the like. With respect to substituted positions of R4, in any of a ortho, meta or para position, and in a  
15 first position in the case of the cyclohexyl group, polyhydroxyalkanoate constituted by corresponding monomer units can be obtained, but if there is no significant differences in functionality and properties for any isomer, constituents at the meta or para  
20 position can be suitably used in terms of yields or ease with which it is captured in the polymer.

Furthermore, the inventors did strenuous research to develop a method for efficiently producing desired PHA having little or no undesired monomer units  
25 coexisting therein, and as a result, found that when the microorganism are cultured in a culture medium containing as the sole carbon source an alkanoate to be

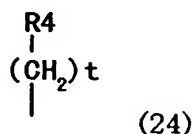
the raw material for PHA and an organic acid associated with the TCA cycle, it is possible to produce PHA having little or no undesired monomer units coexisting therein, leading to the completion of the invention.

5           That is, the method of producing polyhydroxyalkanoate (PHA) of the present invention is a method of producing polyhydroxyalkanoate containing monomer units represented by formula (23) using microorganisms, characterized by having a process in  
10           which microorganisms capable of synthesizing polyhydroxyalkanoate containing monomer units represented by formula (23) from the alkanoate represented by the following general formula (22) are cultured in a culture medium including the alkanoate  
15           and only an organic acid associated with the TCA cycle as a carbon source other than the alkanoate represented by the following formula (22).



(In the above described formulae, R is at least one or more groups represented by formula (24), and R' is one or more groups selected from the groups selected  
25           in formula (22), the group of t-2 in the selected groups, the group of t-4 in the selected groups and the group of t-6 in the selected groups. Herein, t-2, t-4

and t-6 can be only integers of 1 or more.)



(In the above described formula, R4 represents a saturated or unsaturated phenyl group, a saturated or unsaturated phenoxy group, and a saturated or unsaturated cyclohexyl group, and t represents an integer in the range of 1 to 8 independently.)

Herein, more than one kinds of alkanoate represented by formula (22) may be used when culture is carried out, but the appropriate number thereof are preferably used, considering the needed polymer's functionality and properties. Generally, when up to about five kinds of alkanoates represented by formula (22) are used as the raw materials, "secondary" substrates with the chain length shortened by two methylene units are newly produced by  $\beta$  oxidation from a part of the alkanoates as described previously, and captured as the monomer units of PHA. Thus, up to about ten kinds of monomer units, for example, as represented by formula (2), are included in PHA, and it is expected that the above described object be sufficiently achieved. Furthermore, if fine control of the functionality and the property is desired, more kinds of alkanoates can be used.

Substituents at the group of R4 described above



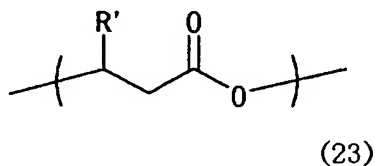
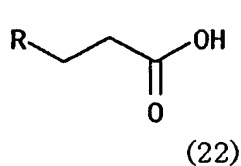
include halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub>, -C<sub>3</sub>F<sub>7</sub> and the like. With respect to substituted positions of R<sub>4</sub>, in any of a ortho, meta or para position, and in a first position in the case of the cyclohexyl group, 5 polyhydroxyalkanoate constituted by corresponding monomer units can be obtained, but if there is no significant differences in functionality and properties for any isomer, constituents at the meta or para position can be suitably used in terms of yields or 10 ease with which it is captured in the polymer.

Also, for organic acids associated with the TCA cycle, organic acids existing in the TCA cycle itself, for example citric acid, succinic acid, fumaric acid, malic acid and salts thereof, and organic acids 15 existing on the main flux to the TCA cycle, for example lactic acid, pyruvic acid and salts thereof may be suitably used.

Furthermore, the inventors did strenuous research to develop a method for obtaining desired PHA having 20 little or no undesired monomer units coexisting therein efficiently, and as a result, found that when microorganisms are cultured in at least two steps: first in a medium containing as the sole carbon source an alkanoate to be a raw material for PHA and 25 polypeptone, and then in a medium containing the alkanoate and pyruvic acid or salts thereof as the sole carbon source with nitrogen limitation, it is possible

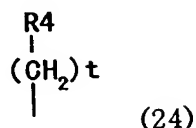
to produce PHA having little or no undesired monomer units coexisting therein.

That is, the method of producing polyhydroxyalkanoate (PHA) of the present invention is a method of manufacturing polyhydroxyalkanoate containing monomer units represented by formula (23) using microorganisms, characterized by having a process in which microorganisms capable of synthesizing polyhydroxyalkanoate containing monomer units represented by formula (23) from the alkanoate represented by the following general formula (22) are cultured in at least two steps: first in a culture medium containing as the sole carbon source the alkanoate represented by formula (22) and polypeptone, and then in a culture medium containing as the sole carbon source the alkanoate represented by formula (22) and pyruvic acid or salts thereof under nitrogen limitation.



(In the above described formulae, R is at least one or more groups represented by formula (24), and R' is one or more groups selected from the groups selected in the above described formula (22), the group of t-2 in the selected groups, the group of t-4 in the

selected groups and the group of t-6 in the selected groups. Herein, t-2, t-4 and t-6 can be only integers of 1 or more.)

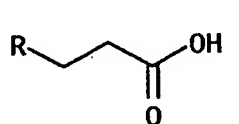


(In the above described formula, R4 represents a saturated or unsaturated phenyl group, a saturated or unsaturated phenoxy group, and a saturated or unsaturated cyclohexyl group, and t represents an integer in the range of 1 to 8 independently.)

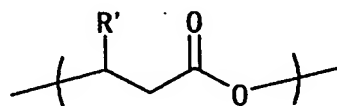
Herein, more than one kinds of alkanoate represented by formula (22) may be used when culture is carried out, but the appropriate number thereof are preferably used, considering the needed polymer's functionality and properties. Generally, when up to about five kinds of alkanoates represented by formula (22) are used as the raw materials, "secondary" substrates shortened by two methylene units are newly produced by  $\beta$  oxidation from part of the alkanoates as described previously, and captured as the monomer units in PHA, up to about ten kinds of monomer units, for example, those represented by formula (2), are included in PHA, and it is expected that the above described object be sufficiently achieved. Furthermore, if fine control of the functionality and the property is desired, more kinds of alkanoate can be used.

Substituents at the group of R4 described above include halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub>, -C<sub>3</sub>F<sub>7</sub>, and the like. With respect to substituted positions of R4, in any of a ortho, meta or para position, and in a first position in the case of the cyclohexyl group, polyhydroxyalkanoate constituted by corresponding monomer units can be obtained, but if there is no significant differences in functionality and properties for any isomer, constituents at the meta or para position can be suitably used in terms of yields or ease with which it is captured in the polymer.

Also, new strains related to the present invention are characterized by having synthetic systems of polyhydroxyalkanoate including alkanoate represented by formula (22) to monomer units represented by formula (23).



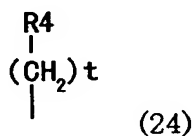
(22)



(23)

(In the above described formulae, R is at least one or more groups represented by formula (24), and R' is one or more groups selected from the groups selected in the above described formula (22), the group of t-2 in the selected groups, the group of t-4 in the selected groups and the group of t-6 in the selected groups. Herein, t-2, t-4 and t-6 can be only integers

of 1 or more.)

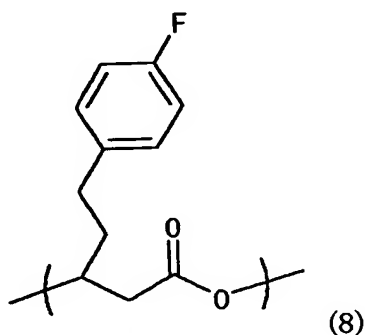


(wherein R4 is a substituted or unsubstituted phenyl group, a substituted or unsubstituted phenoxy group, or a substituted or unsubstituted cyclohexyl group; and t is independently an integer of 1-8).

A substituent in the above described R4 group includes a halogen atom, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub>, -C<sub>3</sub>F<sub>7</sub> or the like.

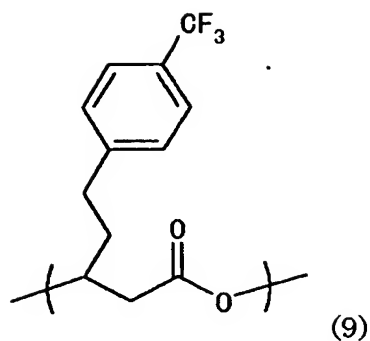
Hereinafter, a new polyhydroxyalkanoate of the present invention will be illustrated.

A new polyhydroxyalkanoate of the present invention is the one of formula (8):



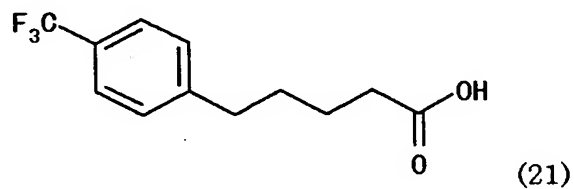
which contains 3-hydroxy-5-(4-fluorophenyl)valeric acid as a monomer unit.

Further, the polyhydroxyalkanoate is of formula (9):



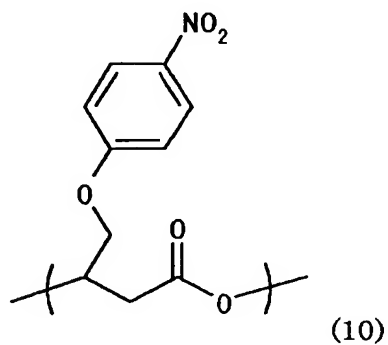
which contains 3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid as a monomer unit.

In addition, a material substrate when producing new PHA of the present invention using microorganisms is 5-(4-trifluoromethylphenyl) valeric acid of the following chemical formula (21):



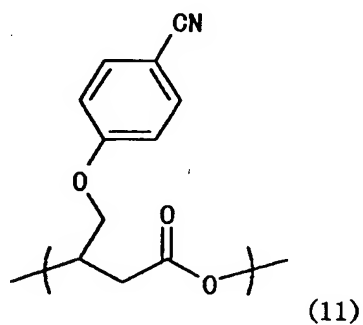
and the material substrate itself is a new compound.

Further, the polyhydroxyalkanoate is of formula (10):



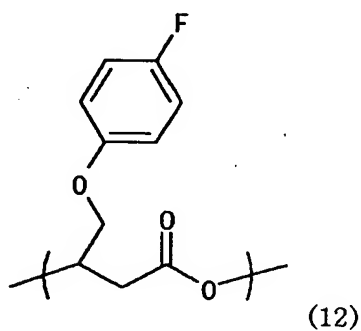
which contains 3-hydroxy-4-(4-nitrophenoxy)butyric acid as a monomer unit.

Further, the polyhydroxyalkanoate is of formula (11):



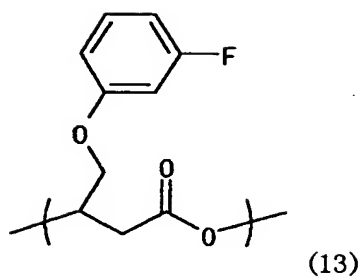
which contains 3-hydroxy-4-(4-cyanophenoxy)butyric acid as a monomer unit.

Further, the polyhydroxyalkanoate is of formula (12):



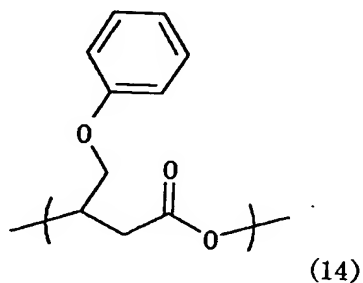
which contains 3-hydroxy-4-(4-fluorophenoxy)butyric acid as a monomer unit.

Further, the polyhydroxyalkanoate is of formula (13):



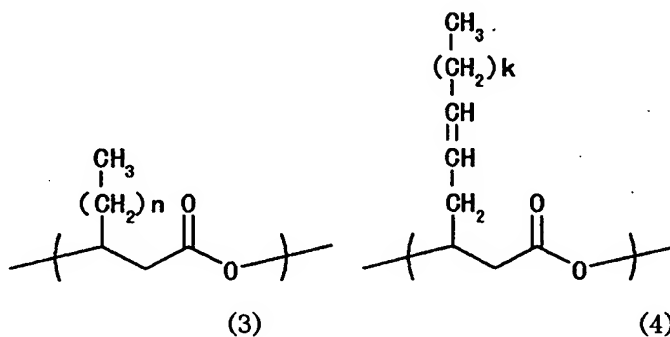
which contains 3-hydroxy-4-(3-fluorophenoxy)butyric acid as a monomer unit.

Further, the polyhydroxyalkanoate is of formula (14):



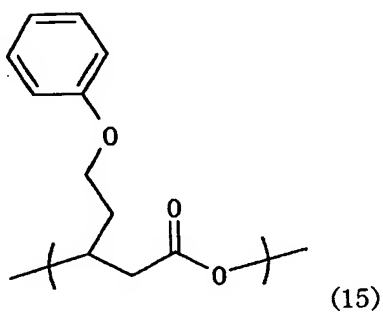
which contains 3-hydroxy-4-phenoxybutyric acid as a monomer unit. Herein, as the monomer unit except 3-hydroxy-4-phenoxybutyric acid of formula (14), at least one or more of the monomer units of formula (3) or (4) are contained.



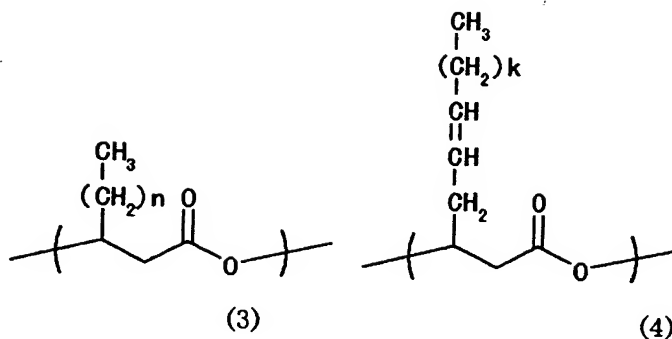


(wherein n is 0-10)(wherein k is 3 or 5)

Further, the polyhydroxyalkanoate is of formula (15):

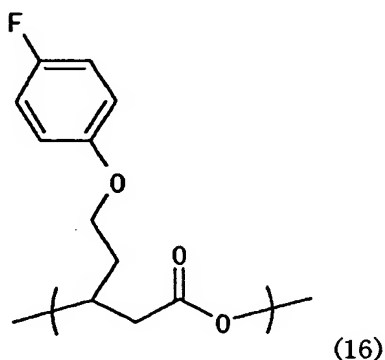


which contains 3-hydroxy-5-phenoxyvaleric acid as a monomer unit. Herein, as the monomer unit except 3-hydroxy-5-phenoxyvaleric acid of formula (15), at least one or more of the monomer units of formula (3) or (4) are contained.

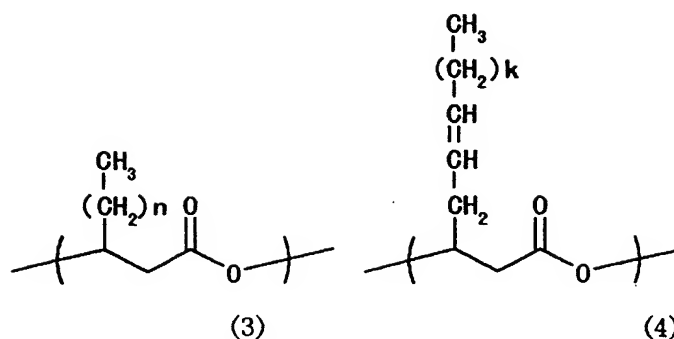


(wherein n is 0-10)(wherein k is 3 or 5)

Further, the polyhydroxyalkanoate is of formula (16):

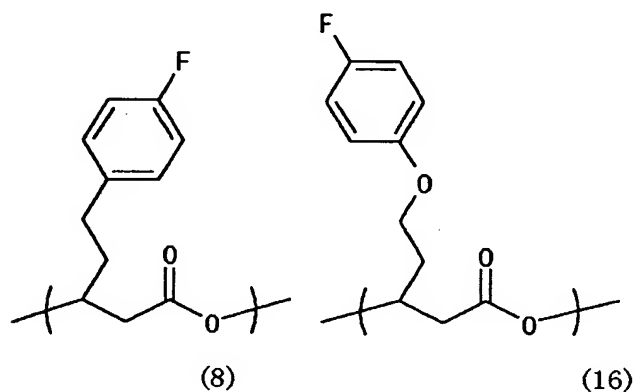


which contains 3-hydroxy-5-(4-fluorophenoxy)valeric acid as a monomer unit. Herein, as the monomer unit except 3-hydroxy-5-(4-fluorophenoxy)valeric acid of formula (16), the polyhydroxyalkanoate contains at least one or more of the monomer units of formula (3) or (4) and excludes the monomer unit of three component system.



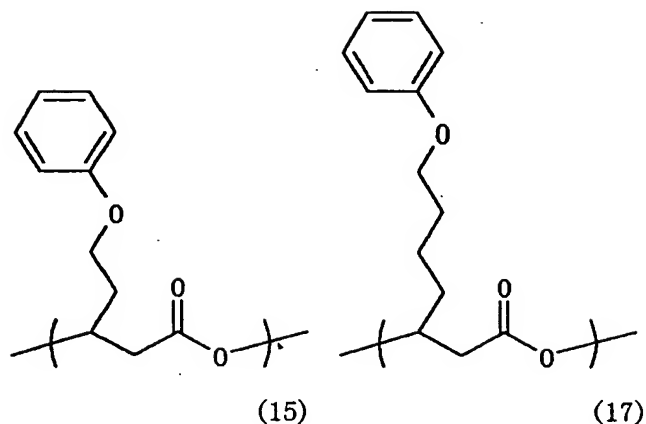
(wherein n is 0-10) (wherein k is 3 or 5)

Further, the polyhydroxyalkanoates are of formulas (8) and (16):

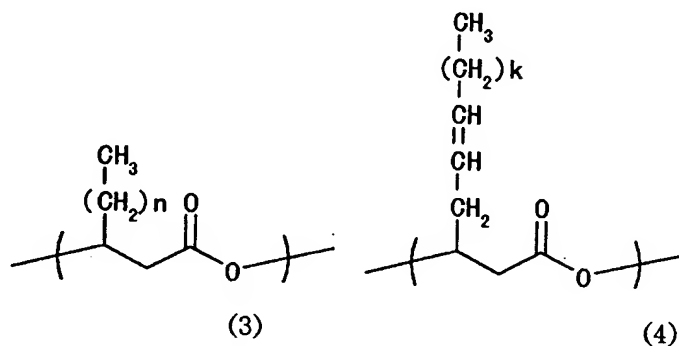


which contain 3-hydroxy-5-(4-fluorophenyl)valeric acid and 3-hydroxy-5-(4-fluorophenoxy)valeric acid as monomer units.

Further, the polyhydroxyalkanoates are of formulas (15) and (17):



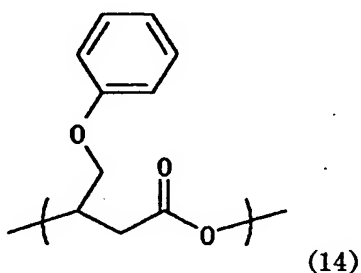
which contain 3-hydroxy-5-phenoxyvaleric acid and 3-hydroxy-7-phenoxyheptanoic acid as monomer units. Herein, as the monomer unit except 3-hydroxy-5-phenoxyvaleric acid and 3-hydroxy-7-phenoxyheptanoic acid of formulas (15) and (17), at least one or more of the monomer units of formula (3) or (4) are contained.



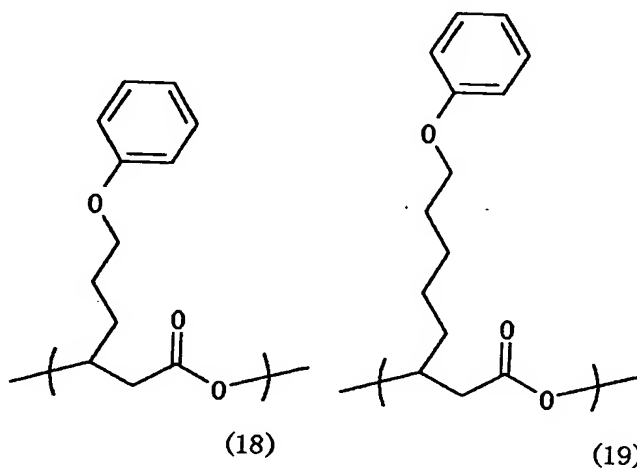
(wherein n is 0-10) (wherein k is 3 or 5)

Further, the polyhydroxyalkanoates are of formulas (14), (18) and (19):

5



10

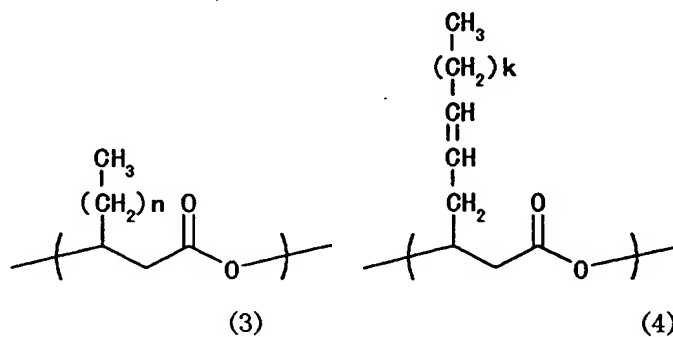


15

which contain 3-hydroxy-4-phenoxybutyric acid, 3-hydroxy-6-phenoxyhexanoic acid and 3-hydroxy-8-phenoxyoctanoic acid as monomer units. Herein, as the monomer unit except 3-hydroxy-4-phenoxybutyric acid, 3-hydroxy-6-phenoxyhexanoic acid and 3-hydroxy-8-phenoxyoctanoic acid of formulas (14), (18) and (19), at least one or more of the monomer units of formula (3) or (4) are contained.

20

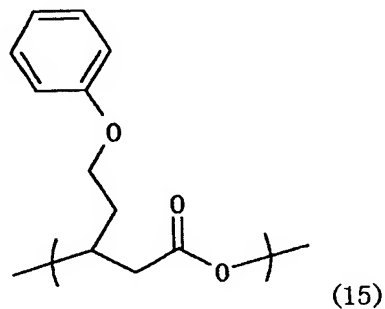
5



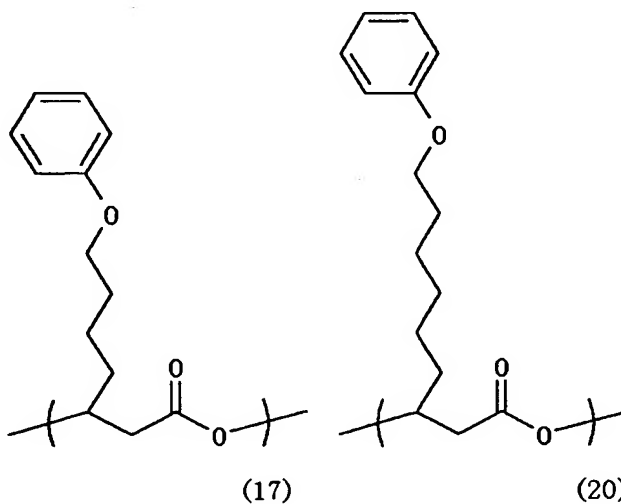
(wherein n is 0-10) (wherein k is 3 or 5)

Further, the polyhydroxyalkanoates are of formulas  
 10 (15), (17) and (20):

15



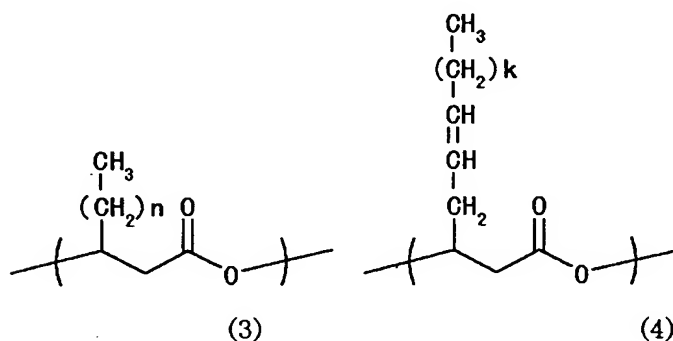
20



25

which contain 3-hydroxy-5-phenoxyvaleric acid, 3-

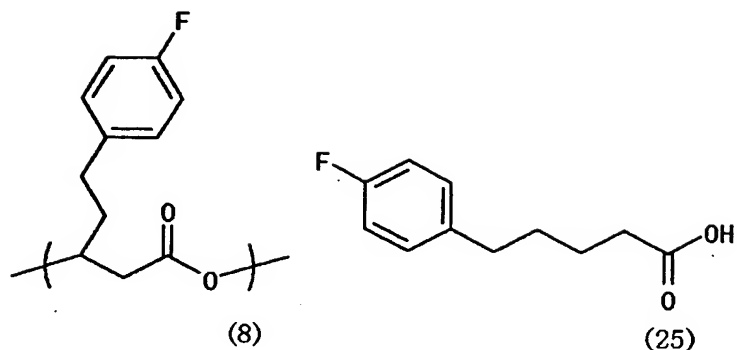
hydroxy-7-phenoxyheptanoic acid and 3-hydroxy-9-phenoxy-nonanoic acid as monomer units. Herein, as the monomer unit except 3-hydroxy-5-phenoxyvaleric acid, 3-hydroxy-7-phenoxyheptanoic acid and 3-hydroxy-9-phenoxy-nonanoic acid of formulas (15), (17) and (20), at least one or more of the monomer units of formula (3) or (4) are contained.



(wherein n is 0-10) (wherein k is 3 or 5)

Hereinafter, a manufacturing method for polyhydroxyalkanoates will be illustrated.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-5-(4-fluorophenyl)valeric acid (3HFPV) of formula (8) by being cultivated in a culture medium containing 5-(4-fluorophenyl)valeric acid (FPVA) of formula (25).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having a step of cultivating microorganisms which produce polyhydroxyalkanoates containing the monomer unit of 3HFPV of formula (8) using FPVA in a culture medium containing FPVA of formula (25).

In addition, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing FPVA of formula (25) and saccharides.

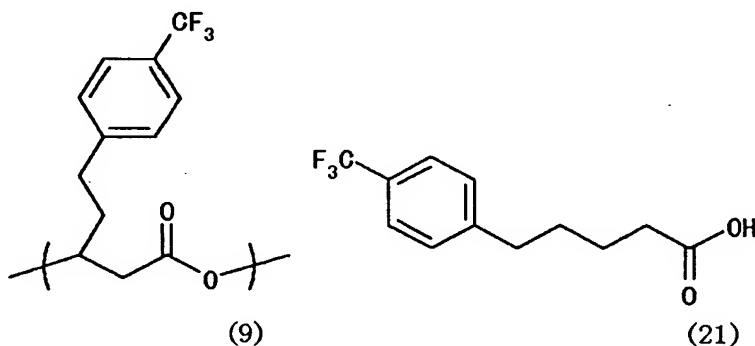
Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing FPVA of formula (25) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing FPVA of formula (25) and organic acids associated with the TCA cycle.



Furthermore, another manufacturing method is characterized in that cultivation of the microorganisms is performed by at least 2 step culturing: first in a culture medium containing FPVA of formula (25) and polypeptone, and then in a culture medium containing FPVA of formula (25) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid (3HCF<sub>3</sub>PV) of formula (9) by being cultured in a culture medium containing 5-(4-trifluoromethylphenyl)valeric acid (CF<sub>3</sub>PVA) of formula (21).



A manufacturing method for polyhydroxyalkanoates of the present invention comprise is characterized by culturing the microorganism which can produce polyhydroxyalkanoates containing the monomer unit of 3HCF<sub>3</sub>PV of formula (9) from CF<sub>3</sub>PVA of formula (21) in a culture medium containing CF<sub>3</sub>PVA.

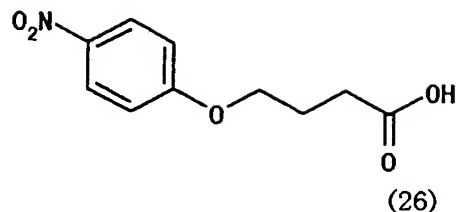
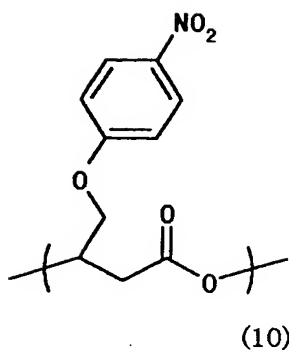
In addition, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing  $\text{CF}_3\text{PVA}$  of formula (21) and a saccharide.

5 Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing  $\text{CF}_3\text{PVA}$  of formula (21) and polypeptone.

Further, another manufacturing method is  
10 characterized in that cultivation of the microorganisms is performed in a culture medium containing  $\text{CF}_3\text{PVA}$  of formula (21) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is  
15 characterized in that cultivation of the microorganisms is performed by at least two step culturing: first in a culture medium containing  $\text{CF}_3\text{PVA}$  of formula (21) and polypeptone, and then in a culture medium containing  $\text{CF}_3\text{PVA}$  of formula (21) and pyruvic acid or its salt with  
20 nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-4-(4-nitrophenoxy)butyric acid ( $3\text{HNO}_2\text{PxB}$ ) formula (10) by  
25 being cultured in a culture medium containing 4-(4-nitrophenoxy)butyric acid ( $\text{NO}_2\text{PxBA}$ ) of formula (26).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by culturing a microorganism which can produce polyhydroxyalkanoates containing the monomer unit of 3HNO<sub>2</sub>PxB represented by formula (10) using NO<sub>2</sub>PxBA in a culture medium containing NO<sub>2</sub>PxBA of formula (26).

In addition, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing NO<sub>2</sub>PxBA of formula (26) and a saccharide.

Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing NO<sub>2</sub>PxBA of formula (26) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing NO<sub>2</sub>PxBA of formula (26) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is



is performed in a culture medium containing CNPxBA of formula (27) and a saccharide.

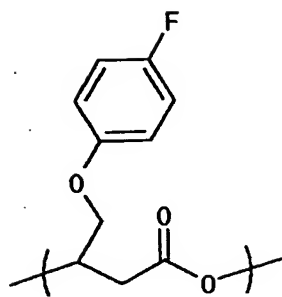
Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing CNPxBA of formula (27) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing CNPxBA of formula (27) and an organic acid associated with the TCA cycle.

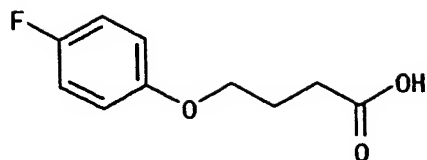
Furthermore, another manufacturing method is characterized in that cultivation of the microorganism is performed by at least 2 steps of culturing: first in a culture medium containing CNPxBA of formula (27) and polypeptone, and then in a culture medium containing CNPxBA of formula (27) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-4-(4-fluorophenoxy)butyric acid (3HFPxB) of formula (12) by being cultivated in a culture medium containing 4-(4-fluorophenoxy)butyric acid (FPxBA) of formula (28).

25



(12)



(28)

A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating a microorganism which can produce polyhydroxyalkanoates containing the monomer unit of 3HFPxB of formula (12) using FPxBA in a culture medium containing FPxBA of formula (28).

In addition, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing FPxBA of formula (28) and a saccharide.

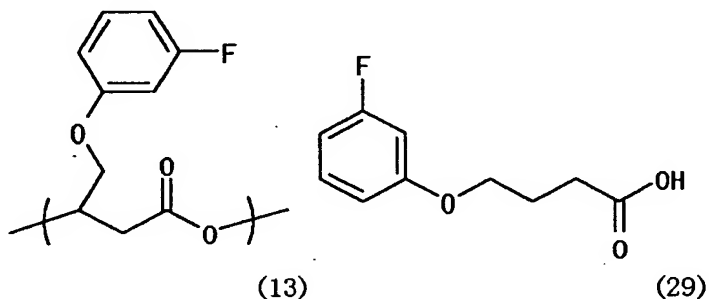
Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing FPxBA of formula (28) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing FPxBA of formula (28) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is

characterized in that cultivation of the microorganisms is performed by at least 2 steps of culturing: first in a culture medium containing FPxBA of formula (28) and polypeptone, and then in a culture medium containing FPxBA of formula (28) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-4-(3-fluorophenoxy)butyric acid (3HmFPxB) of formula (13) by being cultivated in a culture medium containing 4-(3-fluorophenoxy)butyric acid (mFPxBA) of formula (29).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having steps of cultivating microorganisms which produce polyhydroxyalkanoates containing the monomer unit of 3HmFPxB of formula (13) using mFPxBA in a culture medium containing mFPxBA of formula (29).

In addition, another manufacturing method is characterized in that cultivation of the microorganisms

is performed in a culture medium containing mFPxBA of formula (29) and saccharides.

Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing mFPxBA of formula (29) and polypeptone.

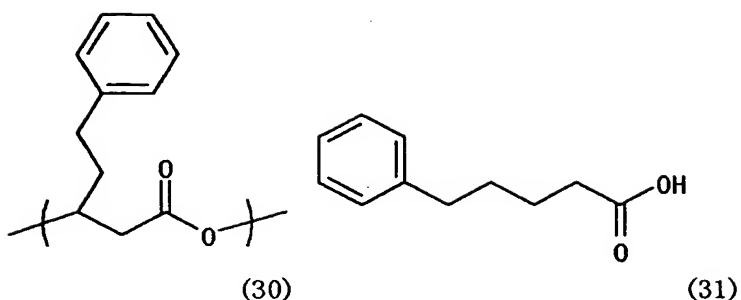
Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing mFPxBA of formula (29) and organic acids associated with the TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganisms is performed by at least 2 steps of culturing: first in a culture medium containing mFPxBA of formula (29) and polypeptone, and then in a culture medium containing mFPxBA of formula (29) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-5-phenylvaleric acid (3HPV) of formula (30) by being cultivated in a culture medium containing 5-phenylvaleric acid (PVA) of formula (31).

25





A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer unit of 3HPV of formula (30) using PVA in a culture medium containing PVA of formula (31) and a saccharide.

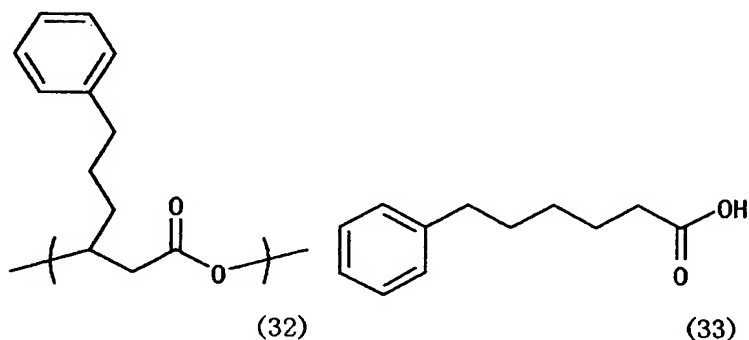
Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing PVA of formula (31) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganisms is performed in a culture medium containing PVA of formula (31) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganism is performed by at least 2 steps of culturing: first in a culture medium containing PVA of formula (31) and polypeptone, and then in a culture medium containing

PVA of formula (31) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-6-phenylhexanoic acid (3HPHx) of formula (32) by being cultivated in a culture medium containing 6-phenylhexanoic acid (PHxA) of formula (33).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer unit of 3HPHx of formula (32) using PHxA in a culture medium containing PHxA of formula (33) and a saccharide.

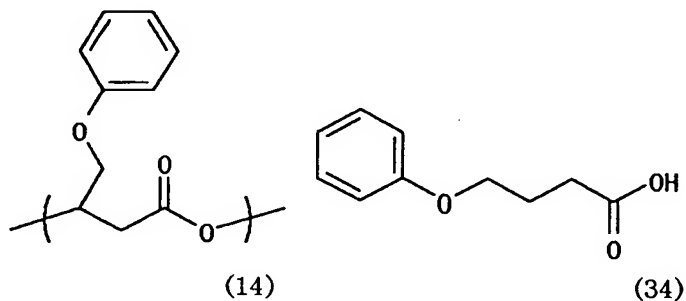
Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PHxA of formula (33) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganism

is performed in a culture medium containing PHxA of formula (33) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganism is performed by at least 2 steps of culturing: first in a culture medium containing PHxA of formula (33) and polypeptone, and then in a culture medium containing PHxA of formula (33) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-4-phenoxybutyric acid (3HPxB) of formula (14) by being cultivated in a culture medium containing 4-phenoxybutyric acid (PxBA) of formula (34).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer unit of 3HPxB of formula (14) using PxBA in a culture medium

containing PxBA of formula (34).

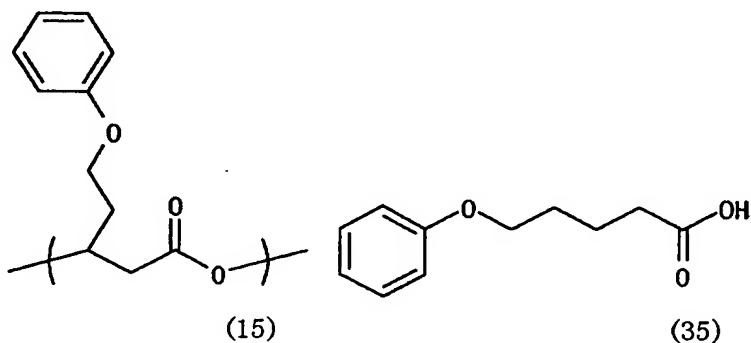
In addition, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxBA of  
5 formula (34) and a saccharide.

Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxBA of formula (34) and polypeptone.

10 Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxBA of formula (34) and an organic acid associated with the TCA cycle.

15 Furthermore, another manufacturing method is characterized in that cultivation of the microorganisms is performed by at least 2 steps of culturing: first in a culture medium containing PxBA of formula (34) and polypeptone, and then in a culture medium containing  
20 PxBA of formula (34) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-5-phenoxyvaleric  
25 acid (3HPxV) of formula (15) by being cultivated in a culture medium containing 5-phenoxyvaleric acid (PxVA) of formula (35).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer unit of 3HPxV of formula (15) using PxVA in a culture medium containing PxVA of formula (35).

In addition, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxVA of formula (35) and a saccharide.

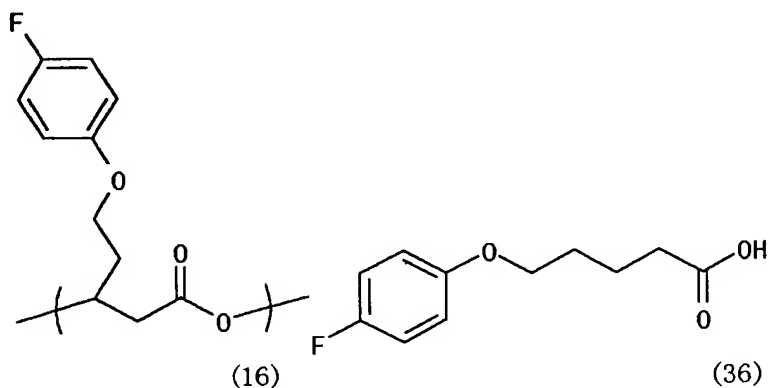
Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxVA of formula (35) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxVA of formula (35) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is

characterized in that cultivation of the microorganism is performed by at least 2 steps of culturing: first in a culture medium containing PxVA of formula (35) and polypeptone, and then in a culture medium containing PxVA of formula (35) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-5-(4-fluorophenoxy)valeric acid (3HFPxV) of formula (16) by being cultivated in a culture medium containing 5-(4-fluorophenoxy)valeric acid (FPxVA) of formula (36).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer unit of 3HFPxV of formula (16) using FPxVA in a culture medium containing FPxVA of formula (36) and a saccharide.

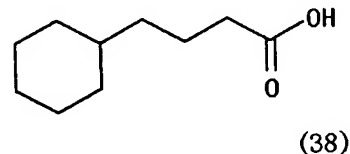
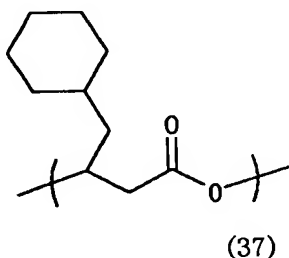
Further, another manufacturing method is

characterized in that cultivation of the microorganism is performed in a culture medium containing FPxVA of formula (36) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing FPxVA of formula (36) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganisms is performed by at least 2 steps of culturing, first in a culture medium containing FPxVA of formula (36) and polypeptone, and then in a culture medium containing FPxVA of formula (36) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing a monomer unit of 3-hydroxy-4-cyclohexylbutyric acid (3HCHB) of formula (37) by being cultivated in a culture medium containing 4-cyclohexylbutyric acid (CHBA) of formula (38).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer unit of  
5 3HCHB of formula (37) using CHBA in a culture medium containing CHBA of formula (38) and a saccharide.

Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing CHBA of  
10 formula (38) and polypeptone.

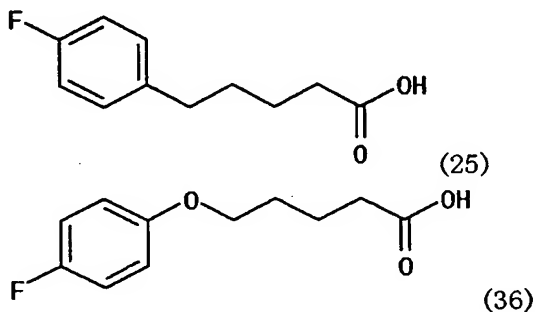
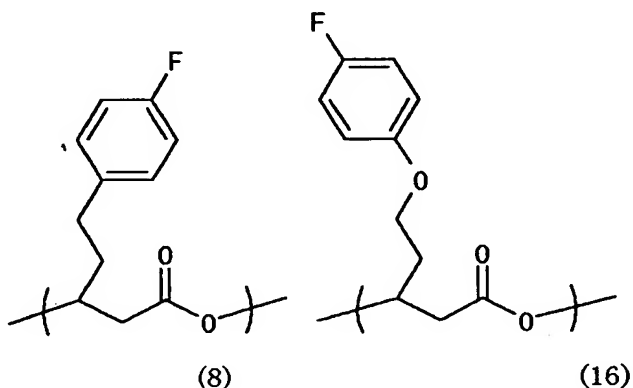
Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing CHBA of formula (38) and an organic acid associated with the  
15 TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganism is performed by at least 2 steps in a culture medium containing CHBA of formula (38) and polypeptone  
20 followed by in a culture medium containing CHBA of formula (38) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing monomer units of 3-hydroxy-5-(4-  
25 fluorophenyl)valeric acid (3HFPV) and 3-hydroxy-5-(4-fluorophenoxy)valeric acid (3HFPxV) of formulas (8) and



(16), respectively, by being cultivated in a culture medium containing 5-(4-fluorophenyl)valeric acid (FPVA) and 5-(4-fluorophenoxy)valeric acid (FPxVA) of formulas (25) and (36), respectively.



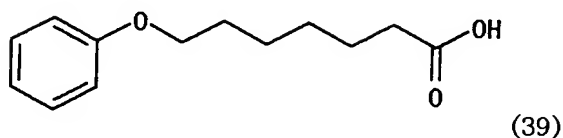
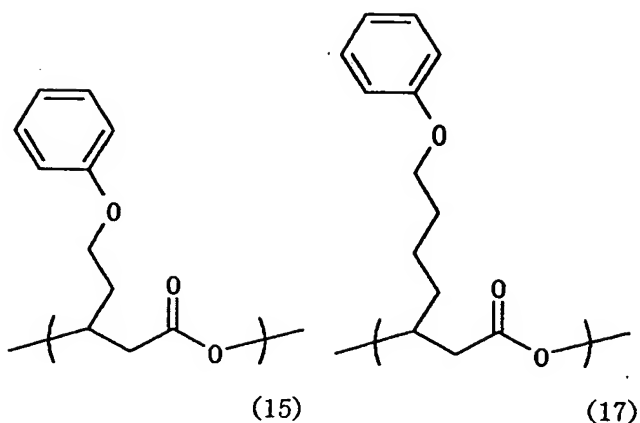
A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer units of 3HFPV and 3HFPxV of formulas (8) and (16), respectively, using FPVA and FPxVA in a culture medium containing FPVA and FPxVA of formulas (25) and (36), respectively, and a saccharide.

Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing FPVA and FPxVA of formulas (25) and (36), respectively, and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing FPVA and FPxVA of formulas (25) and (36), respectively, and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganisms is performed by at least 2 steps of culturing; first in a culture medium containing FPVA and FPxVA of formulas (25) and (36), respectively, and polypeptone, and then in a culture medium containing FPVA and FPxVA of formulas (25) and (36), respectively, and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing monomer units of 3-hydroxy-5-phenoxyvaleric acid (3HPxV) and 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) of formulas (15) and (17), respectively, by being cultivated in a culture medium containing 7-phenoxyheptanoic acid (PxHpA) of formulas (39).



A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer units of 3HPxV and 3HPxHp of formulas (15) and (17), respectively, using PxHpA in a culture medium containing PxHpA of formula (39) and a saccharide.

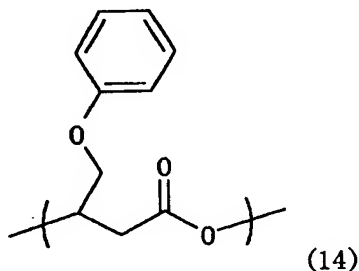
In addition, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxHpA of formula (39) and polypeptone.

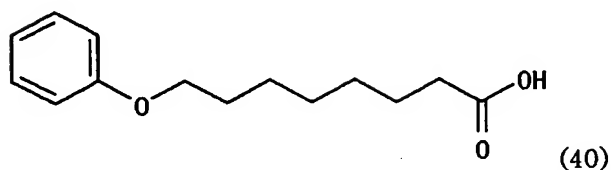
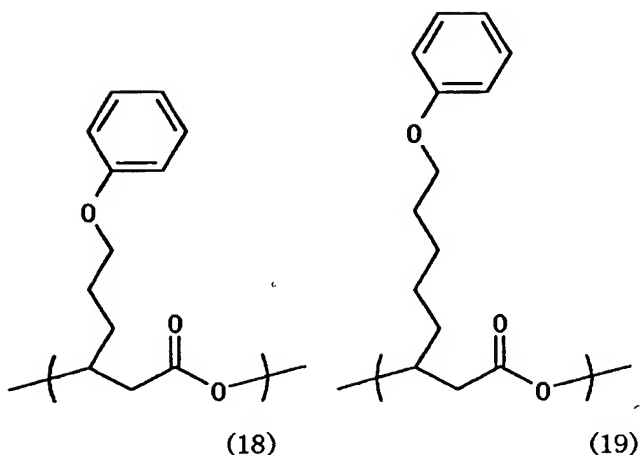
Further, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxHpA of

formula (39) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganisms is performed by at least 2 steps of culturing; first in a culture medium containing PxHpA of formula (39) and polypeptone, and then in a culture medium containing PxHpA of formula (39) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing monomer units of 3-hydroxy-4-phenoxybutyric acid (3HPxB), 3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) and 3-hydroxy-8-phenoxyoctanoic acid (3HPxO) of formulas (14), (18) and (19), respectively, by being cultivated in a culture medium containing 8-phenoxyoctanoic acid (PxOA) of formula (40).





A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer units of 3HPxB, 3HPxHx and 3HPxO of formulas (14), (18) and (19), respectively, using PxOA in a culture medium containing PxOA of formula (40) and a saccharide.

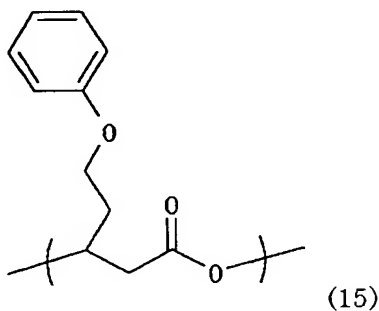
In addition, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxOA of formula (40) and polypeptone.

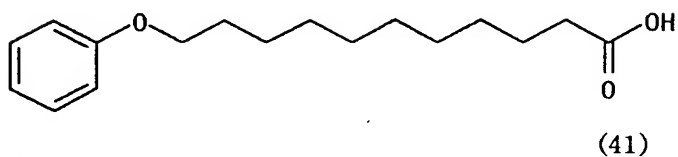
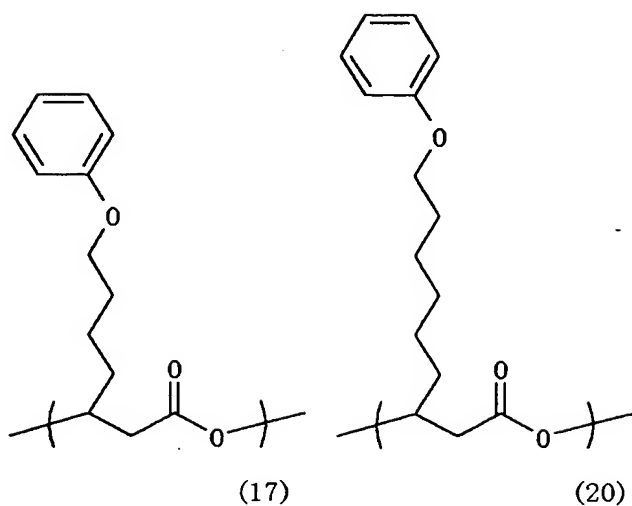
Further, another manufacturing method is characterized in that cultivation of the microorganism is cultivated in a culture medium containing PxOA of

formula (40) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganisms is performed by at least 2 steps of culturing: first in a culture medium containing PxOA of formula (40) and polypeptone, and then in a culture medium containing PxOA of formula (40) and pyruvic acid or its salt with nitrogen limitation.

The present inventors have succeeded in obtaining microorganisms which can produce polyhydroxyalkanoates containing monomer units of 3-hydroxy-5-phenoxyvaleric acid (3HPxV); 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) and 3-hydroxy-9-phenoxyundecanoic acid (3HPxN) of formulas (15), (17) and (20), respectively, by being cultivated in a culture medium containing 11-phenoxyundecanoic acid (PxUDA) of formula (41).





A manufacturing method for polyhydroxyalkanoates of the present invention is characterized by having the step of cultivating microorganisms which can produce polyhydroxyalkanoates containing the monomer units of 3HPxV, 3HPxHp and 3HPxN of formulas (15), (17) and (20), respectively, using PxUDA in a culture medium containing PxUDA of formula (41) and a saccharide.

In addition, another manufacturing method is characterized in that cultivation of the microorganism is performed in a culture medium containing PxUDA of formula (41) and polypeptone.

Further, another manufacturing method is characterized in that cultivation of the microorganism

is performed in a culture medium containing PxUDA of formula (41) and an organic acid associated with the TCA cycle.

Furthermore, another manufacturing method is characterized in that cultivation of the microorganisms is performed by at least 2 steps of culturing; first in a culture medium containing PxUDA of formula (41) and polypeptone, and then in a culture medium containing PxUDA of formula (41) and pyruvic acid or its salt with nitrogen limitation.

Further, four new strains suitably usable for production of the above described polyhydroxyalkanoates of the present invention include *Pseudomonas cichorii* YN2, FERM BP-7375, *Pseudomonas cichorii* H45, FERM BP-7374, *Pseudomonas putida* P91, FERM BP-7373 and *Pseudomonas jessenii* P161, FERM BP-7376.

The PHAs according to the present invention are those PHAs which contain monomer units with a variety of structures having substituents useful as device and medical materials and others on the side chain, and more specifically, those which have the above-mentioned substituted or non-substituted phenoxy, phenyl and cyclohexyl groups on the side chain. In addition, the method according to the present invention enables the production of desired PHAs at a high purity and a high yield by using the microorganisms. Furthermore, the present invention can provide strains capable of



efficient synthesis of the PHAs at a high purity. In general, the PHAs according to the present invention are of only the R form, and are isotactic polymers.

<Saccharides and Organic Acids Involved in the TCA  
5 Cycle: Differences from Conventional Technology>

One method for producing PHA according to the present invention is characterized in that when a microorganism is cultured, in addition to alkanoate for introducing the desired monomer unit, only  
10 saccharide(s) or organic acid(s) involved in the TCA cycle as the carbon source other than the alkanoate are added into the medium, so that the PHA produced by and accumulated in the microorganism has a significantly high content of the monomer unit of interest or  
15 alternatively only the monomer unit of interest. The effect of facilitating the preference of this particular monomer unit is achieved by adding into the medium only saccharide(s) or organic acid(s) involved in the TCA cycle as the carbon source other than the  
20 alkanoate.

That is, the inventors have found that when culture is carried out using saccharide(s) or organic acid(s) involved in the TCA cycle as the co-existing substrate and together with alkanoate for introducing  
25 the desired monomer unit, the PHA of interest is produced at a particularly superior yield and purity, compared to conventional methods employing mcl-

alkanoate, such as nonanoate or octanoate, as the co-existing substrate, and that this effect is achieved by culture methods allowing acetyl-CoA that is a carbon source as well as an energy source of the microorganism to be produced by processes independent on the  $\beta$ -oxidation, and have reached the present invention.

In the methods according to the invention, saccharide compounds, for example, glucose, fructose, mannose, and the like, are utilized as growth substrates for microorganisms, and the produced PHA is of alkanoate for introducing the desired monomer unit which is existing with saccharide(s), so that there is contained none of or an extremely small amount of monomer units derived from the saccharides such as glucose and others. In this point, the methods according to the present invention is essentially different in structures and effects from conventional methods for microbial production of PHA in which saccharides themselves, such as glucose and others, are employed as the raw substrate of the monomer unit to be introduced into the PHA.

#### <Polypeptone: Differences from Conventional Technology>

One method for producing PHA according to the present invention is characterized in that when a microorganism is cultured, in addition to alkanoate, a raw material, for introducing the desired monomer unit, only polypeptone as the carbon source other than the

alkanoate is added into the medium, so that the PHA produced by and accumulated in the microorganism has a significantly high content of the monomer unit of interest or alternatively only the monomer unit of interest. The effect of facilitating the preference of this particular monomer unit is achieved by adding into the medium only polypeptone as the carbon source other than the alkanoate.

As examples of utilizing polypeptone in microbial production of PHA, Japanese Patent Application Laid-Open Nos. 5-49487, 5-64591, 5-214081, 6-145311, 6-284892, 7-48438, 8-89264, 9-191893, and 11-32789 disclose that when PHA is produced by microorganisms, polypeptone is allowed to be contained in the media. However, all of these utilize polypeptone during pre-culture, that is, at the stage of simply growing cells, and there are not contained substrates resulting in the monomer unit of PHA during pre-culture. Furthermore, there are no examples utilizing polypeptone at the stage of allowing cells to produce PHA. In contrast, the present invention is intended to produce and accumulate PHA, as well as to grow cells, with alkanoate for introducing the desired monomer unit and by co-existing only polypeptone as the carbon sources other than the alkanoate. The production method according to the present invention employing polypeptone, therefore, has quite different structures

and effects from conventional examples employing polypeptone. Moreover, there is no mention of the preference of the particular monomer units which is the effect of the present invention, and no indication of the effect of the preference of the particular monomer units having as substituents phenoxy, phenyl and cyclohexyl groups in the composition of PHAs produced by the microorganisms, as in the present invention.

The PHAs, production methods, and microorganisms of the present invention will be explained in more detail below.

#### <Supplying Pathways of PHA Monomer Units>

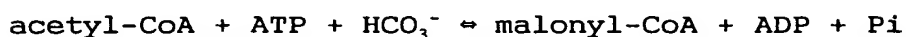
At first will be detailed the "fatty acid synthesis pathway," one of pathways supplying mcl-3HA monomer units to be mixed into the PHA of interest.

In the case where saccharides such as glucose and the like are substrates, alkanoates necessary for cellular components are biosynthesized from the "fatty acid synthesis pathway" in which acetyl-CoA produced from saccharides through the "glycolytic pathway" is a starting substance. The fatty acid synthesis involves the de novo synthesis pathway and the carbon-chain elongation pathway, as explained below.

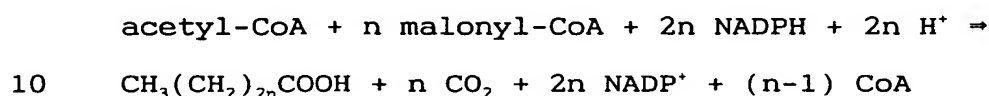
#### 1) De novo Synthesis Pathway

This pathway is catalyzed by two enzymes, acetyl-CoA carboxylase (EC 6.4.1.2) and fatty acid synthase (E.C. 2.3.1.85). Acetyl-CoA carboxylase is an enzyme

interposing biotin, and ultimately catalyzing the following reaction to produce malonyl-CoA from acetyl-CoA. This reaction is of follows:

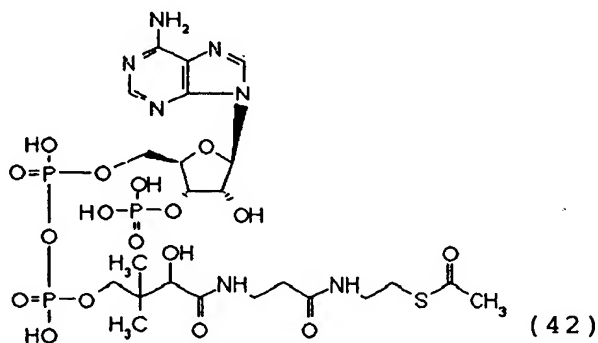


5 Also, fatty acid synthase is an enzyme catalyzing a cycles of reactions of transfer - decarbonation - reduction - dehydration - reduction. The entire reactions are represented as follows:

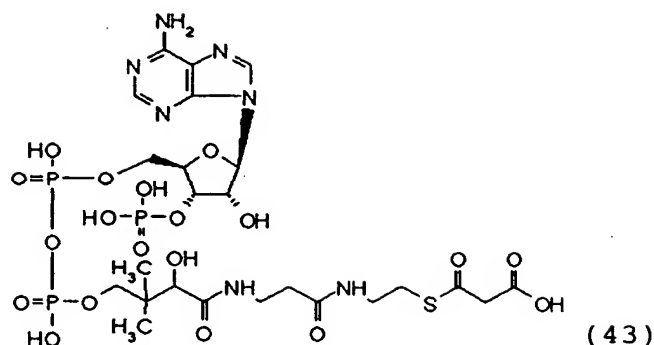


Reaction products may be free acids, CoA-derivatives, or ACP-derivatives, depending on the type of enzymes.

Now, acetyl-CoA and malonyl-CoA are represented by  
15 the following chemical formulae (42) and (43), respectively.

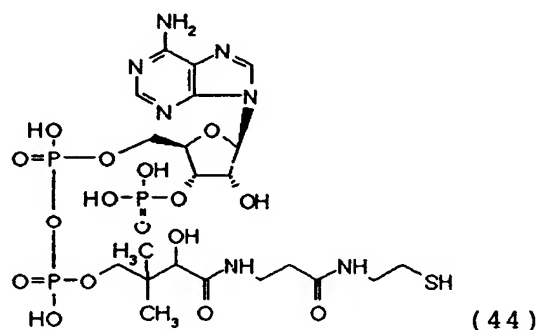


5



In addition, Co-A stands for co-enzyme A, and is  
 10 represented by the following chemical formula (44).

15



Within this reaction pathway, the route described  
 20 below gives "D-3-hydroxyacyl-ACP," an intermediate to  
 be the monomer substrate for the PHA biosynthesis.  
 Additionally, as shown in the following reaction  
 formulae, this route extends finally to palmitate with  
 repeated addition of two carbons. Therefore, as the  
 25 monomer substrate for the PHA biosynthesis are provided  
 seven "D-3-hydroxyacyl-ACPs" having even numbers of the  
 carbons, from "D-3-hydroxybutyryl-ACP" to "D-3-

μm), solvent; (chloroform, polystyrene reduced) to obtain Mn=58200 and Mw=108100.

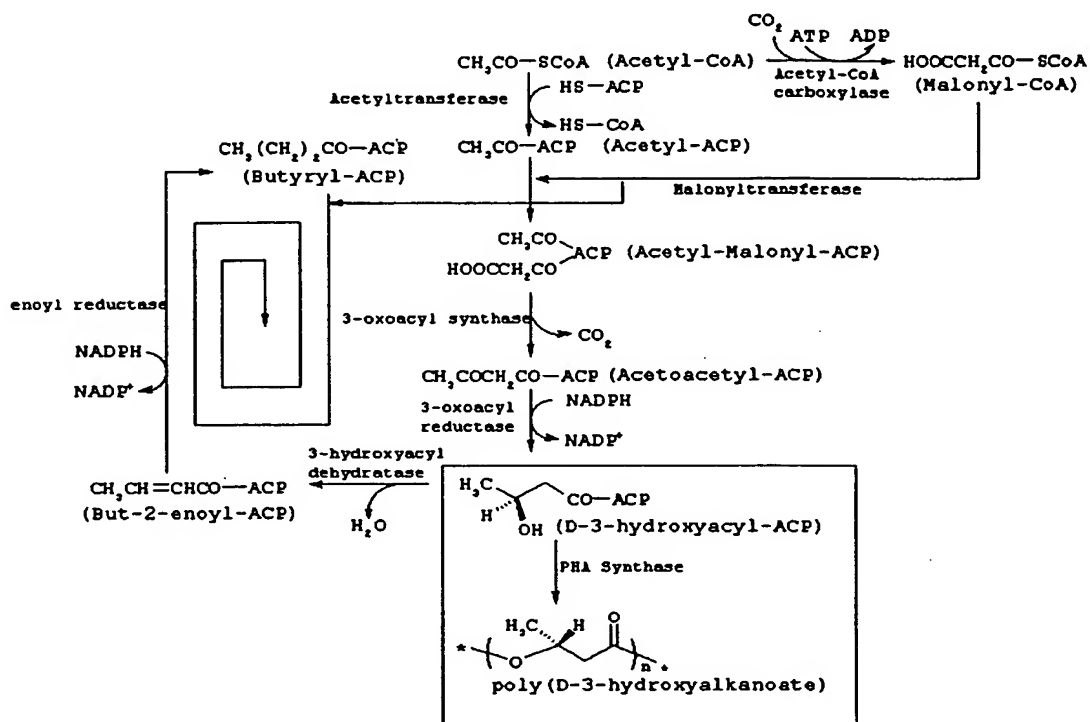
[Example 35]

First, cells of *Pseudomonas cichorii* strain H45  
5 were inoculated into 10 mL of M9 culture medium  
containing 0.5% D-glucose and shake-cultured at 30°C at  
125 stroke/min, and then 2 mL of the culture were added  
into 200 mL of M9 culture medium containing 0.5%  
D-glucose and 0.1% CNPxBA, and shake cultured at 30°C  
10 at 125 stroke/min. 48 hours later, the cells were  
recovered by centrifugation and re-suspended into 200  
mL of M9 culture medium containing 0.5% D-glucose, 0.1%  
CNPxBA and nitrogen (NH<sub>4</sub>Cl)-free, and further shake  
cultured at 30°C at 125 stroke/min. 47 hours later,  
15 the cells were recovered by centrifugation, washed once  
with cold methanol and lyophilized.

This lyophilized pellet was suspended into 20 mL  
of chloroform, and stirred at 60°C for 20 hours to  
extract PHA. After the extracts were filtrated through  
20 a membrane filter with a pore size of 0.45 μm, they  
were concentrated by rotating evaporator, and the  
concentrates were re-precipitated with cold methanol,  
and further the precipitates alone were recovered and  
vacuum dried to provide PHA.

25 The resulting PHA was determined by NMR analysis  
under the condition shown in the Example 34. As a  
result, it was confirmed that the PHA was the PHA

hydroxypalmityl-ACP."

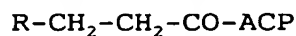
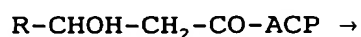
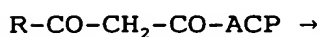


## 2) Carbon-Chain Elongation Pathway

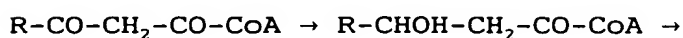
This pathway is broadly divided into two pathways: a pathway in which malonyl-CoA is added to acyl-ACP, which is finally converted to acyl-ACP having the carbon chain extended with two carbons (and  $\text{CO}_2$ ) (referred to as Pathway A) and a pathway in which acetyl-CoA is added to acyl-CoA, which is finally converted to acyl-CoA having the carbon chain extended with two carbons (referred to as Pathway B). Each pathway will be explained below.

- Pathway A





- Pathway B



In either Pathway A or B, it is thought that "D-3-hydroxyacyl-CoA" or "D-3-hydroxyacyl-ACP" is yielded as an intermediate, and "D-3-hydroxyacyl-CoA" is utilized as the monomer substrate for the PHA synthesis as it is, while "D-3-hydroxyacyl-ACP" is utilized as the monomer substrate for the PHA synthesis after converting to "D-3-hydroxyacyl-CoA" by ACP-CoA transferase.

In the case where saccharides such as glucose and the like are used as a substrate, it is thought that an mcl-3HA monomer unit is generated within microbial cells via the "glycolytic pathway" and the "fatty acid synthesis pathway" as described above. In the case where organic acids involved in the TCA cycle are used as a substrate, acetyl-CoA is yielded directly from pyruvic acid by pyruvate dehydrogenase. Organic acids in the TCA cycle, for example, malic acid yields

pyruvic acid by malate dehydrogenase, followed by acetyl-CoA by the above-mentioned reaction.

Oxaloacetic acid yields phosphoenolpyruvic acid by phosphoenolpyruvate carboxykinase, which, in turn, is catalyzed to produce pyruvic acid by pyruvate kinase, followed by acetyl-CoA by the above-mentioned reaction. It is thought that acetyl-CoA produced by these reactions goes through the "fatty acid synthesis pathway" to produce an mcl-3HA monomer unit.

10           In these cases, it is thought that mcl-alkanoates, for example, octanoate, nonanoate, and the like, or alkanoates having functional groups other than straight aliphatic alkyl added at the end, for example, such as 5-phenylvalerate, 5-(4-fluorophenyl)valerate, 6-  
15   phenylhexanoate, 4-phenoxybutyrate, and 4-cyclohexylbutyrate are converted to their CoA-derivatives by CoA-ligases (EC 6.2.1.3, etc), followed by "D-3-hydroxyacyl-CoA" to be the monomer substrate for the PHA biosynthesis directly by a series of  
20   enzymes responsible for the  $\beta$ -oxidation pathway.

          In short, the mcl-3HA monomer units generated from saccharides or organic acids involved in the TCA cycle is produced via quite a lot of enzymatic reaction steps (i.e., indirectly), but the mcl-alkanoates should yield  
25   the mcl-3HA monomer units quite directly.

          There will now be described the generation of acetyl-CoA responsible for microbial growth. In the

method in which mcl-alkanoate is co-existed in addition to alkanoate for introducing the monomer unit of interest, these alkanoates go through the  $\beta$ -oxidation pathway to produce acetyl-CoA. Generally, mcl-  
5 alkanoates are believed to have a superior substrate-affinity for a series of enzymes in the  $\beta$ -oxidation pathway, as compared with alkanoates having a bulky substituent (alkanoates having substituents such as phenyl, phenoxy, cyclohexyl group, or the like), and  
10 thus acetyl-CoA is effectively produced by co-existence with mcl-alkanoates. For this reason, it is advantageous to microbial growth utilizing acetyl-CoA as the energy and carbon source.

However, since mcl-alkanoates pathway are directly  
15 converted to the monomer unit for PHA via the  $\beta$ -oxidation, it is a significant problem that the produced PHAs also contains the mcl-3HA monomer unit a lot, in addition to the monomer unit of interest.

To solve this problem, methods are desirable in  
20 which rather than mcl-alkanoate, such substrates that can provide acetyl-CoA or the energy and carbon source effectively are selected and allowed to co-exist with the alkanoate of interest. As mentioned above, acetyl-CoA can be the monomer unit of PHA by going into the  
25 fatty acid synthesis pathway, but this process is an indirect process of many steps compared with mcl-alkanoate in  $\beta$ -oxidation. Therefore, it is possible to

achieve a production method not to incorporate or decrease mcl-3HA in PHA by selecting culture conditions such as the substrate concentration to generate acetyl-CoA.

5           Alternatively, there are commonly used production methods by which culture is carried out at the first step only for the purpose of microbial growth and at the second step is added to the medium only the alkanoate of interest as the carbon source. In this  
10       case, ATP is required by acyl-CoA ligase which is an initial enzyme of the beta-oxidation pathway converting the alkanoate to acyl-CoA. Consequently, the inventors' investigation has provided the result that the production methods are more effective by which  
15       substrates capable of being utilized as the energy source by microorganisms are also co-existed at the second step, and accomplished the present invention.

          As substrates which can effectively provide acyl-CoA or the energy and carbon source in method according  
20       to the present invention, as long as compounds can yield acyl-CoA or the energy and carbon source without going through the beta-oxidation pathway, for example, aldoses including glyceraldehyde, erythrose, arabinose, xylose, glucose, galactose, mannose, and fructose,  
25       alditols such as glycerol, erythritol, and xylitol, aldonic acids such as gluconic acid, uronic acids such as glucuronic acid and galacturonic acid, saccharides

such as disaccharides including maltose, sucrose, and lactose, and in addition, organic acids involved in the TCA cycle such as lactic, pyruvic, malic, citric, succinic, fumaric acids and their salts, and further, medium components derived from natural products such as polypeptone, beef extract, and casamino acid, and the like, any compounds can be used, or selected as appropriate, based on usefulness as a substrate for the strains employed. Furthermore, if their combinations result in a small degree of mixture with mcl-3HA, two or more compounds also can be selected for use.

<Microorganisms>

As mentioned in Background of the Invention, there are reports of microorganisms that produce and accumulate within the cell PHA containing the monomer unit of, for example, 3-hydroxy-4-phenoxybutyrate, 3-hydroxy-5-fluorophenoxyvalerate, 3-hydroxy-6-cyanophenoxyhexanoate, 3-hydroxy-6-nitrophenoxyhexanoate, 3-hydroxy-7-fluorophenoxyheptanoate, such as *Pseudomonas oleovorance* and *Pseudomonas putida* as described in *Macromolecules*, 29, 3432-3435 (1996), *Can. J. Microbiol.*, 41, 32-43 (1995), Japanese Patent No. 2989175, and others. However, there are no report of microorganisms that produce and accumulate within the cell PHA containing the monomer unit of 3-hydroxy-4-phenoxybutyrates having substituents such as fluorine,

cyano, nitro groups. Macromolecules, 32, 2889-2895  
(1999) reported that *Pseudomonas oleovorance* produces  
PHA containing the monomer units of 3-hydroxy-5-(2,4-  
dinitrophenyl)valerate and 3-hydroxy-5-(4-  
5 nitrophenyl)valerate by culturing it in a medium  
containing 5-(2,4-dinitrophenyl)valerate and nonanoate  
as substrates. However, there are no reports on  
microorganisms that produce and accumulate within the  
cell PHA containing as the monomer unit 3-hydroxy-  
10 phenylalkanoates having a substituent such as fluorine,  
trifluoromethyl group. Therefore, the present  
invention has been achieved by screening microorganisms  
capable of incorporating these new monomer units into  
PHA.

15        Novel microorganisms of the present invention have  
a previously unknown capability of producing and  
accumulating within the cell PHA containing a new  
monomer unit derived from an alkanoate using the  
alkanoate as a substrate. Microorganisms displaying  
20 such a novel enzymatic reaction have been found by the  
inventors by screening. The novel microorganisms of  
the present invention are *Pseudomonas cichorii* strain  
YN2 (FERM BP-7375), *Pseudomonas cichorii* strain H45  
(FERM BP-7374), *Pseudomonas putida* strain P91 (FERM BP-  
25 7373), and *Pseudomonas jessenii* strain P161 (FERM BP-  
7376). Other than these microorganisms, microorganisms  
to be utilized in the production method of PHA

according to the present invention can be obtained by culturing a bacterial strain, for example, of genus *Pseudomonas*, employing the alkanooates as the substrate, for example.

- 5           There will be given details concerning strains YN2, H45, P91, and P161.

<Bacteriological Properties of Strain YN2>

(1) Morphological Properties

- 10   Shape and size of cells: rod, 0.8  $\mu$ m  $\times$  1.5 to 2.0  $\mu$ m  
Polymorphism of cells: negative  
Mobility: motile  
Sporulation: negative  
Gram staining: negative  
15   Colony shape: circular; entire, smooth margin; low convex; smooth surface; glossy; translucent

(2) Physiological Properties

- Catalase: positive  
20   Oxidase: positive  
O/F test: oxidative (non-fermentative)  
Nitrate reduction: negative  
Indole production: positive  
Acid production from glucose: negative  
25   Arginine dihydrolase: negative  
Urease: negative  
Esculin hydrolysis: negative

Gelatin hydrolysis: negative

$\beta$ -Galactosidase: negative

Fluorescent pigment production on King's B agar:  
positive

5 Growth under 4% NaCl: positive (weak growth)

Poly- $\beta$ -hydroxybutyrate accumulation: negative (\*)

Tween 80 hydrolysis: positive

(\*) Colonies cultured on nutrient agar were stained  
with Sudan Black for determination.

10

(3) Substrate Assimilation

Glucose: positive

L-Arabinose: positive

D-Mannose: negative

15 D-Mannitol: negative

N-Acetyl-D-glucosamine: negative

Maltose: negative

Potassium gluconate: positive

n-Caprate: positive

20 Adipate: negative

dl-Malate: positive

Sodium citrate: positive

Phenyl acetate: positive

25 <Bacteriological Properties of Strain H45>

(1) Morphological Properties

Shape and size of cells: rod, 0.8  $\mu$ m  $\times$  1.0 to 1.2  $\mu$ m



Polymorphism of cells: negative

Mobility: motile

Sporulation: negative

Gram staining: negative

- 5 Colony shape: circular; entire, smooth margin; low  
convex; smooth surface; glossy; cream-colored

(2) Physiological Properties

Catalase: positive

- 10 Oxidase: positive

O/F test: oxidative

Nitrate reduction: negative

Indole production: negative

Acid production from glucose: negative

- 15 Arginine dihydrolase: negative

Urease: negative

Esculin hydrolysis: negative

Gelatin hydrolysis: negative

$\beta$ -Galactosidase: negative

- 20 Fluorescent pigment production on the King's B agar:  
positive

Growth under 4% NaCl: negative

Poly- $\beta$ -hydroxybutyrate accumulation: negative

- 25 (3) Ability to assimilate substrates

Glucose: positive

L-Arabinose: negative

- D-Mannose: positive  
D-Mannitol: positive  
N-Acetyl-D-glucosamine: positive  
Maltose: negative
- 5 Potassium gluconate: positive  
n-Caprato: positive  
Adipate: negative  
dl-Malate: positive  
Sodium citrate: positive
- 10 Phenyl acetate: positive
- <Bacteriological Properties of Strain P91>
- (1) Morphological Properties
- Shape and size of cells: rod, 0.6  $\mu\text{m}$   $\times$  1.5  $\mu\text{m}$
- 15 Polymorphism of cells: negative  
Mobility: motile  
Sporulation: negative  
Gram staining: negative  
Colony shape: circle; entire, smooth margin; low
- 20 convex; smooth surface; glossy; cream-colored
- (2) Physiological Properties
- Catalase: positive  
Oxidase: positive
- 25 O/F test: oxidative  
Nitrate reduction: negative  
Indole production: negative

Acid production from glucose: negative

Arginine dihydrolase: positive

Urease: negative

Esculin hydrolysis: negative

5 Gelatin hydrolysis: negative

$\beta$ -Galactosidase: negative

Fluorescent pigment production on the King's B agar:  
positive

10 (3) Substrate assimilation

Glucose: positive

L-Arabinose: negative

D-Mannose: negative

D-Mannitol: negative

15 N-Acetyl-D-glucosamine: negative

Maltose: negative

Potassium gluconate: positive

n-Caprato: positive

Adipate: negative

20 dl-Malate: positive

Sodium citrate: positive

Phenyl acetate: positive

<Bacteriological Properties of the Strain P161>

25 (1) Morphological Properties

Shape and size of cells: spheres,  $\phi 0.6 \mu\text{m}$

rods,  $0.6 \mu\text{m} \times 1.5 \text{ to } 2.0 \mu\text{m}$

Polymorphism of cells: elongated form

Mobility: motile

Sporulation: negative

Gram staining: negative

- 5 Colony shape: circle; entire, smooth margin; low convex; smooth surface; pale yellow

## (2) Physiological Properties

Catalase: positive

- 10 Oxidase: positive

O/F test: oxidative

Nitrate reduction: positive

Indole production: negative

Acid production from glucose: negative

- 15 Arginine dihydrolase: positive

Urease: negative

Esculin hydrolysis: negative

Gelatin hydrolysis: negative

$\beta$ -Galactosidase: negative

- 20 Fluorescent pigment production on the King's B agar: positive

## (3) Substrate assimilation

Glucose: positive

- 25 L-Arabinose: positive

D-Mannose: positive

D-Mannitol: positive

N-Acetyl-D-glucosamine: positive

Maltose: negative

Potassium gluconate: positive

n-Caprato: positive

5 Adipate: negative

dl-Malate: positive

Sodium citrate: positive

Phenyl acetate: positive

10           Based on these bacteriological properties, and  
referring to Bergey's Manual of Systematic  
Bacteriology, vol. 1 (1984) and Bergey's Manual of  
Determinative Bacteriology, 9th ed. (1994), strains YN2  
and H45 were revealed to belong to *Pseudomonas*  
15 *cichorii*, and strain P91 was revealed to belong to  
*Pseudomonas putida*. Accordingly, these strains were  
designated *Pseudomonas cichorii* strain YN2, *Pseudomonas*  
*cichorii* strain H45, and *Pseudomonas putida* strain P91.

          Strain P161, on the other hand, was revealed to  
20 belong to the genus *Pseudomonas* (*Pseudomonas* sp.), but  
its bacteriological properties could not identify its  
taxonomic position. Then, to do classification based  
on genetic properties, the DNA sequence of 16S rRNA  
coding region of strain P161 (SEQ ID NO: 1) has been  
25 determined to examine the homology with the DNA  
sequences of 16S RNA coding region of known  
microorganisms of genus *Pseudomonas*. The results have

shown that there is a remarkably high homology of the DNA sequences between strain P161 and *Pseudomonas jessenii*. Furthermore, bacteriological properties of *Pseudomonas jessenii* described in System. Appl. Microbiol., 20, 137-149 (1997) and System. Appl. Microbiol., 22, 45-58 (1999) were found to have a high similarity to those of strain P161. From these results, strain P161 was designated *Pseudomonas jessenii* strain P161, since it was concluded that it is appropriate that strain P161 should be attributed to belong to *Pseudomonas jessenii*.

Strains YN2, H45, P91, and P161 have been deposited at National Institute of Bioscience and Human-Technology (Patent Microorganism Depository Center), Agency of Industry Science and Technology, Ministry of International Trade and Industry, under the deposition numbers "FERM BP-7375," "FERM BP-7374," "FERM BP-7373," and "FERM BP-7376," respectively.

<Culture: General>

The PHAs of interest can be produced by culturing these microorganisms in a medium containing alkanoate for introducing the desired monomer unit and growth substrates according to the present invention. These PHAs are generally of only the R-form, and are isotactic polymers.

For usual culture of microorganisms to be employed in the production method of PHA according to the

present invention, for example, for preparation of cell stocks, for maintaining of the number and activities of the cells, any type of media can be used, such as common natural media and synthetic media supplemented  
5 with nutrients, unless they have adverse effects on the growth or existence of the microorganisms. Culture conditions such as temperature, aeration, stirring, and the like are selected as appropriate, depending on the microorganism employed.

10 In the case where microorganisms are used to produce and accumulate PHA, inorganic media and others containing alkanoate for introducing the desired monomer unit can be employed as a medium for the PHA production.

15 For inorganic media employed in the above-mentioned culture method, any media can be used, as long as they contain components allowing microorganisms to grow, such as phosphorus sources (for example, phosphates), nitrogen sources (for example, ammonium  
20 salts, nitrates), and the like. Such inorganic media may include, for example, MSB medium, E medium (J. Biol. Chem. 218, 97-106 (1956)), M9 medium, and others.

The composition of M9 medium employed in Examples of the present invention is as follows:

25	$\text{Na}_2\text{HPO}_4$ :	6.2 g
	$\text{KH}_2\text{PO}_4$ :	3.0 g
	$\text{NaCl}$ :	0.5 g

NH<sub>4</sub>Cl: 1.0 g  
(per liter of medium, pH 7.0)

Culture conditions may include, for example,  
shaking culture and stirring culture under aerobic  
5 conditions at 15 to 40°C, and preferably 20 to 35°C.

The culture steps can utilize any processes  
employed for usual culture of microorganisms, such as  
batch, flow batch, semi-continuous, continuous, and  
reactor-type cultures, and may take multi-step  
10 processes connecting plural steps of these processes.

For respective growth substrates, specific culture  
steps will be described as follows:

<Culture: mcl-Alkanoates>

As a method of, for example, two-step culture,  
15 there is a method by which the first-step culture is  
carried out in an inorganic medium or the like  
containing a first alkanoate having 6 to 12 carbon  
atoms, such as octanoate and nonanoate, as the growth  
substrate at an amount of the order of 0.1% by weight  
20 to 0.2% by weight and a second alkanoate for  
introducing the desired monomer unit at an amount of  
the order of 0.01% by weight to 0.5% by weight until  
the time of the late logarithmic growth phase to the  
stationary phase, and at the second step, cells after  
25 the first-step culture is completed are collected by  
centrifugation or the like, followed by further  
culturing them in an inorganic medium containing the



second alkanoate at an amount of the order of 0.01% by weight to 0.5% by weight and no nitrogen sources, and after the culture is completed, the cells are harvested to extract the desired PHA.

5           Alternatively, there is another method by which culture is carried out by supplying a first alkanoate having 6 to 12 carbon atoms, such as octanoate and nonanoate, at an amount of the order of 0.1% by weight to 0.2% by weight and a second alkanoate for  
10       introducing the desired monomer unit at an amount of the order of 0.01% by weight to 0.5% by weight, and cells are harvested at the time of the late logarithmic growth phase to the stationary phase to extract the desired PHA.

15           In the methods in which mcl-alkanoate as the growth substrate is added to the medium, the obtained PHAs are ones in which is mixed a large amount of the monomer unit derived from the mcl-alkanoate added as the growth substrate. Such PHAs are generally of only  
20       the R form, and are isotactic polymers.

<Culture: Saccharides>

          As a method, for example, a two-step culture, there is a method by which the first-step culture is carried out in an inorganic medium or the like  
25       containing saccharide(s) (for example, glucose, mannose, fructose, etc.) as the growth substrate at an amount of the order of 0.1% by weight to 2.0% by weight

and alkanoate for introducing the desired monomer unit at an amount of the order of 0.01% by weight to 0.5% by weight until the time of the late logarithmic growth phase to the stationary phase, and at the second step, 5 cells after the first-step culture is completed are collected by centrifugation or the like, followed by further culturing them in an inorganic medium containing saccharide(s) (for example, glucose, mannose, fructose, etc.) as the growth substrate at an 10 amount of the order of 0.1% by weight to 2.0% by weight, the alkanoate at an amount of the order of 0.01% by weight to 0.5% by weight, and no nitrogen sources, and after the culture is completed, the cells are harvested to extract the desired PHA.

15 Alternatively, there is another method by which culture is carried out by supplying saccharide(s) (for example, glucose, mannose, fructose, etc.) as the growth substrate at amounts of the order of 0.1% by weight to 2.0% by weight and alkanoate for introducing 20 the desired monomer unit at an amount of the order of 0.01% by weight to 0.5% by weight, and cells are harvested at the time of the late logarithmic growth phase to the stationary phase to extract the desired PHA.

25 In these cases, the concentration of the saccharides (for example, glucose, mannose, fructose, etc.) to be added to the medium is selected as

appropriate, depending on the type of alkanoate for introducing the desired monomer unit, the genus and species of the microorganism, the cell density, or the culture process, although addition can be selected such that the content in the medium is usually in the order of 0.1% by weight to 2.0% by weight. On the other hand, the concentration of the alkanoate to be raw material is also selected as appropriate, depending on the genus and species of the microorganism, the cell density, or the culture process, although addition can be selected such that the content in the medium is usually in the order of 0.01% by weight to 0.5% by weight. Thus, by culturing the microorganism in a medium containing saccharide(s) (for example, glucose, mannose, fructose, etc.) and the alkanoate, the desired PHAs can be produced and accumulated in which the monomer unit other than the intended one is incorporated at a small amount or not at all. These PHAs are generally of only the R form, and are isotactic polymers.

<Culture: Polypeptone>

As a method of, for example, a two-step culture, there is a method by which the first-step culture is carried out in an inorganic medium or the like containing polypeptone as the growth substrate at an amount of the order of 0.1% by weight to 2.0% by weight and alkanoate for introducing the desired monomer unit

at an amount of the order of 0.01% by weight to 0.5% by weight until the time of the late logarithmic growth phase to the stationary phase, and at the second step, cells after the first-step culture is completed are  
5 collected by centrifugation or the like, followed by further culturing them in an inorganic medium containing the alkanoate at an amount of the order of 0.01% by weight to 0.5% by weight, and no nitrogen sources, and after the culture is completed, the cells  
10 are harvested to extract the desired PHA.

Alternatively, there is another method by which culture is carried out by supplying polypeptone at an amount of the order of 0.1% by weight to 2.0% by weight and alkanoate for introducing the desired monomer unit  
15 at an amount of the order of 0.01% by weight to 0.5% by weight, and cells are harvested at the time of the late logarithmic growth phase to the stationary phase to extract the desired PHA.

In these cases, the concentration of polypeptone  
20 to be added to the medium is selected as appropriate, depending on the type of alkanoate for introducing the desired monomer unit, the genus and species of the microorganism, the cell density, or the culture process, although addition can be selected such that  
25 the content in the medium is usually in the order of 0.1% by weight to 2.0% by weight. For polypeptone, it is also possible to use, as appropriate, any commercial

available polypeptone that is commonly employed for culturing microorganisms and the like. On the other hand, the concentration of the alkanoate to be raw material is also selected as appropriate, depending on the genus and species of the microorganism, the cell density, or the culture process, although addition may be selected such that the content in the medium is usually in the order of 0.01% by weight to 0.5% by weight. Thus, by culturing the microorganism in a medium containing polypeptone and the alkanoate, the desired PHAs can be produced and accumulated in which the monomer unit other than the intended one is incorporated at a small amount or not at all. These PHAs are generally of only the R form, and are isotactic polymers.

<Culture: Organic Acids of TCA Cycle>

As a method of, for example, a two-step culture, there is a method by which the first-step culture is carried out in an inorganic medium or the like containing organic acid(s) involved in the TCA cycle (for example, lactic, pyruvic, citric, succinic, fumaric, malic acids and the like, and salts thereof) as the growth substrate at an amount of the order of 0.1% by weight to 2.0% by weight and alkanoate for introducing the desired monomer unit at an amount of the order of 0.01% by weight to 0.5% by weight until the time of the late logarithmic growth phase to the

stationary phase, and at the second step, cells after the first-step culture is completed are collected by centrifugation or the like, followed by further culturing them in an inorganic medium containing  
5 organic acid(s) of the TCA cycle (for example, lactic, pyruvic, citric, succinic, fumaric, malic acids and the like, and salts thereof) as the growth substrate at an amount of the order of 0.1% by weight to 2.0% by weight, the alkanoate at an amount of the order of  
10 0.01% by weight to 0.5% by weight, and no nitrogen sources, and after the culture is completed, the cells are harvested to extract the desired PHA.

Alternatively, there is another method by which culture is carried out by supplying organic acid(s) of  
15 the TCA cycle (for example, lactic, pyruvic, citric, succinic, fumaric, malic acids and the like, and salts thereof) at an amount of the order of 0.1% by weight to 2.0% by weight and alkanoate for introducing the desired monomer unit at an amount of the order of 0.01%  
20 by weight to 0.5% by weight, and cells are harvested at the time of the late logarithmic growth phase to the stationary phase to extract the desired PHA.

In these cases, the concentration of organic acids of the TCA cycle (for example, lactic, pyruvic, citric,  
25 succinic, fumaric, malic acids and the like, and salts thereof) to be added to the medium is selected as appropriate, depending on the type of alkanoate for

introducing the desired monomer unit, the genus and species of the microorganism, the cell density, or the culture process, although addition can be selected such that the content in the medium is usually in the order of 0.1% by weight to 2.0% by weight. On the other hand, the concentration of the alkanoate to be raw material is also selected as appropriate, depending on the genus and species of the microorganism, the cell density, or the culture process, although addition can be selected such that the content in the medium is usually in the order of 0.01% by weight to 0.5% by weight. Thus, by culturing the microorganism in a medium containing organic acid(s) of the TCA cycle (for example, lactic, pyruvic, citric, succinic, fumaric, malic acids and the like, and salts thereof) and the alkanoate, the desired PHAs can be produced and accumulated in which the monomer unit other than the intended one is incorporated at a small amount or not at all. These PHAs are generally of only the R form, and are isotactic polymers.

<Culture: Polypeptone + Pyruvic Acid and Salts Thereof>

As a method of, for example, a two-step culture, there is a method by which the first-step culture is carried out in an inorganic medium or the like containing polypeptone as the growth substrate at an amount of the order of 0.1% by weight to 2.0% by weight and alkanoate for introducing the desired monomer unit

at an amount of the order of 0.01% by weight to 0.5% by weight until the time of the late logarithmic growth phase to the stationary phase, and at the second step, cells after the first-step culture is completed are  
5 collected by centrifugation or the like, followed by further culturing them in an inorganic medium containing pyruvic acid or salt thereof as the growth substrate at an amount of the order of 0.1% by weight to 2.0% by weight, the alkanoate at an amount of the  
10 order of 0.01% by weight to 0.5% by weight, and no nitrogen sources, and after the culture is completed, the cells are harvested to extract the desired PHA.

In these cases, the concentration of polypeptone and pyruvic acid or salt thereof to be added to the  
15 medium is selected as appropriate, depending on the type of alkanoate for introducing the desired monomer unit, the genus and species of the microorganism, the cell density, or the culture process, although addition can be selected such that the content in the medium is  
20 usually in the order of 0.1% by weight to 2.0% by weight in each case. On the other hand, the concentration of the alkanoate to be raw material is also selected as appropriate, depending on the genus and species of the microorganism, the cell density, or  
25 the culture process, although addition can be selected such that the content in the medium is usually in the order of 0.01% by weight to 0.5% by weight. Thus, by



culturing the microorganism in two steps utilizing a medium containing polypeptone and the alkanoate and a medium containing pyruvic acid or salt thereof and the alkanoate, the desired PHAs can be produced and  
5 accumulated in which the monomer unit other than the intended one is incorporated at a small amount or not at all. These PHAs are generally of only the R form, and are isotactic polymers.

<PHA Recovery>

10 For PHA recovery from cells in the method according to the present invention, usually-operating extraction with organic solvents such as chloroform is most convenient, but in the circumstances where it is difficult to use organic solvents, it is also possible  
15 to utilize methods of collecting PHA by removing cellular components other than PHA by means of treatment with detergents such as SDS and the like, treatment with enzymes such as lysozyme and the like, treatment with chemicals such as EDTA, sodium  
20 hypochlorite, ammonia, and the like.

<Molecular Weight>

The PHAs according to the present invention can be obtained by utilizing the above-mentioned methods. It is desirable that the PHAs have a number average  
25 molecular weight of more than at least 10,000 or so, in order to allow stable physical properties as polymer, for example, such as glass transition temperature,

softening point, melting point, crystallinity,  
orientation defined by the monomer unit of which the  
polymer is composed, to be fixed. The PHAs according  
to the present invention have a number average  
5 molecular weight of about 20,000 or higher, and  
therefore can be sufficiently expected to display  
stable physical properties as polymer. From the  
viewpoint of convenience of treatments such as  
dissolving processes, the PHAs preferably have a number  
10 average molecular weight of up to 200,000 or so, and  
more preferably not more than 100,000. As mentioned  
above, these PHAs are generally composed of only the R  
form, and are isotactic polymers.

Culturing of the microorganisms, production and  
15 accumulation of PHAs within cells by the  
microorganisms, and recovery of PHAs from the cells are  
not limited to the methods described above. For  
example, in addition to four strains described above,  
microorganisms to be employed in the method of  
20 producing PHAs according to the present invention can  
utilize microorganisms having similar production  
capabilities of producing PHAs according to the present  
invention as those of these four strains.

It is likely that these PHAs are useful, for  
25 example, for device and medical materials and others,  
as well as applications in which common plastics are  
used. Those having fluorine atom(s), trifluoromethyl

group, and the like introduced as a substituent group, in particular, are expected to have a superior biocompatibility, and therefore applications to medical uses. Furthermore, they are predicted to have a water-repellent effect due to containing fluorine atom(s), trifluoromethyl group, and the like, and thus applications to water-repellent treatments in various fields are also possible. Specifically, applications to temporary water-repellent treatments also can be contemplated utilizing biodegradability resulting from aliphatic polyesters.

(Examples)

[Example 1]

The substrate FPVA was first synthesized by the Grignard reaction according to the method described in "Macromolecules, 29, 1762-1766 (1996) and 27, 45-49 (1994)". 5-bromovaleric acid was dissolved in anhydrous tetrahydrofuran (THF), and 3 mol/L of a methyl magnesium chloride THF solution was added dropwise at -20°C in an argon atmosphere. After stirring for about 15 min, a THF solution of 1-bromo-4-fluorobenzene and magnesium was further dropped and a THF solution of 0.1 mol/L  $\text{Li}_2\text{CuCl}_4$  was added (temperature was maintained at -20°C). The reaction solution was resumed to room temperature and further stirred overnight. Then the solution was poured into a 20% sulfuric acid solution cooled on ice, and stirred.

The aqueous layer was recovered, saturated with salt, and extracted with ether. After the extract was further extracted with 100 mL of deionized water, to which 50 g of potassium hydroxide was added, the  
5 extract was acidified with a 20% sulfuric acid solution and the precipitate was recovered.

The precipitate was analyzed with nuclear magnetic resonance equipment (FT-NMR: Bruker DPX400) under the following conditions: nuclide:  $^1\text{H}$  and  $^{13}\text{C}$ ; solvent:  
10 heavy chloroform (containing TMS). The results are shown in FIG. 1 and Table 3.

[Example 2]

Cells of strain H45 was inoculated in 200 mL of M9 medium containing nonanoic acid 0.1% and FPVA 0.1%, and  
15 shake-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, resuspended in 200 mL of M9 medium containing FPVA 0.2% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shake-cultured at 30°C at 125 strokes/min. After 24 hours,  
20 the cells were collected by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 20 hours to extract PHA. After the extract was filtered with a membrane  
25 filter of 0.45  $\mu\text{m}$  pore size, the filtrate was concentrated with a rotary evaporator and the concentrate was reprecipitated in cold methanol. Only

the precipitate was then recovered and vacuum-dried to obtain PHA. After the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. The results are shown in Table 4.

[Example 3]

Cells of strain YN2 was inoculated in 200 mL of M9 medium containing nonanoic acid 0.1% and FPVA 0.1%, and shake-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, resuspended in 200 mL of M9 medium containing FVPA 0.2% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shaken-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in 100 mL of chloroform, stirred at 60°C for 20 hours to extract PHA. After the extract was filtered with a membrane filter of 0.45  $\mu\text{m}$  in diameter, the filtrate was concentrated with a rotary evaporator and the concentrate was reprecipitated in cold methanol. The precipitate was then recovered and vacuum-dried to obtain PHA. After the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer

(GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. The results are shown in Table 5.

[Example 4]

- 5           Cells of strain P91 were inoculated in 200 mL of M9 medium containing nonanoic acid 0.1% and shaken-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, resuspended in 200 mL of M9 medium containing nonanoic acid 0.1% and FPVA 0.1% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shaken-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, washed once with cold methanol and lyophilized.
- 10
- 15           The lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 20 hours to extract PHA. After the extract was filtered with a membrane filter of 0.45  $\mu\text{m}$  in diameter, the filtrate was concentrated with a rotary evaporator and the concentrate was reprecipitated in cold methanol. Only the precipitate was then recovered and vacuum-dried to obtain PHA. After the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer
- 20
- 25           (GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. The results are shown in Table 6.

[Example 5]

Cells of strain P161 were inoculated in 200 mL of M9 medium containing nonanoic acid 0.1% and shaken-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, resuspended in 200 mL of M9 medium containing nonanoic acid 0.1% and FPVA 0.1% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shaken-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 20 hours to extract PHA. After the extract was filtered with a membrane filter of 0.45  $\mu\text{m}$  in diameter, the filtrate was concentrated with a rotary evaporator and the concentrate was reprecipitated in cold methanol. Only the precipitate was then recovered and vacuum-dried to obtain PHA. After the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. The results are shown in Table 7.

[Example 6]

After 100 mg of PHA derived from strain H45 was dissolved in 1 mL of chloroform, n-hexane was added

until it was clouded. This was centrifuged to recover and vacuum-dried the precipitate. This was again dissolved in 1 mL of chloroform, n-hexane was added, and the procedure of recovering the precipitate was repeated three times.

After the precipitate obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. As a result, the precipitate was found to be PHA whose monomer unit consisted of 3HFPV monomer alone as shown in Table 8.

In addition, nuclear magnetic resonance equipment (FT-NMR: Bruker DPX400) was used for analysis under the following condition: nuclide:  $^1\text{H}$  and  $^{13}\text{C}$ ; solvent: heavy chloroform (containing TMS). The results are shown in FIG. 2, Table 9, FIG. 3 and Table 10.

[Example 7]

(Synthesis of FPxVA)

After 240 mL of dehydrated acetone was put into a three-necked round-bottom flask, sodium iodide (0.06 mol), potassium carbonate (0.11 mol) and 4-fluorophenol (0.07 mol) were added and thoroughly stirred. 5-bromoethylvalerate (0.06 mol) was dropped into the solution in a nitrogen atmosphere, refluxed at  $60 \pm 5^\circ\text{C}$  and allowed to react for 24 hours. After reaction, the



reaction solution was concentrated to dryness with an evaporator and redissolved in methylene chloride. Water was added and the solution was separated. The organic layer was dehydrated with anhydrous magnesium sulfate and concentrated to dryness with an evaporator.

Hot methanol was added to the reactant, dissolved, slowly cooled and reprecipitated to obtain 5-(4-fluorophenoxy)ethylvalerate. At this time, the yield from 5-bromoethylvalerate was 68 mol %.

The reactant (ester) obtained was dissolved in ethanol-water (9.1 (v/v)) so as to be 5 weight %. Ten-fold molar quantity of potassium hydroxide was added and allowed to react at 0 to 4°C for 4 hours to hydrolyze the ester.

The reaction solution was poured into 10 volumes of a 0.1 mol/L hydrochloric acid solution and the precipitate was recovered by filtration. The precipitate (reactant) recovered was vacuum-dried at room temperature for 36 hours. The dried substance obtained was dissolved in a small quantity of hot ethanol, and the solution was gradually cooled, reprecipitated, vacuum-dried at room temperature for 24 hours to obtain the target compound 5-(4-fluorophenoxy)valeric acid. The yield of this compound from 5-bromoethyl valerate was 49 mol %.

The compound obtained was analyzed by NMR under the following conditions:

<Equipment>

FT-NMR: Bruker DPX400

$^1\text{H}$  resonance frequency: 400 MHz

<Measurement conditions>

5 nuclide:  $^1\text{H}$

solvent:  $\text{CDCl}_3$

reference: capillary-contained TMS/ $\text{CDCl}_3$

temperature: room temperature

10 The spectral chart is shown in FIG. 4 and the  
results of identification are shown in Table 11.

The above results confirmed that the desired FPxVA  
was certainly synthesized.

[Example 8]

(Production of PHA by strain P91)

15 Cells of strain P91 were inoculated in 200 mL of  
M9 medium containing nonanoic acid 0.1 weight % and  
FPxVA 0.1 weight %, and shaken-cultured at 30°C at 125  
strokes/min. After 24 hours, the cells were collected  
by centrifugation, resuspended in 200 mL of M9 medium  
20 containing nonanoic acid 0.1 weight % and FPxVA 0.1  
weight % but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further  
shaken-cultured at 30°C at 125 strokes/min. After 24  
hours, the cells were collected by centrifugation,  
washed once with cold methanol and lyophilized.

25 After the lyophilized pellet was weighed, it was  
suspended in 100 mL of chloroform and stirred at 60°C  
for 20 hours to extract PHA. After the extract was

filtered with a membrane filter of 0.45  $\mu\text{m}$  in pore size, the filtrate was concentrated with a rotary evaporator and the concentrate was reprecipitated in cold methanol. Only the precipitate was then recovered  
5 and vacuum-dried to obtain and weigh PHA. The yields are shown in Table 12.

The PHA obtained was measured for the molecular weight by gel permeation chromatography (GPC; Toso HCL-8020, column: Polymer Laboratory PLgel MIXED-C (5  $\mu\text{m}$ ),  
10 solvent: chloroform, converted on a polystyrene basis). The molecular weight is shown in Table 13.

In addition, after the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed by GC-MS and the methyl esters of the  
15 PHA monomer unit were identified. The TIC and the mass spectrum of each peak are shown in FIG. 5 to FIGS. 7A and 7B. Peak (1) was shown to represent 3-hydroxymethylheptanoate, Peak (2) 3-hydroxymethyloctanoate, Peak (3) 3-hydroxymethylnonanoate, and Peak (4) 3 hydroxy-4-(4-fluorophenoxy) methylvalerate.  
20

The above results indicated that the polymer obtained was PHA containing the units of 3-hydroxyheptanoic acid, 3-hydroxyoctanoic acid, 3-hydroxynonanoic acid and 3-hydroxy-4-(4-fluorophenoxy)valeric acid.  
25

[Example 9]

(Production of PHA by strain YN2 (1))

Except that strain P91 was replaced by strain YN2, the same procedures as described in Example 8 were used to produce PHA, and each analysis was performed. The yields are shown in Table 12, the molecular weight in Table 13, and the TIC of GC-MS in FIG. 8. Peak (1) was shown to represent 3-hydroxymethylheptanoate, Peak (2) 3-hydroxymethyloctanoate, Peak (3) 3-hydroxymethylnonanoate, and Peak (4) 3-hydroxy-4-(4-fluorophenoxy) methylvalerate.

The above results indicated that the polymer obtained was PHA containing the units of 3-hydroxyheptanoic acid, 3-hydroxyoctanoic acid, 3-hydroxynonanoic acid and 3-hydroxy-4-(4-fluorophenoxy)valeric acid.

[Example 10]

(Production of PHA by strain P161)

Except that strain P91 was replaced by strain P161, the same procedures as described in Example 8 were used to produce PHA, and each analysis was performed. The yields are shown in Table 12, the molecular weight in Table 13, and the TIC of GC-MS in FIG. 9. Peak (1) was shown to represent 3-hydroxymethylheptanoate, Peak (2) 3-hydroxymethyloctanoate, Peak (3) 3-hydroxymethylnonanoate, and Peak (4) 3 hydroxy-4-(4-fluorophenoxy) methylvalerate.

The above results indicated that the polymer obtained was PHA containing the units of 3-hydroxyheptanoic acid, 3-hydroxyoctanoic acid, 3-hydroxynonanoic acid and 3-hydroxy-4-(4-fluorophenoxy)valeric acid.

[Example 11]

(Production of PHA by strain H45)

Except that strain P91 was replaced by H45 strain, the same procedures as described in Example 8 were used to produce PHA, and each analysis was performed. The yields are shown in Table 12, the molecular weight in Table 13, and the TIC of GC-MS in FIG. 10. Peak (1) was shown to represent 3-hydroxymethylheptanoate, Peak (2) 3-hydroxymethyloctanoate, Peak (3) 3-hydroxymethylnonanoate, and Peak (4) 3-hydroxy-4-(4-fluorophenoxy) methylvalerate.

The above results indicated that the polymer obtained was PHA containing the units of 3-hydroxyheptanoic acid, 3-hydroxyoctanoic acid, 3-hydroxynonanoic acid and 3-hydroxy-4-(4-fluorophenoxy)valeric acid.

[Example 12]

(Production of PHA by strain YN2 (2))

Cells of strain YN2 were inoculated in 200 mL of M9 medium containing hexanoic acid 0.1 weight % and FPxVA 0.1 weight %, and shaken-cultured at 30°C at 125 strokes/min. After 72 hours, the cells were collected

by centrifugation and resuspended in 200 mL of M9 medium containing hexanoic acid 0.1 weight % and FPxVA 0.1 weight % but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shaken-cultured at 30°C at 125 strokes/min.

- 5 After 30 hours, the cells were collected by centrifugation, washed once with cold methanol and lyophilized.

PHA was extracted with the same procedures as described in Example 8, and each analysis was  
10 performed. The yields are shown in Table 12 and the molecular weight is shown in Table 13. The results of GC-MS analysis revealed that the PHA obtained by this method had the following composition:

- 3-hydroxybutyric acid: 8.1%  
15 3-hydroxyhexanoic acid: 51.2%  
3-hydroxyoctanoic acid: 1.3%  
3-hydroxydecanoic acid: 7.0%  
3-hydroxydodecanoic acid: 10.6%  
unidentified substances: 9.9%  
20 3 hydroxy-4-(4-fluorophenoxy)valeric acid: 11.9%

The above results indicated that the polymer obtained was PHA containing the units of 3-hydroxy-4-(4-fluorophenoxy)valeric acid.

[Example 13]

- 25 (Synthesis of TFMPVA)

The substrate TFMPVA was first synthesized by the Grignard reaction according to the method described in

"Macromolecules, 29, 1762-1766 (1996) and 27, 45-49 (1994)." 5-bromovaleric acid was dissolved in anhydrous tetrahydrofuran (THF) and 3 mol/L of a methyl magnesium chloride THF solution was added dropwise at - 20°C in an argon atmosphere. After stirring for about 15 min, a THF solution of 1-bromo-4-trifluorobenzene and magnesium was further dropped and a THF solution of 0.1 mol/L  $\text{Li}_2\text{CuCl}_2$  was added (temperature was maintained at -20°C). The reaction solution was restored to room temperature and further stirred overnight. Then the solution was poured into a 20% sulfuric acid solution cooled on ice, and stirred. The aqueous layer was recovered, saturated with salt, and extracted with ether. After the extract was further extracted with 100 mL of deionized water, to which 50 g of potassium hydroxide was added, the extract was acidified with a 20% sulfuric acid solution and the precipitate was recovered.

The compound recovered was methylesterified by the conventional method and analyzed with a gas chromatograph-mass spectrometer (GC-MS, Shimadzu GC-MS QP-5050, column: DB-WAXETR (30 m  $\times$  0.32 mm  $\times$  0.5  $\mu\text{m}$ ) (manufactured by J&W Inc.) The TIC (total ion chromatogram) and mass spectrum are shown in FIGS. 11A and 11B. These results revealed that the target TFMPVA was synthesized.

[Example 14]

(Production of polymer by strain H45)

Cells of strain H45 were inoculated in 200 mL of M9 medium containing nonanoic acid 0.1% and TFMPVA 0.1%, and shaken-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, resuspended in 200 mL of M9 medium containing TFMPVA 0.2% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shaken-cultured at 30°C at 125 strokes/min. After 24 hours, the cells were collected by centrifugation, washed once with cold methanol, lyophilized and weighed.

The lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 20 hours to extract PHA. After the extract was filtered with a membrane filter of 0.45  $\mu\text{m}$  in pore size, the filtrate was concentrated with a rotary evaporator and the concentrate was reprecipitated in cold methanol. Only the precipitate was then recovered and vacuum-dried to obtain and weigh PHA. The yields are shown in Table 14.

Evaluation of the molecular weight of the PHA by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5  $\mu\text{m}$ ), solvent: chloroform, converted on a polystyrene basis) revealed  $M_n=64,000$  and  $M_w=110,000$ .

After the PHA obtained was subjected to methanolysis according to the conventional method, it



was analyzed by gas chromatography-spectrometry (GC-MS, Shimadzu QP-5050, EI method, column: DB-WAXETR (30 m × 0.32 mm × 0.5 μm)) and the methyl esters of the PHA monomer unit were identified. The TIC (total ion chromatogram) and the mass spectrum of a peak (close to 36.5') representing the target unit 3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid are shown in FIGS. 12A and 12B, respectively. The TIC area ratio of each unit of the PHA is shown in Table 15.

The above results indicated that one method of the present invention produced PHA containing 3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid as a monomer unit.

[Example 15]

(Production of polymer by strain P91)

Cells of strain P91 were inoculated in 200 mL of M9 medium containing nonanoic acid 0.1% and TFMPVA 0.1%, and shaken-cultured at 30°C at 125 strokes/min. After 30 hours, the cells were collected by centrifugation, resuspended in 200 mL of M9 medium containing TFMPVA 0.1% and nonanoic acid 0.05% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shaken-cultured at 30°C at 125 strokes/min. After 30 hours, the cells were collected by centrifugation, washed once with cold methanol, lyophilized and weighed.

The lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 20 hours to extract

PHA. After the extract was filtered with a membrane filter of 0.45  $\mu$ m in pore size, the filtrate was concentrated with a rotary evaporator and the concentrate reprecipitated in cold methanol. Only the precipitate was then recovered and vacuum-dried to obtain and weigh PHA.

The yield is shown in Table 16.

Evaluation of the molecular weight of the PHA by gel permeation chromatography (GPC; Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5  $\mu$ m), solvent: chloroform, converted on a polystyrene basis) revealed  $M_n$ =69,000 and  $M_w$ =120,000.

After the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method, column: DB-WAXETR (30 m  $\times$  0.32 mm  $\times$  0.5  $\mu$ m)) and the methyl esters of the PHA monomer unit were identified. The TIC (total ion chromatogram) and the mass spectrum of a peak (close to 36.5') representing the target unit 3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid are shown in FIGS. 13A and 13B, respectively. The TIC area ratio of each unit of the PHA is shown in Table 17.

The above results indicated that one method of the present invention produced PHA containing 3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid as a monomer unit.

[Example 16]

Cells of strain YN2 were inoculated in 200 mL of M9 medium containing D-glucose 0.5% or n-nonanoic acid 0.1% and PVA 0.1%, and shaken-cultured at 30°C at 125  
5 strokes/min. After 40 hours, the cells were collected by centrifugation, washed once with cold methanol and lyophilized to obtain a lyophilized pellet.

The lyophilized pellet was suspended in 20 mL of chloroform and stirred at 60°C for 28 hours to extract  
10 PHA. After the extract was filtered with a membrane filter of 0.45  $\mu$ m in pore size, the filtrate was concentrated with a rotary evaporator and the concentrate was resuspended in cold methanol. Only the precipitate was then recovered and vacuum-dried to  
15 obtain PHA. After the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. As a  
20 result, PHA containing 3HPV being the desired PVA-derived monomer unit at a higher ratio was obtained in high yield by using D-glucose as a carbon source for growth as shown in Table 18.

[Example 17]

25 Cells of strain YN2 were inoculated in 200 mL of M9 medium containing D-glucose 0.5% or n-nonanoic acid 0.1% and PVA 0.1%, and shaken-cultured at 30°C at 125

strokes/min. After 48 hours, the cells were collected by centrifugation, resuspended in 200 mL of M9 medium containing D-glucose 0.5% or n-nonanoic acid 0.1% and PVA 0.1% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further  
5 shaken-cultured at 30°C at 125 strokes/min. After 40 hours, the cells were collected by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in 20 mL of chloroform, stirred at 60°C for 24 hours to extract  
10 PHA. After the extract was filtered with a membrane filter of 0.45  $\mu\text{m}$  in pore size, the filtrate was concentrated with a rotary evaporator and the concentrate was reprecipitated in cold methanol. Only the precipitate was then recovered and vacuum-dried to  
15 obtain PHA. After the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. As a  
20 result, PHA containing 3HPV being the desired PVA-derived monomer unit at a higher ratio was obtained in a high yield by using D-glucose as a carbon source for growth, as shown in Table 19.

[Example 18]

25 Cells of strain YN2 were inoculated in 200 mL of M9 medium containing D-mannose 0.5% or D-fructose 0.5% and PVA 0.1%, and cultured with shaking at 30°C at 125

strokes/min. After 100 hours in the D-mannose system and 40 hours in the D-fructose system, the cells were collected by centrifugation, resuspended in 200 mL of M9 medium containing D-mannose 0.5% or D-fructose 0.5% and PVA 0.1% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shaken-cultured at 30°C at 125 strokes/min. After 48 hours, the cells were collected by centrifugation, washed once with cold methanol and lyophilized.

10           The lyophilized pellet was suspended in 20 mL of chloroform and stirred at 60°C for 24 hours to extract PHA. After the extract was filtered with a membrane filter of 0.45  $\mu\text{m}$  in pore size, the filtrate was concentrated with a rotary evaporator and the  
15           concentrate was reprecipitated in cold methanol. The precipitate was then recovered and vacuum-dried to obtain PHA. After the PHA obtained was subjected to methanolysis according to the conventional method, it was analyzed with a gas chromatograph-mass spectrometer  
20           (GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. As a result, as shown in Table 20, D-mannose and D-fructose were also as effective as D-glucose as the carbon  
25           source to obtain PHA having a high proportion of 3HPV being the desired PVA-derived monomer unit at a high PHA yield.

[Example 19]

Cells of strain P161 were inoculated in 200 mL of M9 medium containing D-glucose 0.5% or n-nonanoic acid 0.1% and PVA 0.1%, and cultured with shaking at 30°C at 125 strokes/min. After 48 hours, the cells were  
5 collected by centrifugation, resuspended in 200 mL of M9 medium containing D-glucose 0.5% or n-nonanoic acid 0.1% and PVA 0.1% but not a nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further cultured at 30°C at 125 strokes/min. After 40 hours, the cells were collected by centrifugation,  
10 washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in 20 mL of chloroform and stirred at 60°C for 24 hours to extract PHA. After the extract was filtered with a membrane filter of 0.45  $\mu\text{m}$  in pore size, the filtrate was  
15 concentrated with a rotary evaporator and the concentrate was reprecipitated in cold methanol. Only the precipitate was then recovered and vacuum-dried to obtain PHA. After the PHA obtained was subjected to methanolysis according to the conventional method, it  
20 was analyzed with a gas chromatograph-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) and the methyl esters of the PHA monomer unit were identified. As a result, PHA having a higher proportion of 3HPV as the desired PVA-derived monomer unit was obtained in high  
25 yield by using D-glucose as a carbon source, as shown in Table 21.

[Example 20]

The cells of strain YN2 were shake-cultured in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of FPVA under the conditions of 30°C and 125 strokes/min for 48 hours.

5 The cells were recovered by centrifugal separation, and resuspended in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of FPVA but no nitrogen source ( $\text{NH}_4\text{Cl}$ ), where they were further shake-cultured under the conditions of 30°C and 125  
10 strokes/min for 40 hours. The cells were recovered by centrifugal separation, washed once with cold methanol, and lyophilized.

The lyophilized pellet was suspended in 20 mL of chloroform, and stirred at 60°C for 24 hours, to  
15 extract the PHA. The extract solution was filtered through a membrane having a pore size of 0.45  $\mu\text{m}$  and concentrated by a rotary evaporator. The concentrated solution was reprecipitated in cold methanol, and the precipitate only was recovered and dried under a  
20 vacuum, to obtain the PHA. The PHA thus prepared was subjected to methanolysis by the normal procedure, and analyzed by a gas chromatograph/mass spectrometer (GC-MS, Shimadzu QP-5050, based on the EI method), to identify the methyl-esterified product of the PHA  
25 monomer units. The results are given in Table 22. As shown, culturing with D-glucose as the carbon source for growth gives the PHA having a higher proportion of

3-hydroxy-5-(4-fluorophenyl)valeric acid as the desired FPVA-derived monomer unit in higher yield.

[Example 21]

Cells of strain P161 was shake-cultured in 200 mL  
5 of M9 medium, containing 0.5% of D-glucose or 0.1% of  
n-nonanoic acid and 0.1% of PHxA under the conditions  
of 30°C and 125 strokes/min for 48 hours. The cells  
were recovered by centrifugation, and resuspended in  
200 mL of M9 medium, containing 0.5% of D-glucose or  
10 0.1% of n-nonanoic acid and 0.1% of PHxA but no  
nitrogen source ( $\text{NH}_4\text{Cl}$ ), where they were further  
shake-cultured under the conditions of 30°C and 125  
strokes/min for 40 hours. The cells were recovered by  
centrifugation, washed once with cold methanol, and  
15 lyophilized.

The lyophilized pellet was suspended in 20 mL of  
chloroform, and stirred at 60°C for 24 hours, to  
extract the PHA. The extract solution was filtered  
through a membrane having a pore size of 0.45  $\mu\text{m}$  and  
20 concentrated by a rotary evaporator. The concentrated  
solution was reprecipitated in cold methanol, and the  
precipitate only was recovered and dried under a  
vacuum, to obtain the PHA. The PHA thus prepared was  
subjected to methanolysis by the normal procedure, and  
25 analyzed by a gas chromatograph/mass spectrometer  
(GC-MS, Shimadzu QP-5050, based on the EI method), to  
identify the methyl-esterified product of the PHA



monomer units. The results are given in Table 23. As shown, culturing with D-glucose as the carbon source for growth gives the PHA having a higher proportion of 3-hydroxy-6-phenylhexanoic acid as the desired PHxA-derived monomer unit in higher yield.

[Example 22]

Cells of strain YN2 were shake-cultured in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of PxBA under the conditions of 30°C and 125 strokes/min for 48 hours. The cells were recovered by centrifugation, and resuspended in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of PxBA but no nitrogen source ( $\text{NH}_4\text{Cl}$ ), where they were further shake-cultured under the conditions of 30°C and 125 strokes/min for 40 hours. The cells were recovered by centrifugation, washed once with cold methanol, and lyophilized.

The lyophilized pellet was suspended in 20 mL of chloroform, and stirred at 60°C for 24 hours, to extract the PHA. The extract solution was filtered through a membrane having a pore size of 0.45  $\mu\text{m}$  and concentrated by a rotary evaporator. The concentrated solution was reprecipitated in cold methanol, and the precipitate only was recovered and dried under a vacuum, to obtain the PHA. The PHA thus prepared was subjected to methanolysis by the normal procedure, and

analyzed by a gas chromatograph/mass spectrometer (GC-MS, Shimadzu QP-5050, based on the EI method), to identify the methyl-esterified product of the PHA monomer units. The results are given in Table 24. As shown, culturing with D-glucose as the carbon source for growth gives the PHA having a higher proportion of 3-hydroxy-4-phenoxybutyric acid as the desired PxBA-derived monomer unit in higher yield.

[Example 23]

Cells of strain H45 was shake-cultured in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of PxBA under the conditions of 30°C and 125 strokes/min for 48 hours. The cells were recovered by centrifugation, and resuspended in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of PxBA but no nitrogen source ( $\text{NH}_4\text{Cl}$ ), where they were further shake-cultured under the conditions of 30°C and 125 strokes/min for 40 hours. The cells were recovered by centrifugation, washed once with cold methanol, and lyophilized.

The lyophilized pellet was suspended in 20 mL of chloroform, and stirred at 60°C for 24 hours, to extract the PHA. The extract solution was filtered through a membrane having a pore size of 0.45  $\mu\text{m}$  and concentrated by a rotary evaporator. The concentrated solution was reprecipitated in cold methanol, and the

precipitate only was recovered and dried under a vacuum, to obtain the PHA. The PHA thus prepared was subjected to methanolysis by the normal procedure, and analyzed by a gas chromatograph/mass spectrometer (GC-MS, Shimadzu QP-5050, based on the EI method), to identify the methyl-esterified product of the PHA monomer units. The results are given in Table 25. As shown, culturing with D-glucose as the carbon source for growth gives the PHA having a higher proportion of 3-hydroxy-4-phenoxybutyric acid as the desired PxBA-derived monomer unit in higher yield.

[Example 24]

Cells of strain P161 were shake-cultured in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of PxBA under the conditions of 30°C and 125 strokes/min for 48 hours. The cells were recovered by centrifugation, and resuspended in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of PxBA but no nitrogen source ( $\text{NH}_4\text{Cl}$ ), where they were further shake-cultured under the conditions of 30°C and 125 strokes/min for 40 hours. The cells were recovered by centrifugation, washed once with cold methanol, and lyophilized.

The lyophilized pellet was suspended in 20 mL of chloroform, and stirred at 60°C for 24 hours, to extract the PHA. The extract solution was filtered

through a membrane having a pore size of 0.45  $\mu\text{m}$  and concentrated by a rotary evaporator. The concentrated solution was reprecipitated in cold methanol, and the precipitate only was recovered and dried under a vacuum, to obtain the PHA. The PHA thus prepared was subjected to methanolysis by the normal procedure, and analyzed by a gas chromatograph/mass spectrometer (GC-MS, Shimadzu QP-5050, based on the EI method), to identify the methyl-esterified product of the PHA monomer units. The results are given in Table 26. As shown, culturing with D-glucose as the carbon source for growth gives the PHA having a higher proportion of 3-hydroxy-4-phenoxybutyric acid as the desired PxBA-derived monomer unit in higher yield.

[Example 25]

Cells of strain YN2 were shake-cultured in 200 mL of M9 medium, containing 0.5% of D-glucose or 0.1% of n-nonanoic acid and 0.1% of CHBA under the conditions of 30°C and 125 strokes/min for 40 hours. The cells were recovered by centrifugation, washed once with cold methanol, and lyophilized.

The lyophilized pellet was suspended in 20 mL of chloroform, and stirred at 60°C for 28 hours, to extract the PHA. The extract solution was filtered through a membrane having a pore size of 0.45  $\mu\text{m}$  and concentrated by a rotary evaporator. The concentrated solution was reprecipitated in cold methanol, and the

precipitate only was recovered and dried under a vacuum, to obtain the PHA. The PHA thus prepared was subjected to methanolysis by the normal procedure, and analyzed by a gas chromatograph/mass spectrometer (GC-MS, Shimadzu QP-5050, based on the EI method), to identify the methyl-esterified product of the PHA monomer units. The results are given in Table 27. As shown, culturing with D-glucose as the carbon source for growth gives the PHA having a higher proportion of 3HCHB as the desired CHBA-derived monomer unit in higher yield.

[Example 26]

(Production of poly-3-hydroxy-5-phenylvaleric acid by strain YN2)

A colony of strain YN2 on M9 agar medium containing 0.1% of nonanoic acid (hereinafter referred as to NA) was inoculated in a total of 4 types of media (each 200 mL), (1) M9 liquid medium containing 0.5% of a yeast extract (DIFCO, hereinafter referred to as YE) and 0.1% of 5-phenylvaleric acid, (2) M9 liquid medium containing 0.5% of a beef extract (DIFCO, hereinafter referred to as BE) and 0.1% of 5-phenylvaleric acid, (3) M9 liquid medium containing 0.5% of Casamino acid (DIFCO, hereinafter referred to as CA) and 0.1% of 5-phenylvaleric acid, and (4) M9 liquid medium containing 0.5% of polypeptone (Wako Jun-yaku, hereinafter referred to as PP) and 0.1% of

5-phenylvaleric acid, and cultured at 30°C for 24 hours. The cells were recovered from each medium by centrifugation, washed once with cold methanol, and lyophilized.

5           The lyophilized pellets from each medium were weighed and suspended in 100 mL of chloroform, and stirred at 55°C for 20 hours, to extract the PHA. Each extract solution was filtered through a membrane having a pore size of 0.45  $\mu$ m and concentrated by a  
10 evaporator. The concentrated solution was reprecipitated in cold methanol, to obtain the polymer, which was dried under a vacuum at room temperature and weighed. The yield results are given in Table 28.

          The PHA composition thus prepared was analyzed by  
15 the following procedure. Approximately 10 mg of the PHA was dissolved in 2 mL of chloroform in a 25 mL egg-plant type flask, to which 2 mL of a methanol solution containing 3% sulfuric acid was added. The mixture was heated at 100°C with reflux for 3.5 hours  
20 for the reactions. On completion of the reactions, the effluent was incorporated with 10 mL of deionized water and vigorously shaken for 10 min. It was separated into two layers, and the lower chloroform layer was taken out, dehydrated with magnesium sulfate, and  
25 analyzed by a gas chromatograph/mass spectrometer (GC-MS, Shimadzu QP-5050, based on the EI method, with a 0.32 mm by 30 m column (J&W, DB-WAX)), to identify

the methyl-esterified product of the PHA monomer units. The PHA monomer units were found to comprise 96% of 3-hydroxy-5-phenylvaleric acid unit and 4% of 3-hydroxybutyric acid unit.

5           The results indicate that one of the embodiments of the present invention gives the PHA containing a very high proportion of the 3-hydroxy-5-phenylvaleric acid unit in a high yield.

[Example 27]

10          (Production of poly-3-hydroxy-4-cyclohexylbutyric acid by strain YN2)

A colony of strain YN2 grown on M9 agar medium containing 0.1% of NA was inoculated into 2 types of media (each 200 mL), (1) M9 liquid medium containing  
15       0.5% of YE and 0.1% of 4-cyclohexylbutyric acid, and (2) M9 liquid medium containing 0.5% of PP and 0.1% of 4-cyclohexylbutyric acid, and cultured at 30°C for 24 hours. The cells were recovered from each medium by centrifugation, washed once with cold methanol, and  
20       lyophilized.

The lyophilized pellets from each medium were weighed and suspended in 100 mL of chloroform, and stirred at 55°C for 20 hours, to extract the PHA. Each extract solution was filtered through a membrane having  
25       a pore size of 0.45  $\mu$ m and concentrated by a evaporator. The concentrated solution was reprecipitated in cold methanol, to obtain the polymer,

which was dried under a vacuum at room temperature and weighed. The yield results are given in Table 29.

The PHA composition thus prepared was analyzed in a manner similar to that for EXAMPLE 26. The PHA  
5 monomer units were found to comprise 97% of 3-hydroxy-4-cyclohexylbutyric acid unit and 3% of 3-hydroxybutyric acid unit.

The results indicate that one of the embodiments of the present invention gives the PHA containing a  
10 very high proportion of the 3-hydroxy-4-cyclohexylbutyric acid unit in a high yield.

[Example 28]

(Production of poly-3-hydroxy-5-phenoxyvaleric acid by  
15 strain YN2)

A colony of strain YN2 grown on M9 agar medium containing 0.1% of NA was inoculated in 2 types of media (each 200 mL), (1) M9 liquid medium containing 0.5% of YE and 0.1% of 5-phenoxyvaleric acid, and (2)  
20 M9 liquid medium containing 0.5% of PP and 0.1% of 5-phenoxyvaleric acid, and cultured at 30°C for 26 hours. The cells were recovered from each medium by centrifugation, washed once with cold methanol, and lyophilized.

25 The lyophilized pellets from each medium were weighed and suspended in 100 mL of chloroform, and stirred at 55°C for 20 hours, to extract the PHA. Each



extract solution was filtered through a membrane having a pore size of 0.45  $\mu$ m and concentrated by an evaporator. The concentrated solution was reprecipitated in cold methanol, to obtain the polymer, which was dried under a vacuum at room temperature and weighed. The yield results are given in Table 30.

The PHA composition thus prepared was analyzed in a manner similar to that in EXAMPLE 26. The PHA monomer units were found to comprise 95% of 3-hydroxy-5-phenoxyvaleric acid unit and 5% of 3-hydroxybutyric acid unit.

The results indicate that one of the embodiments of the present invention gives the PHA containing a very high proportion of the 3-hydroxy-5-phenoxyvaleric acid unit in a high yield.

[Example 29]

(Production of poly-3-hydroxy-5-phenylvaleric acid by strain H45)

A colony of strain H45 grown on M9 agar medium containing 0.1% of NA was inoculated in 4 types of media (each 200 mL), (1) M9 liquid medium containing 0.5% of YE and 0.1% of 5-phenylvaleric acid, (2) M9 liquid medium containing 0.5% of sodium glutamate (Kishida Kagaku, hereinafter referred to as SG) and 0.1% of 5-phenylvaleric acid, (3) M9 liquid medium containing 0.5% of CA and 0.1% of 5-phenylvaleric acid, and (4) M9 liquid medium containing 0.5% of PP and 0.1%

of 5-phenylvaleric acid, and cultured at 30°C for 28 hours. The cells were recovered from each medium by centrifugation, washed once with cold methanol, and lyophilized.

5           The lyophilized pellets from each medium were weighed and suspended in 100 mL of chloroform, and stirred at 55°C for 20 hours, to extract the PHA. Each extract solution was filtered through a membrane having a pore size of 0.45  $\mu$ m and concentrated by a  
10       evaporator. The concentrated solution was reprecipitated in cold methanol, to obtain the polymer, which was dried under a vacuum at room temperature and weighed. The yield results are given in Table 31.

          The PHA composition thus prepared was analyzed in  
15       a manner similar to that in EXAMPLE 26. The PHA was found to be essentially the homopolymer of 3-hydroxy-5-phenylvaleric acid.

          The results indicate that one of the embodiments of the present invention gives the PHA containing a  
20       very high proportion of the 3-hydroxy-5-phenylvaleric acid unit in a high yield.

[Example 30]

(Production of poly-3-hydroxy-5-phenylvaleric acid by strain P161)

25           A colony of strain P161 grown on M9 agar medium containing 0.1% of NA was inoculated in 4 types of media (each 200 mL), (1) M9 liquid medium containing

0.5% of YE and 0.1% of 5-phenylvaleric acid, (2) M9 liquid medium containing 0.5% of SG and 0.1% of 5-phenylvaleric acid, (3) M9 liquid medium containing 0.5% of BE and 0.1% of 5-phenylvaleric acid, and (4) M9  
5 liquid medium containing 0.5% of PP and 0.1% of 5-phenylvaleric acid, and cultured at 30°C for 24 hours. The cells were recovered from each medium by centrifugation, washed once with cold methanol, and lyophilized.

10 The lyophilized pellets from each medium were weighed and suspended in 100 mL of chloroform, and stirred at 55°C for 20 hours, to extract the PHA. Each extract solution was filtered through a membrane having a pore size of 0.45  $\mu$ m and concentrated by a  
15 evaporator. The concentrated solution was reprecipitated in cold methanol, to obtain the polymer, which was dried under a vacuum at room temperature and weighed. The yield results are given in Table 32.

The PHA composition thus prepared was analyzed in  
20 a manner similar to that in EXAMPLE 26. The PHA monomer units were found to comprise 97% of 3-hydroxy-5-phenylvaleric acid unit and 3% of 3-hydroxybutyric acid unit.

The results indicate that one of the embodiments  
25 of the present invention gives the PHA containing a very high proportion of the 3-hydroxy-5-phenylvaleric acid unit in a high yield.

[Example 31]

Pseudomonas cichorii strain YN2 was shake-cultured in 10 mL of M9 medium containing 0.5% of D-glucose under the conditions of 30°C and 125 strokes/min for 72 hours, and 2 mL of the culture was transferred for further shake-culture in 200 mL of M9 medium containing 0.5% of D-glucose and 0.1% of NO<sub>2</sub>PxBA under the conditions of 30°C and 125 strokes/min for another 72 hours. The cells were recovered by centrifugation, and resuspended in 200 mL of M9 medium, containing 0.5% of D-glucose and 0.1% of NO<sub>2</sub>PxBA but no nitrogen source (NH<sub>4</sub>Cl), where they were further shake-cultured under the conditions of 30°C and 125 strokes/min for 48 hours. The cells were recovered by centrifugation, washed once with cold methanol, and lyophilized.

The lyophilized pellets were suspended in 20 mL of chloroform, and stirred at 60°C for 20 hours, to extract the PHA. The extract solution was filtered through a membrane having a pore size of 0.45 µm and concentrated by a rotary evaporator. The concentrated solution was reprecipitated in cold methanol, and the precipitate only was recovered and dried under a vacuum, to obtain the PHA.

The PHA thus prepared was analyzed by NMR under the following conditions:

<Analyzer>

FT-NMR: Bruker DPX400

Resonance frequency:  $^1\text{H}$ =400 MHz

<Analysis conditions>

Nuclide to be analyzed:  $^1\text{H}$

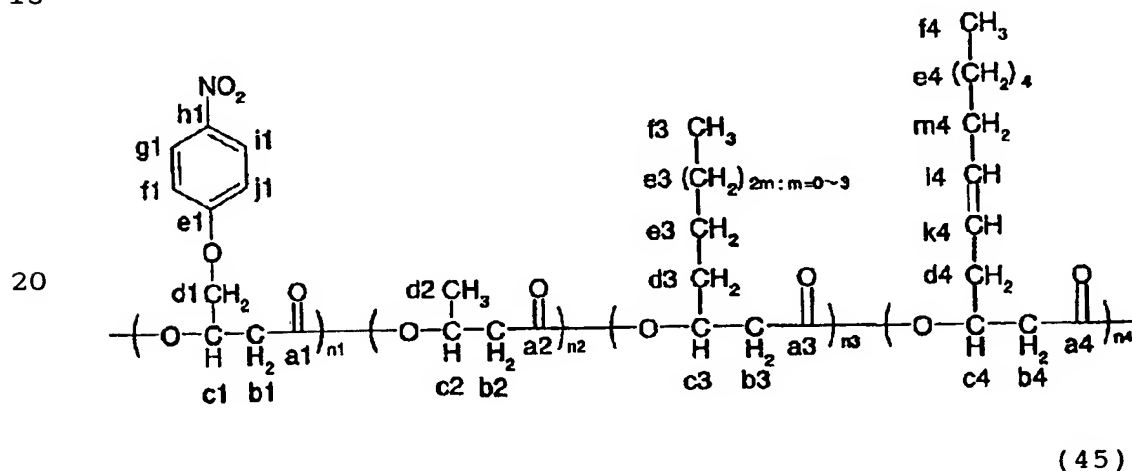
Solvent:  $\text{CDCl}_3$

5 Reference: TMS/ $\text{CDCl}_3$  sealed in a capillary

Temperature: room temperature

FIG. 14 shows the  $^1\text{H}$ -NMR spectral patterns, Table 33 the results of identification of the patterns, and Table 34 the composition of the monomer units of 3-hydroxy-4-(4-nitrophenoxy)butyric acid. As shown in Table 33, the PHA is the one represented by the chemical formula (45), containing 3-hydroxy-4-(4-nitrophenoxy)butyric acid as the monomer unit.

15



25

The PHA thus prepared was subjected to methanolysis by the normal procedure, and analyzed by a gas chromatograph/mass spectrometer (GC-MS, Shimadzu

QP-5050, based on the EI method), to identify the methyl-esterified product of the PHA monomer units, other than 3-hydroxy-4-(4-nitrophenoxy)butyric acid. The results are given in Table 34.

5           The PHA had a number-average molecular weight (Mn) of 81,900 and weight-average molecular weight (Mw) of 226,200, as determined by a gel permeation chromatograph (GPC; Toso HLC-8020, column: Polymer Laboratory's PLgel MIXED-C(5  $\mu$ m), solvent: chloroform,  
10 as polystyrene).

[Example 32]

Cells of *Pseudomonas cichorii* strain YN2 were inoculated into 200 mL of M9 culture medium containing 0.5% D-glucose and 0.1% NO<sub>2</sub>PxBA, and shake cultured at  
15 30°C at 125 stroke/min. After 45 hours, the cells were recovered by centrifugation and re-suspended into 200 mL of M9 culture medium containing 0.5% D-glucose, 0.1% NO<sub>2</sub>PxBA and nitrogen (NH<sub>4</sub>Cl)-free, and further shake  
cultured at 30°C at 125 stroke/min. After 48 hours,  
20 the cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

This lyophilized pellet was suspended into 20 mL of chloroform, and stirred at 60°C for 20 hours to extract PHA. After the extracts were filtrated through  
25 a membrane filter with a pore size of 0.45  $\mu$ m, they were concentrated by a rotating evaporator, and the concentrates were re-precipitated with cold methanol,

and further the precipitates alone were recovered and vacuum dried to provide PHA.

The resulting PHA was determined by NMR analysis under the condition shown in the Example 31. As a  
5 result, it was confirmed that the PHA was the PHA containing 3-hydroxy-4-(4-nitrophenoxy)butyric acid as a monomer unit, as shown in Table 35.

Further, the resulting PHA was done methanolysis according to conventional method, and then analyzed by  
10 gas chromatography-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit and identify monomer unit other than 3-hydroxy-4-(4-nitrophenoxy)butyric acid. The result was shown in the Table 35.

15 [Example 33]

Cells of *Pseudomonas cichorii* strain YN2 were inoculated into 200 mL of M9 culture medium containing 0.5% Polypeptone and 0.1%  $\text{NO}_2\text{PxBA}$ , and shake cultured at 30°C at 125 stroke/min. After 21 hours, the cells were  
20 recovered by centrifugation and re-suspended into 200 mL of M9 culture medium containing 0.5% sodium pyruvate, 0.1%  $\text{NO}_2\text{PxBA}$  and nitrogen ( $\text{NH}_4\text{Cl}$ )-free, and further shake cultured at 30°C at 125 stroke/min. After 24 hours, the cells were recovered by  
25 centrifugation, washed once with cold methanol and lyophilized.

This lyophilized pellet was suspended into 20 mL

of chloroform, and stirred at 60°C for 20 hours to extract PHA. After the extracts were filtrated through a membrane filter with a pore size of 0.45  $\mu$ m, they were concentrated by rotating evaporator, and the  
5 concentrates were re-precipitated with cold methanol, and further the precipitates alone were recovered and vacuum dried to provide PHA.

The resulting PHA was determined by NMR analysis under the condition shown in the Example 31. As a  
10 result, it was confirmed that the PHA was the PHA containing 3-hydroxy-4-(4-nitorophenoxy)butyric acid as a monomer unit, as shown in Table 36.

Further, the resulting PHA was done methanolysis according to the conventional method, and then analyzed  
15 by gas chromatography-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit and identify monomer unit other than 3-hydroxy-4-(4-nitrophenoxy)butyric acid. The result  
20 was shown in the Table 36.

[Example 34]

First, cells of *Pseudomonas cichorii* strain YN2 were inoculated into 10 mL of M9 culture medium containing 0.5% D-glucose and shake-cultured at 30°C at  
25 125 stroke/min, and then 2 mL of the culture was added into 200 mL of M9 culture medium containing 0.5% D-glucose and 0.1% CNPxBA, and shake cultured at 30°C



at 125 stroke/min. 48 hours later, the cells were recovered by centrifugation and re-suspended into 200 mL of M9 culture medium containing 0.5% D-glucose, 0.1% CNPxBA and nitrogen ( $\text{NH}_4\text{Cl}$ )-free, and further shake  
5 cultured at 30°C at 125 stroke/min. 47 hours later, the cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

This lyophilized pellet was suspended into 20 mL of chloroform, and stirred at 60°C for 20 hours to  
10 extract PHA. After the extracts were filtrated through a membrane filter with a pore size of 0.45  $\mu\text{m}$ , they were concentrated by rotating evaporator, and the concentrates were re-precipitated with cold methanol, and further the precipitates alone were recovered and  
15 vacuum dried to provide PHA.

The resulting PHA was determined by NMR analysis on the following conditions.

<Measuring apparatus>

FT-NMR: Bruker DPX 400

20 Resonance frequency:  $^1\text{H}$ =400 MHz

<Measuring condition>

Measuring nuclide:  $^1\text{H}$

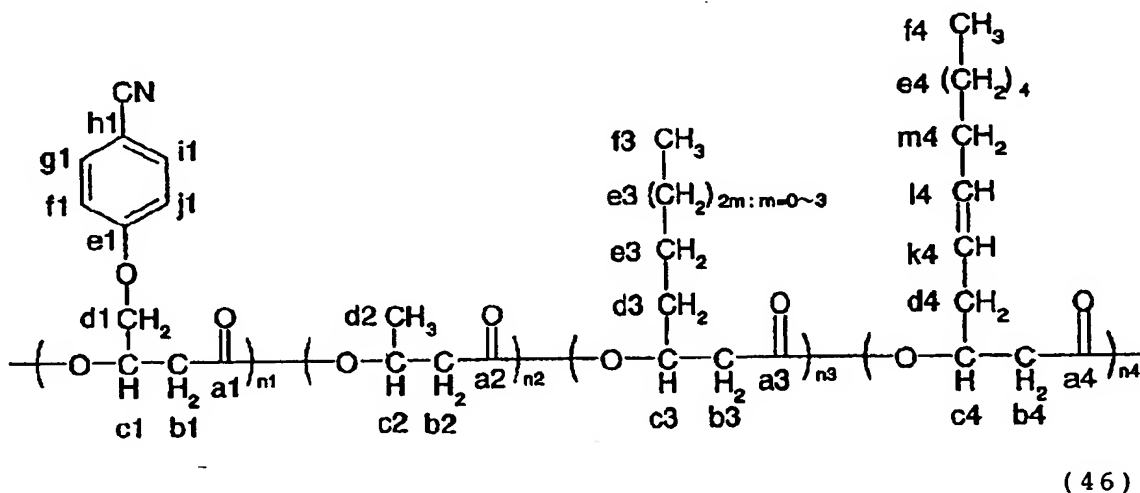
Measuring solvent:  $\text{CDCl}_3$

Reference: capillary inclusion TMS/ $\text{CDCl}_3$

25 Measuring temperature: room temperature

FIG. 15 illustrates  $^1\text{H}$ -NMR spectra, Table 37 shows their corresponding results, and Table 38 shows the

ratio of 3-hydroxy-4-(4-cyanophenoxy)butyric acid monomer unit contained in PHA. As shown in the Table 37, it was confirmed that the PHA was the PHA expressed by the chemical formula (46) containing 3-hydroxy-4-(4-cyanophenoxy)butyric acid as a monomer unit.



Further, the resulting PHA was done methanolysis according to the conventional method, and then analyzed by gas chromatography-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit and identify monomer unit other than 3-hydroxy-4-(4-cyanophenoxy)butyric acid. The result was shown in the Table 38.

Moreover, the molecular weight of the PHA was evaluated by gel permeation chromatography (GPC; Tosoh HLC-8020, column; Polymer Laboratory PL gel MIXED-C (5

containing 3-hydroxy-4-(4-cyanophenoxy)butyric acid as a monomer unit, as shown in Table 39.

Further, the resulting PHA was done methanolysis according to the conventional method, and then analyzed  
5 by gas chromatography-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit and identify monomer unit other than  
3-hydroxy-4-(4-cyanophenoxy)butyric acid. The result  
10 was shown in the Table 39.

[Example 36]

Cells of *Pseudomonas cichorii* strain YN2 were inoculated into 200 mL of M9 culture medium containing 0.5% D-glucose and 0.1% CNP×BA, and shake cultured at  
15 30°C at 125 stroke/min. After 48 hours, the cells were recovered by centrifugation and re-suspended into 200 mL of M9 culture medium containing 0.5% D-glucose, 0.1% CNP×BA and nitrogen (NH<sub>4</sub>Cl)-free, and further shake  
cultured at 30°C at 125 stroke/min. After 48 hours,  
20 the cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

This lyophilized pellet was suspended into 20 mL of chloroform, and stirred at 60°C for 20 hours to extract PHA. After the extracts were filtrated through  
25 a membrane filter with a pore size of 0.45 μm, they were concentrated by rotating evaporator, and the concentrates were re-precipitated with cold methanol,

and further the precipitates alone were recovered and vacuum dried to provide PHA.

The resulting PHA was determined by NMR analysis under the condition shown in the Example 34. As a  
5 result, it was confirmed that the PHA was the PHA containing 3-hydroxy-4-(4-cyanophenoxy)butyric acid as a monomer unit, as shown in Table 40.

Further, the resulting PHA was done methanolysis according to the conventional method, and then analyzed  
10 by gas chromatography-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit and identify monomer unit other than  
3-hydroxy-4-(4-cyanophenoxy)butyric acid. The result  
15 was shown in the Table 40.

[Example 37]

Cells of *Pseudomonas cichorii* strain H45 were inoculated into 200 mL of M9 culture medium containing 0.5% D-glucose and 0.1% CNPxBA, and shake cultured at  
20 30°C at 125 stroke/min. After 48 hours, the cells were recovered by centrifugation and re-suspended into 200 mL of M9 culture medium containing 0.5% D-glucose, 0.1% CNPxBA and nitrogen ( $\text{NH}_4\text{Cl}$ )-free, and further shake  
cultured at 30°C at 125 stroke/min. After 48 hours,  
25 the cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

This lyophilized pellet was suspended into 20 mL

of chloroform, and stirred at 60°C for 20 hours to extract PHA. After the extracts were filtrated through a membrane filter with a pore size of 0.45  $\mu$ m, they were concentrated by rotating evaporator, and the  
5 concentrates were re-precipitated with cold methanol, and further the precipitates alone were recovered and vacuum dried to provide PHA.

The resulting PHA was determined by NMR analysis under the condition shown in the Example 34. As a  
10 result, it was confirmed that the PHA was the PHA containing 3-hydroxy-4-(4-cyanophenoxy)butyric acid as a monomer unit, as shown in Table 41.

Further, the resulting PHA was done methanolysis according to the conventional method, and then analyzed  
15 by gas chromatography-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit and identify monomer unit other than 3-hydroxy-4-(4-cyanophenoxy)butyric acid. The result  
20 was shown in the Table 41.

[Example 38]

Cells of *Pseudomonas Cichorii* strain YN2 were inoculated into 200 mL of M9 culture medium containing 0.5% polypeptone and 0.1% CNPxBA and cultured at 30°C  
25 with shaking at 125 stroke/min. After 23 hours, the cells were recovered by centrifugation and re-suspended into 200 mL of M9 culture medium containing 0.5% sodium

pyruvate, 0.1% CNPxBA but no nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further cultured at 30°C at 125 stroke/min. After 24 hours, the cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

5           The lyophilized pellet was suspended into 20 mL of chloroform, and stirred at 60°C for 20 hours to extract PHA. After the extracts were filtrated through a membrane filter with a pore size of 0.45  $\mu\text{m}$ , they were concentrated by rotating evaporator, and the  
10 concentrates were re-precipitated with cold methanol, and further the precipitates alone were recovered and vacuum dried to provide PHA.

          The resulting PHA was determined by NMR analysis under the condition shown in the Example 34. As a  
15 result, it was confirmed that the PHA was the PHA containing 3-hydroxy-4-(4-cyanophenoxy)butyric acid as a monomer unit, as shown in Table 42.

          Further, the resulting PHA was done methanolysis according to the conventional method, and then analyzed  
20 by gas chromatography-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit and identify monomer unit other than  
3-hydroxy-4-(4-cyanophenoxy)butyric acid. The result  
25 was shown in the Table 42.

[Example 39]

Cells of *Pseudomonas cichorii* strain H45 were inoculated into 200 mL of M9 culture medium containing 0.5% polypeptone and 0.1% CNPxBA, and cultured at 30°C with shaking at 125 stroke/min. After 23 hours, the  
5 cells were recovered by centrifugation and re-suspended into 200 mL of M9 culture medium containing 0.5% Sodium pyruvate, 0.1% CNPxBA but no nitrogen source ( $\text{NH}_4\text{Cl}$ ), and further shake cultured at 30°C at 125 stroke/min. After 24 hours, the cells were recovered by  
10 centrifugation, washed once with cold methanol and lyophilized.

This lyophilized pellet was suspended into 20 mL of chloroform, and stirred at 60°C for 20 hours to extract PHA. After the extracts were filtrated through  
15 a membrane filter with a pore size of 0.45  $\mu\text{m}$ , they were concentrated by rotating evaporator, and the concentrates were re-precipitated with cold methanol, and further the precipitates alone were recovered and vacuum dried to provide PHA.

20 The resulting PHA was determined by NMR analysis under the condition shown in the Example 34. As a result, it was confirmed that the PHA was the PHA containing 3-hydroxy-4-(4-cyanophenoxy)butyric acid as a monomer unit, as shown in Table 43.

25 Further, the resulting PHA was done methanolysis according to the conventional method, and then analyzed by gas chromatography-mass spectrometer (GC-MS,

Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit and identify monomer unit other than 3-hydroxy-4-(4-cyanophenoxy)butyric acid. The result was shown in the Table 43.

[Example 40]

(Synthesis of 4-(4-fluorophenoxy)butyric acid)

A new compound, 4-(4-fluorophenoxy)butyric acid was prepared by the synthetic method mentioned below.

Into a round-bottom flask with four opening was put 240 mL of dehydrated acetone, added 15.2 g (0.11 mol) of potassium carbonate, and stirred under nitrogen atmosphere. Into this solution were added 9.0 g (0.06 mol) of sodium iodide and 7.9 g (0.07 mol) of 4-fluorophenol, and thoroughly stirred at room temperature under nitrogen atmosphere. 11.7 g (0.06 mol) of 4-bromoethyl butyrate was then added, and heat refluxed at 65°C for 24 hours.

After aforesaid reaction was completed, the solvent acetone was removed by rotating evaporator, and its residue was re-dissolved in chloroform. Water was added for phase separation, and the organic layer was collected. After the organic layer was dehydrated with anhydrous magnesium sulfate, chloroform was removed by rotating evaporator. Then, it was dried by a vacuum pump to provide 14.0 g of crude 4-(4-fluorophenoxy)ethyl butyrate (gas



chromatograph-mass spectrometer: GC-MS Shimadzu QP-5050, GC-MS peak ratio purity: 65.2% with EI method). Without purifying the resulting crude 4-(4-fluorophenoxy)ethyl butyrate, it was used in the following ester hydrolytic reaction.

The resulting crude 4-(4-fluorophenoxy)ethyl butyrate was dissolved in 300 mL of ethanol-water (1:9 (V/V)) mixed solution, and approximately ten-fold molar equivalent weight of potassium hydroxide was added to react for four hours under ice cooling (at 0 to 4°C). This cocktail was poured into 3 L of 0.1 mol/L hydrochloric acid to precipitate. Precipitates were filtrated, separated, and dried by vacuum pump to provide crude 4-(4-fluorophenoxy)butyric acid.

The resulting crude 4-(4-fluorophenoxy)butyric acid (precipitate) was dissolved in a small amount of hot methanol, and gradually cooled to recrystallize. Filtrated recrystallization materials were dried by vacuum pump to provide objective compound, 4-(4-fluorophenoxy)butyric acid. In this series of processes, the resulting 4-(4-fluorophenoxy)butyric acid was 52.7% in yield based on the raw material 4-bromoethylbutyrate.

In order to verify that the resulting compound was the objective 4-(4-fluorophenoxy)butyric acid, NMR analysis was carried out by the following measuring apparatus under measuring condition to identify the

structure.

<Measuring apparatus>

FT-NMR: Bruker DPX 400

<Measuring condition>

5           Resonance frequency:  $^1\text{H}$  400 MHz

$^{13}\text{C}$  100 MHz

Measuring nuclide:  $^1\text{H}$ ,  $^{13}\text{C}$

Used solvent :  $\text{CDCl}_3$

Reference: capillary inclusion TMS/ $\text{CDCl}_3$

10           Measuring temperature: room temperature

Determined  $^1\text{H}$ -NMR spectra chart and  $^{13}\text{C}$ -NMR spectra chart were illustrated in FIG. 16 and 17, respectively. Table 44 and 45 show analytical (corresponding) result of each signal for NMR spectra illustrated in the FIG.

15           16 and 17. This analytical (corresponding) result confirmed that the resulting compound was the objective 4-(4-fluorophenoxy)butyric acid.

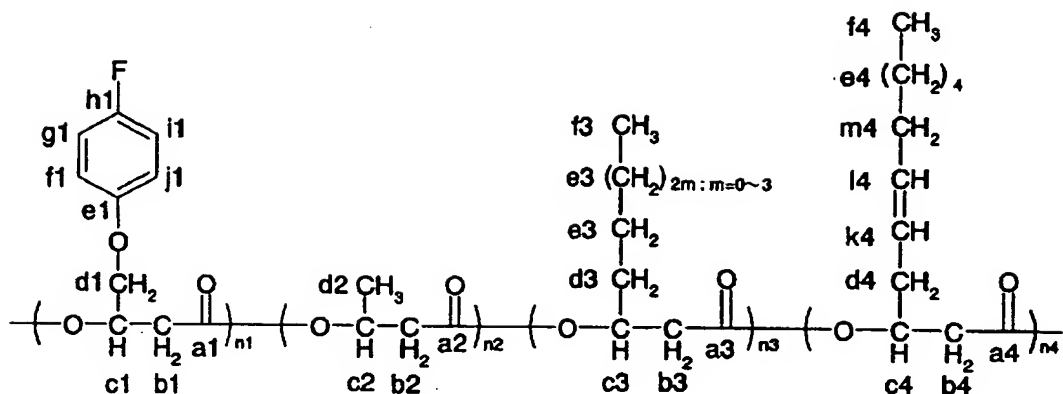
[Example 41]

First, cells of *Pseudomonas cichorii* strain YN2  
20           (FERM BP-7375) were inoculated into 10 mL of M9 culture medium containing 0.5% D-glucose, and shake cultured at 30°C at 125 stroke/min for 72 hours. Then 2 mL of the culture was added into 200 mL of M9 culture medium (no inorganic nitrogen source,  $\text{NH}_4\text{Cl}$ ) containing 0.5%  
25           D-glucose and 0.1% pFPxBA, and continuously shake cultured at 125 stroke/min. After 45 hours, the cells were recovered by centrifugation and re-suspended into

200 mL of M9 culture medium containing 0.5% D-glucose and 0.1% pFPxBA, and further shake cultured at 30°C at 125 stroke/min. After 46 hours, the cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

This lyophilized pellet was suspended into 20 mL of chloroform, and stirred at 60°C for 20 hours to extract PHA. After the extracts were filtrated through a membrane filter with a pore size of 0.45  $\mu$ m, they were concentrated by rotating evaporator, and the concentrates were re-precipitated with cold methanol, and further the precipitates alone were recovered and vacuum dried to provide PHA.

The resulting PHA was done methanolysis according to the conventional method, and then analyzed by gas chromatography-mass spectrometer (GC-MS, Shimadzu QP-5050, EI method) to identify methylesterified materials of PHA monomer unit. FIG. 18 illustrates measured GC-MS spectra data, and the upper chart shows GC spectrum, and the lower chart shows an MS spectrum corresponding to the main peak on the GC spectrum. This result shows that the resulting PHA contains 3-hydroxy-4-(4-fluorophenoxy)butyric acid (3HpFPxB) as a main component of monomer unit, in addition it also contains a small amount of six kinds of monomer units, and can be represented by the following chemical formula (47).



10

(47)

Furthermore, the molecular weight of the PHA was determined by gel permeation chromatography (GPC; Toso·HLC-8020, column; Polymer laboratory·PL gel·MIX ED-C·5  $\mu$ m, solvent; chloroform, polystyrene reduced molecular weight).

Identification result, average molecular weight, as well as yield of lyophilized pellet and recovered polymer are shown in Table 46. It shows that the resulting PHA is the PHA containing 3-hydroxy-4-(4-fluorophenoxy)butyric acid (3HpFPxB) as a monomer unit. Further, it is likely that the extracted PHA includes 3HpFPxB unit as a major component, however, it is the mixture containing more than one kind of component as a monomer unit selected from the group consisting of 3-hydroxybutyric acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid, 3-hydroxydecanoic acid, 3-hydroxydodecanoic acid,

3-hydroxydodecenoic acid. Estimated average molecular weight was  $M_n=42400$  for number-average molecular weight, on the other hand,  $M_w=90600$  for weight-average molecular weight.

5            This PHA was also determined by NMR analysis with the same measuring apparatus and same measuring condition as shown in the Example 40. The measured  $^1\text{H}$ -NMR spectra chart is shown in FIG. 19. Table 47 shows the analytical (corresponding) result of each  
10           signal for major peak for NMR spectra illustrated in the FIG. 19. This analytical (corresponding) result confirmed that the resulting PHA contained 3HpFPxB unit as a major component.

[Example 42]

15           Cells of *Pseudomonas cichorii* H45; FERM BP-7374 were inoculated in 10 mL of M9 medium containing D-glucose 0.5%, and was shake-cultured at 30°C in 125 strokes/min. for 72 hours. Then 2 ml of the culture was transferred into 200 ml of M9 medium containing  
20           0.5% D-glucose and 0.1% pFPxBA, and further shake cultured at 30°C in 125 strokes/min. After 45 hours, cultured cells were recovered by centrifugation, and re-suspended in M9 medium containing D-glucose 0.5% and pFPxBA 0.1%, but not containing inorganic nitrogen  
25           source ( $\text{NH}_4\text{Cl}$ ), and further cultured at 30°C in 125 strokes/min. After 46 hours, cells were recovered by centrifugation, washed once with cold methanol and

lyophilized.

The lyophilized pellet was suspended in chloroform 20 mL, and stirred at 60°C for 20 hours to extract PHA. The extract was filtered using membrane filter, pore size 0.45 µm, concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

The thus obtained PHA was subjected to methanolysis, and the product was analyzed using gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA monomer units. In Table 48, result of identification, and, weight obtained and yield of the lyophilized pellet and the recovered polymer are shown. The thus obtained PHA is the PHA with 3HpFPxB as a monomer unit. The extracted PHA mainly consists of 3HpFPxB as a main component, and is considered to be a mixture containing more than one of compounds selected from the group consisting of 3-hydroxybutyric acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid, 3-hydroxydecanoic acid, 3-hydroxydodecanoic acid and 3-hydroxydodecenoic acid, as a monomer unit.

[Example 43]

Cells of *Pseudomonas cichorii* YN2 were inoculated in 200 ml M9 medium containing D-glucose 0.5% and pFPxBA 0.1% and was shake cultured at 30°C in 125

strokes/min. After 96 hours, the cultured cells were recovered by centrifugation, and re-suspended in 200 ml of M9 medium containing D-glucose 0.5% and pFPxBA 0.1% but not containing an inorganic nitrogen source ( $\text{NH}_4\text{Cl}$ ),  
5 and further shake cultured at 30°C in 125 strokes/min. After 64 hours, cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in chloroform 20 mL, and stirred at 60°C for 20 hours to extract PHA.  
10 The extract was filtered using membrane filter, pore size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

15 The thus obtained PHA was subjected to methanolysis, and the product was analyzed using gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA monomer units. In Table 49, result of identification,  
20 and weight obtained and yield of the lyophilized pellet and the recovered polymer are shown. The thus obtained PHA is the PHA with 3HpFPxB as a major monomer unit.

[Example 44]

Cells of *Pseudomonas cichorii* H45 were inoculated  
25 in M9 medium 200 mL containing D-glucose 0.5% and pFPxBA 0.1% and was shake cultured at 30°C in 125 strokes/min. After 96 hours, cultured cells were

recovered by centrifugation, and re-suspended in M9 medium not containing inorganic nitrogen source  $\text{NH}_4\text{Cl}$ , containing D-glucose 0.5% and pFPxBA 0.1% (200 ml), then further shake cultured at 30°C in 125 strokes/min. After 64 hours, cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in chloroform 20 mL, and stirred at 60°C for 20 hours to extract PHA. The extract was filtered using membrane filter of pore size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

The thus obtained PHA was subjected to methanolysis, and the product was analyzed using gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA monomer units. In Table 50, result of identification, and, weight obtained and yield of the lyophilized pellet and the recovered polymer are shown. The thus obtained PHA is the PHA with 3HpFPxB as a major monomer unit.

[Example 45]

Cells of *Pseudomonas cichorii* YN2 were inoculated in 200 ml of M9 medium containing polypeptone 0.5% and pFPxBA 0.1% and was shake cultured at 30°C in 125 strokes/min. After 24 hours, cultured cells were



recovered by centrifugation, and re-suspended in 200 ml of M9 medium containing sodium pyruvate 0.5% and pFPxBA 0.1%, but not containing inorganic nitrogen source  $\text{NH}_4\text{Cl}$ , and further shake cultured at 30°C in 125  
5 strokes/min. After 24 hours, cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in chloroform 20 mL, and stirred at 60°C for 20 hours to extract PHA.  
10 The extract was filtered using membrane filter (pore size 0.45  $\mu\text{m}$ ), concentrated by rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

15 The thus obtained PHA was subjected to methanolysis, and the product was analyzed using gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA monomer units. In Table 51, result of identification,  
20 weight obtained and yield of the lyophilized pellet and the recovered polymer are shown. The thus obtained PHA is the PHA with 3HpFPxB as a major monomer unit.

[Example 46]

Cells of *Pseudomonas cichorii* H45 were inoculated  
25 in 200 ml of M9 medium containing polypeptone 0.5% and pFPxBA 0.1% and was shake cultured at 30°C in 125 strokes/min. After 24 hours, the cells were recovered

by centrifugation, and re-suspended in 200 ml of M9 medium not containing inorganic nitrogen source  $\text{NH}_4\text{Cl}$ , and containing sodium pyruvate 0.5% and pFPxBA 0.1%, then further shake cultured at 30°C in 125 strokes/min.

5 After 24 hours, the cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in chloroform 20 mL, and stirred at 60°C for 20 hours to extract PHA.

10 The extract was filtered using membrane filter, pore size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

15 The thus obtained PHA was subjected to methanolysis, and the product was analyzed using gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA monomer units. In Table 52, result of identification, and, weight obtained and yield of the lyophilized  
20 pellet and the recovered polymer are shown. The thus obtained PHA is the PHA with 3HpFPxB as a major monomer unit.

[Example 47]

25 (Synthesis of 4-(3-fluorophenoxy)butyric acid)

Novel compound 4-(3-fluorophenoxy)butyric acid was prepared by the following synthetic method.

Dehydrated acetone 240 mL was poured in a four-neck round-bottom flask, and potassium carbonate 15.2 g (0.11 mol) was added thereto, then stirred under nitrogen atmosphere. Sodium iodide 9.0 g (0.06 mol) and 3-fluorophenol 7.9 g (0.07 mol) were added to the solution, and the mixture was stirred sufficiently at room temperature under nitrogen atmosphere. Then ethyl 4-bromobutyrate 11.7 g (0.06 mol) and refluxed at 65°C for 24 hours.

After completed the above reaction, solvent acetone was distilled off using rotary evaporator, and the residue was dissolved in chloroform. Water was added therein for separation and the organic layer was collected. The organic layer was dehydrated by adding anhydrous magnesium sulfate and the chloroform was distilled off by using rotary evaporator. Further, the residue was dried in vacuo using vacuum pump to obtain crude ethyl 4-(3-fluorophenoxy) butyrate 14 g (purification 87.9%, GC-MS peak ratio, determined by EI method using gas chromatography-mass spectrograph: GC-MS Shimadzu QP-5050). The thus obtained crude ethyl 4-(3-fluorophenoxy) butyrate was used subsequent ester hydrolysis without purification.

The thus obtained crude ester 3.0 g was dissolved in a mixture of ethanol-water [1:9 (V/V)] 100 mL, to which about 10-fold excess molar equivalent of potassium hydroxide was added, and the mixture was

reacted at room temperature for 4 hours. The reaction mixture was poured into about 200 mL of 0.1 mol/l aqueous hydrochloric acid to precipitate. The precipitate was filtered, separated and dried in vacuo  
5 under the vacuum pump to obtain crude 4-(3-fluorophenoxy) butyric acid.

The thus obtained crude 4-(3-fluorophenoxy) butyric acid (precipitate) was dissolved in a small amount of hot methanol and gradually cooled for  
10 recrystallization. Filtered recrystallized product was dried using vacuum pump to obtain 4-(3-fluorophenoxy) butyric acid.

The yield of 4-(3-fluorophenoxy) butyric acid obtained from crude ester 3.0 g was 2.4 g.  
15 Consequently, total yield in the whole process based on the raw material ethyl 4-bromobutyrate is 93.2%.

For verification of the objective compound. 4-(3-fluorophenoxy) butyric acid, structure of the obtained compound was identified by NMR using following  
20 measuring apparatus and conditions.

<Measuring apparatus>

FT-NMR: Bruker DPX400

<Measuring condition>

Resonance frequency:  $^1\text{H}$  400 MHz

25  $^{13}\text{C}$  100 MHz

Measuring nuclide:  $^1\text{H}$  and  $^{13}\text{C}$

Solvent used:  $\text{CDCl}_3$

Reference: Capillary sealed TMS/ $\text{CDCl}_3$ ,

Measuring temperature: room temperature

$^1\text{H}$ -NMR spectrum and  $^{13}\text{C}$ -NMR spectrum are shown in  
FIG. 20 and FIG. 21, respectively. In Table 53 and  
5 Table 54, results of analyses (assignments) of  
respective signals of NMR spectra, which are shown in  
FIG. 20 and FIG. 21, are shown. According to the  
result of analysis (assignment), the obtained compound  
is confirmed to be the objective compound  
10 4-(3-fluorophenoxy) butyric acid.

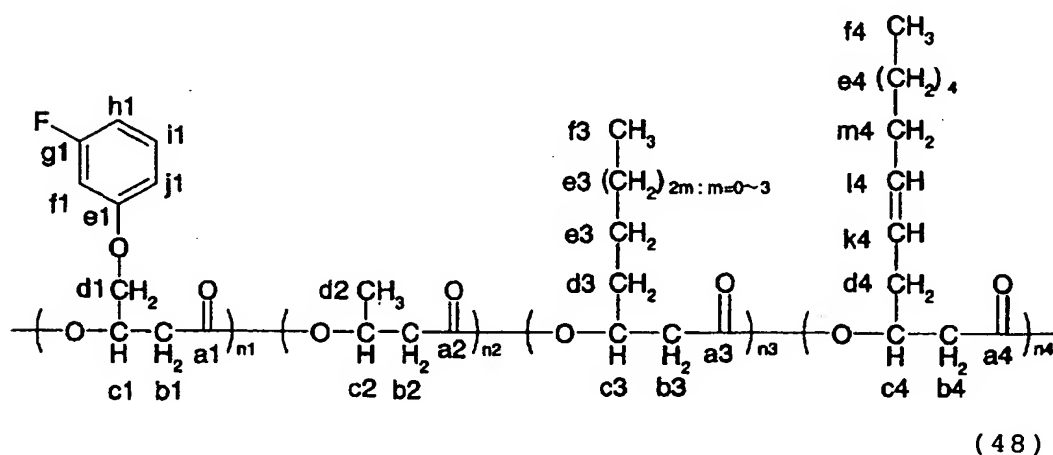
[Example 48]

Cells of *Pseudomonas cichorii* YN2; FERM BP-7375  
were inoculated in 10 mL of M9 medium containing  
D-glucose 0.5% and shake cultured at 30°C at 125  
15 strokes/min for 72 hours. Then 2 mL of the culture was  
transferred into 200 ml of M9 medium containing  
D-glucose 0.5% and mFPxBA 0.1% and shake cultured at  
125 strokes/min. After 45 hours, the cells were  
recovered by centrifugation, and re-suspended in 200 ml  
20 of M9 medium containing D-glucose 0.5% and mFPxBA 0.1%  
but not containing inorganic nitrogen source  $\text{NH}_4\text{Cl}$ , and  
further shake cultured at 30°C at 125 strokes/min.  
After 47 hours, cells were recovered by centrifugation,  
washed once with cold methanol and lyophilized.

25 The lyophilized pellet was suspended in chloroform  
20 mL, and stirred at 60°C for 20 hours to extract PHA.  
The extract was filtered using membrane filter, pore

size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

5           The thus obtained PHA was subjected to methanolysis, and the product was analyzed using gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA monomer units. In FIG. 22, measured GC-MS spectrum data is shown. Upper part of FIG. 22 indicates GC spectrum, and the lower part indicates MS spectrum corresponding to main peaks on the above GC spectrum. As a result, the thus obtained PHA is the PHA, which contains 3-hydroxy-4-(3-fluorophenoxy) butyric acid (3HmFPxB) as a major monomer unit as well as small amount of 6 types of monomer units, and the compound can be illustrated as the following chemical structure (48).



Molecular weight of PHA was measured by using gel-permeation chromatography (GPC: Toso HLC-8020; Column: Polymer Laboratory PLgel MIX ED-C, 5  $\mu$ m; Solvent: chloroform: Molecular weight: reduced value  
5 for polystyrene).

In Table 55, result of identification, average molecular weight, and weight obtained and yield of the lyophilized pellet and the recovered polymer are shown. The thus obtained PHA is the PHA, which contains  
10 3-hydroxy-4-(3-fluorophenoxy) butyric acid (3HmFPxB) as a monomer unit. The extracted PHA mainly consists of 3HmFPxB unit as a main component, and is considered to be a mixture containing more than one of compounds selected from the group consisting of 3-hydroxybutyric  
15 acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid, 3-hydroxydecanoic acid, 3-hydroxydodecanoic acid and 3-hydroxydodecenoic acid, as a monomer unit. The evaluated average molecular weight is: number average molecular weight  $M_n=34500$  and weight-average molecular  
20 weight  $M_w=75200$ .

NMR analysis of the PHA was performed by the same procedures using same measuring apparatus and measuring conditions as described in example 47. Measured  $^1\text{H}$ -NMR spectrum is shown in FIG. 23. In Table 56, result of  
25 analysis (assignment) of major peak signals of NMR spectrum in FIG. 23 is shown. According to the result of analysis (assignment), the obtained compound is

confirmed to have 3HmFPxB unit as a major component.

[Example 49]

Cells of *Pseudomonas cichorii* H45; FERM BP-7374 were inoculated in 10 mL of M9 medium containing  
5 D-glucose 0.5% and was cultured at 30°C with shaking at 125 strokes/min for 72 hours. Then 2 mL of the culture was transferred into 200 mL of M9 medium containing D-glucose 0.5% and mFPxBA 0.1% and shake cultured at 30°C at 125 strokes/min. After 45 hours, the cells  
10 were recovered by centrifugation, and were re-suspended in 200 mL of M9 medium containing D-glucose 0.5% and mFPxBA 0.1% but not containing inorganic nitrogen source ( $\text{NH}_4\text{Cl}$ ), then further shake cultured at 30°C in 125 strokes/min. After 47 hours, cells were recovered  
15 by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in chloroform 20 mL, and stirred at 60°C for 20 hours to extract PHA. The extract was filtered using membrane filter, pore  
20 size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

The thus obtained PHA was subjected to  
25 methanolysis, and the product was analyzed using gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA



monomer units. In Table 57, result of identification, and weight obtained and yield of the lyophilized pellet and the recovered polymer are shown. The thus obtained PHA is the PHA, which contains 3HmFPxB as a monomer unit. The extracted PHA mainly consists of 3HmFPxB unit as a main component, and is considered to be a mixture containing more than one of compounds selected from the group consisting of 3-hydroxybutyric acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid, 3-hydroxydecanoic acid, and 3-hydroxydodecanoic acid, as a monomer unit.

[Example 50]

*Pseudomonas cichorii* H45 was inoculated in 200 ml of M9 medium containing D-glucose 0.5% and mFPxBA 0.1% and was cultured at 30°C with shaking at 125 strokes/min. After 96 hours, cultured cells were recovered by centrifugation, and re-suspended in M9 medium containing D-glucose 0.5% and mFPxBA 0.1% but not containing the inorganic nitrogen source  $\text{NH}_4\text{Cl}$ , then further shake cultured at 30°C, 125 strokes/min. After 64 hours, cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in chloroform 20 mL, and stirred at 60°C for 20 hours to extract PHA. The extract was filtered using membrane filter, pore size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and

recovered the precipitate to obtain PHA by vacuum drying.

The thus obtained PHA was subjected to methanolysis, and the product was analyzed using  
5 gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA monomer units. In Table 58, result of identification, and weight obtained and yield of the lyophilized pellet and the recovered polymer are shown. The thus obtained  
10 PHA is the PHA containing 3HmFPxB as a major monomer unit.

[Example 51]

*Pseudomonas cichorii* YN2 was inoculated in 200 mL of M9 medium containing polypeptone 0.5% and mFPxBA  
15 0.1% and was cultured at 30°C with shaking at 125 strokes/min. After 24 hours, cultured cells were recovered by centrifugation, and re-suspended in 200 ml of M9 medium containing sodium pyruvate 0.5% and mFPxBA 0.1% but not containing the inorganic nitrogen source  
20 NH<sub>4</sub>Cl, then further shake cultured at 30°C, 125 strokes/min. After 24 hours, bacterial cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in chloroform  
25 20 mL, and stirred at 60°C for 20 hours to extract PHA. The extract was filtered using membrane filter, pore size 0.45 µm, concentrated using rotary evaporator,

precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

The thus obtained PHA was subjected to  
5 methanolysis, and the product was analyzed using  
gas-chromatography mass spectrograph (GC-MS, Shimadzu  
QP-5050, EI method) to identify methyl esterified PHA  
monomer units. In Table 59, result of identification,  
and weight obtained and yield of the lyophilized pellet  
10 and the recovered polymer are shown. The thus obtained  
PHA is the PHA containing 3HmFPxB as a major monomer  
unit.

[Example 52]

*Pseudomonas cichorii* H45 was inoculated in 200 mL  
15 of M9 medium containing polypeptone 0.5% and mFPxBA  
0.1% and was cultured at 30°C with shaking at 125  
strokes/min. After 24 hours, cultured cells were  
recovered by centrifugation, and were re-suspended in  
200 mL of M9 medium containing sodium pyruvate 0.5% and  
20 mFPxBA 0.1% but not containing inorganic nitrogen  
source  $\text{NH}_4\text{Cl}$ , then further shake cultured at 30°C, 125  
strokes/min. After 24 hours, the cells were recovered  
by centrifugation, washed once with cold methanol and  
lyophilized.

25 The lyophilized pellet was suspended in chloroform  
20 mL, and stirred at 60°C for 20 hours to extract PHA.  
The extract was filtered using membrane filter, pore

size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

5           The thus obtained PHA was subjected to methanolysis, and the product was analyzed using gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, EI method) to identify methyl esterified PHA monomer units. In Table 60, result of identification,  
10           and weight obtained and yield of the lyophilized pellet and the recovered polymer are shown. The thus obtained PHA is the PHA containing 3HmFPxB as a major monomer unit.

[Example 53]

15           <Production of PHA containing HFPxV unit by strain YN2 (one step culture using polypeptone)>

          The strain YN2 was inoculated in 200 mL of M9 medium containing polypeptone (Wako Pure Chemicals Co.) 0.5% and 5-(4-fluorophenoxy) valeric acid (FPxVA) 0.1%  
20           and was cultured at 30°C with shaking at 125 strokes/min. After 27 hours, cells were recovered by centrifugation, washed once with cold methanol, lyophilized and were weighed.

          The lyophilized pellet was suspended in chloroform  
25           100 mL, and stirred at 60°C for 20 hours to extract polymer. The extract was filtered using membrane filter, pore size 0.45  $\mu\text{m}$ , concentrated using rotary

evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain the polymer by vacuum drying, then weighed as such.

Molecular weight of the obtained polymer was  
5 measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C 5  $\mu$ m; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was  
10 analyzed by the following manner. The polymer sample 5 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at 100°C for 3.5 hours, further added water thereto for  
15 separation. The organic layer was analyzed by means of gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of bacterial cells and polymer, molecular weight and  
20 analytical result of the monomer unit are shown in Table 61. Ratio of monomer unit was calculated by area ratio of GC-MS total ion chromatogram (TIC). Mass spectra obtained by GC-MS of 3-hydroxybutyric acid methyl ester and 3-hydroxy-5-(4-fluorophenoxy) valeric  
25 acid methyl ester are shown in FIG. 24 and FIG. 25.

Result indicates that PHA copolymer containing 3-hydroxy-5-(4-fluorophenoxy) valeric acid unit can be

produced by the strain YN2 with a substrate  
5-(4-fluorophenoxy) valeric acid.

[Example 54]

<Production of PHA containing HFPV unit by strain YN2  
5 (one step culture using polypeptone)>

The strain YN2 was inoculated in 200 mL of M9  
medium containing polypeptone (Wako Pure Chemicals Co.)  
0.5% and 5-(4-fluorophenyl) valeric acid (FPVA) 0.1%  
and was cultured at 30°C with shaking at 125  
10 strokes/min. After 27 hours, the cells were recovered  
by centrifugation, washed once with cold methanol,  
lyophilized and weighed.

The lyophilized pellet was suspended in chloroform  
100 mL, and stirred at 60°C for 20 hours to extract  
15 polymer. The extract was filtered using membrane  
filter, pore size 0.45  $\mu$ m, concentrated using rotary  
evaporator, precipitated the concentrate with cold  
methanol, and recovered the precipitate to obtain the  
polymer by vacuum drying, then weighed as such.

20 Molecular weight of the obtained polymer was  
measured by means of gel-permeation chromatography  
(GPC: Toso, HLC-8020; Column: Polymer Laboratory,  
PL-gel, MIXED-C 5  $\mu$ m; Solvent: chloroform; Molecular  
weight: reduced value for polystyrene).

25 Unit composition of the obtained polymer was  
analyzed by the following manner. The polymer sample 5  
mg was poured into the 25 mL round-neck flask, and

chloroform 2 mL and 2 mL of methanol containing  
sulfuric acid (3%, v/v). The mixture was refluxed at  
100°C for 3.5 hours, further added water thereto for  
separation. The organic layer was analyzed by means of  
5 gas-chromatography mass spectrograph (GC-MS, Shimadzu  
QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to  
identify methyl esterified PHA monomer unit. Yields of  
bacterial cells and polymer, molecular weight and  
analytical result of the monomer unit are shown in  
10 Table 62. Ratio of monomer unit was calculated by area  
ratio of GC-MS total ion chromatogram (TIC). Mass  
spectra obtained by GC-MS of 3-hydroxybutyric acid  
methyl ester and 3-hydroxy-5-(4-fluorophenyl) valeric  
acid methyl ester are shown in FIG. 26 and FIG. 27.

15        Result indicates that PHA copolymer containing  
3-hydroxy-5-(4-fluorophenyl) valeric acid unit can be  
produced by the strain YN2 with a substrate  
5-(4-fluorophenyl) valeric acid.

[Example 55]

20        <Production of PHA containing HFPxV unit by strain YN2  
(two step culture using glucose)>

      The strain YN2 was inoculated in 200 mL of M9  
medium containing glucose 0.5% and 5-(4-fluorophenoxy)  
valeric acid (FPxVA) 0.1% and was cultured at 30°C with  
25 shaking at 125 strokes/min. After 24 hours, the  
cultured cells were recovered by centrifugation, and  
were re-suspended in 200 mL of M9 medium containing

glucose 0.5% and FPxBA 0.1% but not containing nitrogen source ( $\text{NH}_4\text{Cl}$ ), then further shake cultured at  $30^\circ\text{C}$ , 125 strokes/min. After 62 hours, the cells were recovered by centrifugation, washed once with cold methanol and  
5 lyophilized.

The lyophilized pellet was suspended in chloroform 100 mL, and stirred at  $60^\circ\text{C}$  for 20 hours to extract polymer. The extract was filtered using membrane filter, pore size  $0.45\ \mu\text{m}$ , concentrated using rotary  
10 evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA by vacuum drying.

Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography  
15 (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C  $5\ \mu\text{m}$ ; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5  
20 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at  $100^\circ\text{C}$  for 3.5 hours, further added water thereto for separation. The organic layer was analyzed by means of  
25 gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of



bacterial cells and polymer, and analytical result of the monomer unit are shown in Table 63. Mass spectra obtained by GC-MS of 3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic acid methyl ester and  
5 3-hydroxy-5-(4-fluorophenoxy) valeric acid (3HFPxV) methyl ester obtained by GC-MS measurement are shown in FIG. 28 to FIG. 30.

Result indicates that PHA copolymer containing 3-hydroxy-5-(4-fluorophenoxy) valeric acid (3HFPxV)  
10 unit can be produced by the strain YN2 with a substrate 5-(4-fluorophenoxy) valeric acid.

[Example 56]

<Production of PHA containing HFPV unit by strain YN2 (two step culture using glucose)>

15 The strain YN2 was inoculated in 200 mL of M9 medium containing glucose 0.5% and 5-(4-fluorophenyl) valeric acid (FPVA) 0.1% and was cultured at 30°C with shaking at 125 strokes/min. After 24 hours, the cultured cells were recovered by centrifugation, and  
20 were re-suspended in 200 mL of M9 medium containing glucose 0.5% and FPxVA 0.1% but not containing the nitrogen source ( $\text{NH}_4\text{Cl}$ ), then further shake cultured at 30°C in 125 strokes/min. After 62 hours, the cells were recovered by centrifugation, washed once with cold  
25 methanol and lyophilized.

The lyophilized pellet was suspended in chloroform 100 mL, and stirred at 60°C for 20 hours to extract

polymer. The extract was filtered using membrane filter, pore size 0.45  $\mu$ m, concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA  
5 by vacuum drying.

Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C 5  $\mu$ m; Solvent: chloroform; Molecular  
10 weight: reduced value for polystyrene).

Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing  
15 sulfuric acid (3%, v/v). The mixture was refluxed at 100°C for 3.5 hours, further added water thereto for separation. The organic layer was analyzed by means of gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to  
20 identify methyl esterified PHA monomer unit. Yields of bacterial cells and polymer, and analytical result of the monomer unit are shown in Table 64. Mass spectra obtained by GC-MS of 3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic acid methyl ester and  
25 3-hydroxy-5-(4-fluorophenyl) valeric acid (3HFPV) methyl ester obtained by GC-MS measurement are shown in FIG. 31 to FIG. 33.

Result indicates that PHA copolymer containing 3-hydroxy-5-(4-fluorophenyl) valeric acid (3HFPV) unit can be produced by the strain YN2 with a substrate 5-(4-fluorophenyl) valeric acid.

5 [Example 57]

<Production of PHA containing HFPV unit and HFPxV unit by strain YN2 (two step culture using glucose)>

Two M9 media containing glucose 0.5% and 5-(4-fluorophenyl) valeric acid (FPVA) 0.1%, and  
10 containing glucose 0.5% and 5-(4-fluorophenoxy) valeric acid (FPxVA) 0.1% were prepared respectively. Cells of strain YN2 were inoculated in 200 mL of each M9 medium, and cultured at 30°C with shaking at 125 strokes/min. After 94 hours, the cultured cells were recovered by  
15 centrifugation, and the cells of these two cultures were re-suspended together in 200 mL of M9 medium containing glucose 0.5%, FPVA 0.1% and FPxVA 0.1% but not containing nitrogen source ( $\text{NH}_4\text{Cl}$ ), then further shake cultured at 30°C, 125 strokes/min. After 24  
20 hours, the cells were recovered by centrifugation, washed once with cold methanol and lyophilized.

The lyophilized pellet was suspended in chloroform 100 mL, and stirred at 60°C for 20 hours to extract PHA. The extract was filtered using membrane filter,  
25 pore size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain PHA

by vacuum drying.

Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C 5  $\mu$ m; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at 100°C for 3.5 hours, further added water thereto for separation. The organic layer was analyzed by means of gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of bacterial cells and polymer, and analytical result of the monomer unit are shown in Table 65. Mass spectra obtained by GC-MS of 3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic acid methyl ester, 3-hydroxy-5-(4-fluorophenyl) valeric acid (3HFPV) methyl ester and 3-hydroxy-5-(4-fluorophenoxy) valeric acid (3HFPxV) methyl ester obtained by GC-MS measurement are shown in FIG. 34 to FIG. 37.

Results indicate that PHA copolymer containing 3-hydroxy-5-(4-fluorophenyl) valeric acid (3HFPV) unit and 3-hydroxy-5-(4-fluorophenoxy) valeric acid (3HFPxV)

unit can be produced by the strain YN2 with a substrate 5-(4-fluorophenyl) valeric acid and 5-(4-fluorophenoxy) valeric acid.

[Example 58]

5           <Production of PHA containing HPxN unit, HPxHp unit and HPxV unit by strain YN2 (one step culture using polypeptone)>

          The strain YN2 was inoculated in 200 mL of M9 medium containing polypeptone (Wako Pure Chemicals Co.) 10 0.5% and 11-phenoxyundecanoic acid (PxUDA) 0.1% and was cultured at 30°C with shaking 125 strokes/min. After 64 hours, the cells were recovered by centrifugation, washed once with cold methanol, lyophilized and weighed.

15           The lyophilized pellet was suspended in acetone 100 mL, and stirred at room temperature (23°C) for 72 hours to extract polymer. The extract was filtered using membrane filter, pore size 0.45 µm, concentrated using rotary evaporator, precipitated the concentrate 20 with cold methanol, and recovered the precipitate to obtain the polymer by vacuum drying, then weighed as such.

          Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography 25 (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C 5 µm; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at 100°C for 3.5 hours, further added water thereto for separation. The organic layer was analyzed by means of gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of bacterial cells and polymer, and analytical result of the monomer unit are shown in Table 66. Mass spectra obtained by GC-MS of 3-hydroxybutyric acid methyl ester, 3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic acid methyl ester, 3-hydroxy-5-phenoxyvaleric acid (3HPxV) methyl ester, 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) methyl ester and 3-hydroxy-9-phoxynonanoic acid (3HPxN) methyl ester are shown in FIG. 38 to FIG. 43.

Result indicates that PHA copolymer containing three units of 3-hydroxy-5-phenoxy valeric acid (3HPxV), 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) and 3-hydroxy-9-phoxynonanoic acid (3HPxN) can be produced by strain YN2 with a substrate 11-phenoxyundecanoic acid.

[Example 59]

<Production of PHA containing HPxN unit, HPxHp unit and

HPxV unit by strain YN2 (two steps culture using glucose)>

The strain YN2 was inoculated in 200 mL of M9 .  
medium containing glucose 0.5% and 11-phenoxyundecanoic  
5 acid (PxUDA) 0.1% and was cultured at 30°C with shaking  
at 125 strokes/min. After 64 hours, the cultured cells  
were recovered by centrifugation, and were re-suspended  
in 200 mL of M9 medium containing glucose 0.5%  
and PxUDA 0.1% but not containing nitrogen source  
10 (NH<sub>4</sub>Cl), then further shake cultured at 30°C, 125  
strokes/min. After 24 hours, the cells were recovered  
by centrifugation, washed once with cold methanol,  
lyophilized and weighed.

The lyophilized pellet was suspended in chloroform  
15 100 mL, and stirred at 60°C for 24 hours to extract  
polymer. The extract was filtered using membrane  
filter, pore size 0.45 µm, concentrated using rotary  
evaporator, precipitated the concentrate with cold  
methanol, and recovered the precipitate to obtain the  
20 polymer by vacuum drying, then weighed as such.

Molecular weight of the obtained polymer was  
measured by means of gel-permeation chromatography  
(GPC: Toso, HLC-8020; Column: Polymer Laboratory,  
PL-gel, MIXED-C 5 µm; Solvent: chloroform; Molecular  
25 weight: reduced value for polystyrene).

Unit composition of the obtained polymer was  
analyzed by the following manner. The polymer sample 5

mg was poured into the 25 mL round-neck flask, and  
chloroform 2 mL and 2 mL of methanol containing  
sulfuric acid (3%, v/v). The mixture was refluxed at  
100°C for 3.5 hours, further added water thereto for  
5 separation. The organic layer was analyzed by means of  
gas-chromatography mass spectrograph (GC-MS, Shimadzu  
QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to  
identify methyl esterified PHA monomer unit. Yields of  
bacterial cells and polymer, and analytical result of  
10 the monomer unit are shown in Table 67. Mass spectra  
obtained by GC-MS of 3-hydroxybutyric acid methyl  
ester, 3-hydroxyhexanoic acid methyl ester,  
3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic  
acid methyl ester, 3-hydroxydodecanoic acid methyl  
15 ester, 3-hydroxydodecenoic acid methyl ester,  
3-hydroxy-5-phenoxyvaleric acid (3HPxV) methyl ester,  
3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) methyl ester  
and 3-hydroxy-9-phoxynonanoic acid (3HPxN) methyl  
ester are shown in FIG. 44 to FIG. 52.

20 Result indicates that PHA copolymer containing  
three units of 3-hydroxy-5-phenoxyvaleric acid (3HPxV),  
3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) and  
3-hydroxy-9-phoxynonanoic acid (3HPxN) can be  
produced by the strain YN2 with a substrate  
25 11-phenoxyundecanoic acid.

[Example 60]

<Production of PHA containing HPxN unit, HPxHp unit and



HPxV unit by strain H45 (two steps culture using glucose)>

Strain H45 was inoculated in 200 mL of M9 medium containing glucose 0.5% and 11-phenoxyundecanoic acid (PxUDA) 0.1% and was cultured at 30°C with shaking at 125 strokes/min. After 64 hours, the cultured cells were recovered by centrifugation, and were re-suspended in 200 mL of M9 medium containing glucose 0.5% and PxUDA 0.1% but not containing nitrogen source ( $\text{NH}_4\text{Cl}$ ), then further shake cultured at 30°C, 125 strokes/min. After 24 hours, the cells were recovered by centrifugation, washed once with cold methanol, lyophilized and weighed.

The lyophilized pellet was suspended in chloroform 100 mL, and stirred at 60°C for 24 hours to extract polymer. The extract was filtered using membrane filter, pore size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain the polymer by vacuum drying, then weighed as such.

Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C 5  $\mu\text{m}$ ; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5

mg was poured into the 25 mL round-neck flask, and  
chloroform 2 mL and 2 mL of methanol containing  
sulfuric acid (3%, v/v). The mixture was refluxed at  
100°C for 3.5 hours, further added water thereto for  
5 separation. The organic layer was analyzed by means of  
gas-chromatography mass spectrograph (GC-MS, Shimadzu  
QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to  
identify methyl esterified PHA monomer unit. Yields of  
bacterial cells and polymer, and analytical result of  
10 the monomer unit are shown in Table 68. Mass spectra  
obtained by GC-MS of 3-hydroxybutyric acid methyl  
ester, 3-hydroxyhexanoic acid methyl ester,  
3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic  
acid methyl ester, 3-hydroxydodecanoic acid methyl  
15 ester, 3-hydroxydodecenoic acid methyl ester,  
3-hydroxy-5-phenoxyvaleric acid (3HPxV) methyl ester,  
3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) methyl ester  
and 3-hydroxy-9-phoxynonanoic acid (3HPxN) methyl  
ester are shown in FIG. 53 to FIG. 61.

20 Result indicates that PHA copolymer containing  
three units of 3-hydroxy-5-phenoxyvaleric acid (3HPxV),  
3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) and  
3-hydroxy-9-phoxynonanoic acid (3HPxN) can be  
produced by the strain H45 with a substrate  
25 11-phenoxyundecanoic acid.

[Example 61]

<Production of PHA containing HPxO unit, HPxHx unit and

HPxB unit by strain YN2 (one step culture using polypeptone)>

The strain YN2 was inoculated in 200 mL of M9 medium containing polypeptone (Wako Pure Chemicals Co.) 0.5% and 8-phenoxyoctanoic acid (PxOA) 0.1% and was  
5 shake cultured at 30°C in 125 strokes/min. After 24 hours, the cells were recovered by centrifugation, washed once with cold methanol, lyophilized and weighed.

10 The lyophilized pellet was suspended in chloroform 100 mL, and stirred at 60°C for 24 hours to extract polymer. The extract was filtered using membrane filter, pore size 0.45  $\mu$ m, concentrated using rotary evaporator, precipitated the concentrate with cold  
15 methanol, and recovered the precipitate to obtain the polymer by vacuum drying, then weighed as such.

Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory,  
20 PL-gel, MIXED-C 5  $\mu$ m; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5 mg was poured into the 25 mL round-neck flask, and  
25 chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at 100°C for 3.5 hours, further added water thereto for

separation. The organic layer was analyzed by means of gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of bacterial cells and polymer, and analytical result of the monomer unit are shown in Table 69. Mass spectra obtained by GC-MS of 3-hydroxybutyric acid methyl ester, 3-hydroxy-4-phenoxybutyric acid (3HPxB) methyl ester, 3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) methyl ester and 3-hydroxy-8-phenoxyoctanoic acid (3HPxO) methyl ester are shown in FIG. 62 to FIG. 65.

Result indicates that PHA copolymer containing three units of 3-hydroxy-4-phenoxybutyric acid (3HPxB), 3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) and 3-hydroxy-8-phenoxyoctanoic acid (3HPxO) can be produced by the strain YN2 with a substrate 8-phenoxyoctanoic acid.

[Example 62]

<Production of PHA containing HPxO unit, HPxHx unit and HPxB unit by strain H45 (one step culture using polypeptone)>

The strain H45 was inoculated in 200 mL of M9 medium containing polypeptone (Wako Pure Chemicals Co.) 0.5% and 8-phenoxyoctanoic acid (PxOA) 0.1% and was shake cultured at 30°C in 125 strokes/min. After 24 hours, the cells were recovered by centrifugation, washed once with cold methanol, lyophilized and

weighed.

The lyophilized pellet was suspended in chloroform 100 mL, and stirred at 60°C for 24 hours to extract polymer. The extract was filtered using membrane  
5 filter, pore size 0.45  $\mu$ m, concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain the polymer by vacuum drying, then weighed as such.

Molecular weight of the obtained polymer was  
10 measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C 5  $\mu$ m; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was  
15 analyzed by the following manner. The polymer sample 5 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at 100°C for 3.5 hours, further added water thereto for  
20 separation. The organic layer was analyzed by means of gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of bacterial cells and polymer, and analytical result of  
25 the monomer unit are shown in Table 70. Mass spectra obtained by GC-MS of 3-hydroxybutyric acid methyl ester, 3-hydroxy-4-phenoxybutyric acid (3HPxB) methyl

ester, 3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) methyl ester and 3-hydroxy-8-phenoxyoctanoic acid (3HPxO) methyl ester are shown in FIG. 66 to FIG. 69.

Result indicates that PHA copolymer containing  
5 three units of 3-hydroxy-4-phenoxybutyric acid (3HPxB),  
3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) and  
3-hydroxy-8-phenoxyoctanoic acid (3HPxO) can be  
produced by the strain H45 with a substrate  
8-phenoxyoctanoic acid.

10 [Example 63]

<Production of PHA containing HPxO unit, HPxHx unit and  
HPxB unit by strain YN2 (two steps culture using  
glucose)>

The strain YN2 was inoculated in 200 mL of M9  
15 medium containing glucose 0.5% and 8-phenoxyoctanoic  
acid (PxOA) 0.1% and was cultured at 30°C with shaking  
at 125 strokes/min. After 48 hours, the cultured cells  
were recovered by centrifugation, and were re-suspended  
in 200 mL of M9 medium containing glucose 0.5% and PxOA  
20 0.1% but not containing nitrogen source ( $\text{NH}_4\text{Cl}$ ), then  
further shake cultured at 30°C in 125 strokes/min.  
After 24 hours, bacterial cells were recovered by  
centrifugation, washed once with cold methanol,  
lyophilized and were weighed.

25 The lyophilized pellet was suspended in chloroform  
100 mL, and stirred at 60°C for 24 hours to extract  
polymer. The extract was filtered using membrane

filter, pore size 0.45  $\mu\text{m}$ , concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain the polymer by vacuum drying, then weighed as such.

5           Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C 5  $\mu\text{m}$ ; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

10           Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at  
15           100°C for 3.5 hours, further added water thereto for separation. The organic layer was analyzed by means of gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of  
20           bacterial cells and polymer, and analytical result of the monomer unit are shown in Table 71. Mass spectra obtained by GC-MS measurement of 3-hydroxybutyric acid methyl ester, 3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic acid methyl ester,  
25           3-hydroxy-4-phenoxybutyric acid (3HPxB) methyl ester, 3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) methyl ester and 3-hydroxy-8-phenoxyoctanoic acid (3HPxO) methyl

ester are shown in FIG. 70 to FIG. 75.

Result indicates that PHA copolymer containing three units of 3-hydroxy-4-phenoxybutyric acid (3HPxB), 3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) and  
5 3-hydroxy-8-phenoxyoctanoic acid (3HPxO) can be produced by the strain YN2 with a substrate 8-phenoxyoctanoic acid.

[Example 64]

<Production of PHA containing HPxO unit, HPxHx unit and  
10 HPxB unit by strain H45 (two steps culture using glucose)>

The strain H45 was inoculated in 200 mL of M9 medium containing glucose 0.5% and 8-phenoxyoctanoic acid (PxOA) 0.1% and cultured at 30°C with shaking at  
15 125 strokes/min. After 48 hours, cultured bacterial cells were recovered by centrifugation, and were re-suspended in 200 mL of M9 medium containing glucose 0.5% and PxOA 0.1% but not containing nitrogen source (NH<sub>4</sub>Cl), then further shake cultured at 30°C, 125  
20 strokes/min. After 24 hours, the cells were recovered by centrifugation, washed once with cold methanol, lyophilized and weighed.

The lyophilized pellet was suspended in chloroform 100 mL, and stirred at 60°C for 24 hours to extract  
25 polymer. The extract was filtered using membrane filter, pore size 0.45 µm, concentrated using rotary evaporator, precipitated the concentrate with cold



methanol, and recovered the precipitate to obtain the polymer by vacuum drying, then weighed as such.

Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory, PL-gel, MIXED-C 5  $\mu$ m; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at 100°C for 3.5 hours, further added water thereto for separation. The organic layer was analyzed by means of gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of bacterial cells and polymer, and analytical result of the monomer unit are shown in Table 72. Mass spectra obtained by GC-MS measurement of 3-hydroxybutyric acid methyl ester, 3-hydroxyhexanoic acid methyl ester, 3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic acid methyl ester, 3-hydroxydodecanoic acid methyl ester, 3-hydroxydodecenoic acid methyl ester, 3-hydroxy-4-phenoxybutyric acid (3HPxB) methyl ester, 3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) methyl ester and 3-hydroxy-8-phenoxyoctanoic acid (3HPxO) methyl

ester are shown in FIG. 76 to FIG. 84.

Result indicates that PHA copolymer containing three units of 3-hydroxy-4-phenoxybutyric acid (3HPxB), 3-hydroxy-6-phenoxyhexanoic acid (3HPxHx) and  
5 3-hydroxy-8-phenoxyoctanoic acid (3HPxO) can be produced by the strain H45 with a substrate 8-phenoxyoctanoic acid.

[Example 65]

<Production of PHA containing HPxHp unit and HPxV unit  
10 by strain YN2 (one step culture using polypeptone)>

The strain YN2 was inoculated in 200 mL of M9 medium containing polypeptone (Wako Pure Chemicals Co.) 0.5% and 7-phenoxyheptanoic acid (PxHpA) 0.1% and was cultured at 30°C with shaking at 125 strokes/min.  
15 After 64 hours, the cells were recovered by centrifugation, washed once with cold methanol, lyophilized and weighed.

The lyophilized pellet was suspended in chloroform 100 mL, and stirred at 60°C for 24 hours to extract  
20 polymer. The extract was filtered using membrane filter, pore size 0.45  $\mu$ m, concentrated using rotary evaporator, precipitated the concentrate with cold methanol, and recovered the precipitate to obtain the polymer by vacuum drying, then weighed as such.

25 Molecular weight of the obtained polymer was measured by means of gel-permeation chromatography (GPC: Toso, HLC-8020; Column: Polymer Laboratory,

PL-gel, MIXED-C 5  $\mu$ m; Solvent: chloroform; Molecular weight: reduced value for polystyrene).

Unit composition of the obtained polymer was analyzed by the following manner. The polymer sample 5  
5 mg was poured into the 25 mL round-neck flask, and chloroform 2 mL and 2 mL of methanol containing sulfuric acid (3%, v/v). The mixture was refluxed at 100°C for 3.5 hours, further added water thereto for separation. The organic layer was analyzed by means of  
10 gas-chromatography mass spectrograph (GC-MS, Shimadzu QP-5050, Column: DB-WAXETR (J & W Inc.), EI method) to identify methyl esterified PHA monomer unit. Yields of bacterial cells and polymer, and analytical result of the monomer unit are shown in Table 73. Mass spectra  
15 obtained by GC-MS of 3-hydroxybutyric acid methyl ester, 3-hydroxyoctanoic acid methyl ester, 3-hydroxydecanoic acid methyl ester, 3-hydroxy-5-phenoxyvaleric acid (3HPxV) methyl ester and 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) methyl  
20 ester are shown in FIG. 85 to FIG. 89.

Result indicates that PHA copolymer containing two units of 3-hydroxy-5-phenoxyvaleric acid (3HPxV) and 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) can be produced by strain YN2 with a substrate  
25 7-phenoxyheptanoic acid.

[Example 66]

<Production of PHA Containing HPxHp unit and HPxV Unit

by Using strain H45 (Polypeptone, One-Step Culture)>

Strain H45 was inoculated in 200 mL of M9 medium containing 0.5% polypeptone (Wako Jun-Yaku Kogyo available) and 0.1% 7-phenoxyheptanoic acid (PxHpA) and  
5 cultured with shaking at 125 strokes/min at 30°C. After 64 hr, the cells were collected by centrifugation, washed once with cold methanol, lyophilized and weighed.

This lyophilized pellet was suspended in 100 mL of  
10 chloroform and stirred at 60°C for 24 hr to extract the polymer. After filtered through a 0.45 µm pore-size membrane filter, the extract was concentrated by using a rotary evaporator and the concentrated solution was re-precipitated in cold methanol, and the precipitate  
15 was collected, vacuum-dried and weight as the polymer.

The molecular weight of the obtained polymer was measured by using gel permeation chromatography (GPC: Toso/HLC-8020; column: Polymer Laboratory/PL gel/MIXED-C/5 µm; solvent: chloroform; polystyrene  
20 reduced molecular weight).

The unit composition of the obtained polymer was analyzed as follows: To 5 mg of a polymer sample put in a 25 mL volume round bottom flask, 2 mL of chloroform and 2 mL of methanol containing 3% (v/v) sulfuric acid  
25 was added, the mixture was subjected to 100°C and 3.5 hr reflux and separated with a further addition of water, then the organic layer was analyzed by a gas

chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; column: DB-WAXETR (J & W Co. available); EI method) to identify a methyl esterified substance of the PHA monomer unit. Table 74 shows the yield of the cells and polymers and the analyzed result of the monomer unit. Besides, FIGS. 90 to 92 show the mass spectra, obtained by the GC-MS measurement of 3-hydroxybutyrate methyl ester, 3-hydroxy-5-phenoxyvalerate (3HPxV) methyl ester and 3-hydroxy-7-phenoxyheptanoate (3HPxHp) methyl ester, respectively.

From this result, it was revealed that PHA copolymer containing two units of 3-hydroxy-5-phenoxyvaleric acid (3HPxV) and 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) could be produced using strain H45 and 7-phenoxyheptanoic acid as the substrate.

[Example 67]

<Production of PHA Containing HPxHp unit and HPxV Unit by Using strain YN2 (Glucose, Two-Step Culture)>

Strain YN2 was inoculated in 200 mL of M9 culture medium containing 0.5% glucose and 0.1% 7-phenoxyheptanoic acid (PxHpA) and cultured with 125 strokes/min of shaking at 30°C. After 64 hr, the cells were collected by centrifugation, re-suspended into 200 mL of M9 culture medium containing 0.5% glucose and 0.1% PxHpA and no nitrogen source (NH<sub>4</sub>Cl) and further

cultured with 125 strokes/min of shaking at 30°C. After 24 hr, the cells were collected by centrifugation, washed once with cold methanol, lyophilized and weighed.

5           This lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 24 hr to extract polymer. After filtered through a 0.45  $\mu$ m pore-size membrane filter, the extract was concentrated by using a rotary evaporator and the concentrated solution was  
10 re-precipitated in cold methanol, and the precipitate was collected and vacuum-dried to obtain a polymer, then this polymer was weighed.

          The molecular weight of the obtained polymer was measured by using gel permeation chromatography (GPC:  
15 Toso/HLC-8020; column: Polymer Laboratory/PL gel/MIXED-C/5  $\mu$ m; solvent: chloroform; polystyrene reduced molecular weight).

          The unit composition of the obtained polymer was analyzed as follows: To 5 mg of a polymer sample put in  
20 a 25 mL volume round bottom flask, 2 mL of chloroform and 2 mL of methanol containing 3% (v/v) sulfuric acid was added, the mixture was subjected to 100°C and 3.5 hr reflux and separated with a further addition of water, then the organic layer was analyzed on a gas  
25 chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; column: DB-WAXETR (J & W Co. available); EI

method) to identify the methyl esterified substance of the PHA monomer unit. Table 75 shows the yield of the cells and polymer and the analyzed result of the monomer unit. Besides, FIGS. 93 to 100 show the mass spectra, obtained by the GC-MS measurement of 3-hydroxybutyrate methyl ester, 3-hydroxyhexanoate methyl ester, 3-hydroxyoctanoate methyl ester, 3-hydroxydecanoate methyl ester, 3-hydroxydodecanoate methyl ester, 3-hydroxydodecenoate methyl ester, 3-hydroxy-5-phenoxyvalerate (3HPxV) methyl ester and 3-hydroxy-7-phenoxyheptanoate (3HPxHp) methyl ester, respectively.

From this result, it was revealed that PHA copolymer containing two units of 3-hydroxy-5-phenoxyvaleric acid (3HPxV) and 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) could be produced using strain YN2 and 7-phenoxyheptanoic acid as the substrate.

[Example 68]

<Production of PHA Containing HPxHp unit and HPxV Unit by Using strain H45 (Glucose, Two-Step Culture)>

Strain H45 was inoculated in 200 mL of M9 culture medium containing 0.5% glucose and 0.1% 7-phenoxyheptanoic acid (PxHpA) and cultured with 125 strokes/min of shaking at 30°C. After 64 hr, the cells were collected by centrifugation, re-suspended into 200 mL of M9 culture medium containing 0.5% glucose and

0.1% PxHpA and no nitrogen source ( $\text{NH}_4\text{Cl}$ ) and further cultured with 125 strokes/min of shaking at 30°C. After 24 hr, the cells were collected by centrifugation, washed once with cold methanol, lyophilized and weighed.

This lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 24 hr to extract polymer. After filtered through a 0.45  $\mu\text{m}$  pore-size membrane filter, the extract was concentrated using a rotary evaporator and the concentrated solution was re-precipitated in cold methanol, and the precipitate was collected and vacuum-dried to obtain a polymer, then this polymer was weighed.

The molecular weight of the obtained polymer was measured by using gel permeation chromatography (GPC: Tosoh/HLC-8020; column: Polymer Laboratory/PL gel/MIXED-C/5  $\mu\text{m}$ ; solvent: chloroform; polystyrene reduced molecular weight).

The unit composition of the obtained polymer was analyzed as follows: To 5 mg of a polymer sample put in a 25 mL volume round bottom flask, 2 mL of chloroform and 2 mL of methanol containing 3% (v/v) sulfuric acid was added, the mixture was subjected to 100°C and 3.5 hr reflux and separated with a further addition of water, then the organic layer was analyzed on a gas chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; column: DB-WAXETR (J & W Co. available); EI



method) to identify a methyl esterified substance of the PHA monomer unit. Table 76 shows the yield of the cells and polymer and the analyzed result of the monomer unit. Besides, FIGS. 101 to 107 show the mass spectra, obtained by the GC-MS measurement, of 3-hydroxyhexanoate methyl ester, 3-hydroxyoctanoate methyl ester, 3-hydroxydecanoate methyl ester, 3-hydroxydodecanoate methyl ester, 3-hydroxydodecenoate methyl ester, 3-hydroxy-5-phenoxyvalerate (3HPxV) methyl ester and 3-hydroxy-7-phenoxyheptanoate (3HPxHp) methyl ester, respectively.

From this result, it was revealed that PHA copolymer containing two units of 3-hydroxy-5-phenoxyvaleric acid (3HPxV) and 3-hydroxy-7-phenoxyheptanoic acid (3HPxHp) could be produced using strain H45 with 7-phenoxyheptanoic acid as the substrate.

[Example 69]

<Production of PHA Containing PHPxV Unit by Using Strain YN2 (Sodium Malate Two-Step Culture)>

Strain YN2 was inoculated in 200 mL of M9 culture medium containing 0.5% sodium malate and 0.1% 5-phenoxyvaleric acid (PxVA) and cultured with 125 strokes/min of shaking at 30°C. After 60 hr, the cells were collected by centrifugation, re-suspended into 200 mL of M9 culture medium containing 0.5% sodium malate and 0.1% PxVA and no nitrogen source ( $\text{NH}_4\text{Cl}$ ) and further

cultured with 125 strokes/min of shaking at 30°C. After 24 hr, the cells were collected by centrifugation, washed once with cold methanol, lyophilized and weighed.

5           This lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 24 hr to extract polymer. After filtered through a 0.45  $\mu$ m pore-size membrane filter, the extract was concentrated by using a rotary evaporator and the concentrated solution was  
10 re-precipitated in cold methanol, further the precipitate alone was collected and vacuum-dried to obtain a polymer, then this polymer was weighed.

          The molecular weight of the obtained polymer was measured by using gel permeation chromatography (GPC:  
15 Toso/HLC-8020; column: Polymer Laboratory/PL gel/MIXED-C/5  $\mu$ m; solvent: chloroform; polystyrene reduced molecular weight).

          The unit composition of the obtained polymer was analyzed as follows: To 5 mg of a polymer sample put in  
20 a 25 mL volume round bottom flask, 2 mL of chloroform and 2 mL of methanol containing 3% (v/v) sulfuric acid was added, the mixture was subjected to 100°C and 3.5 hr reflux and separated with a further addition of water, then the organic layer was analyzed on a gas  
25 chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; column: DB-WAXETR (J & W Co. available); EI method) to identify a methyl esterified substance of

the PHA monomer unit. Table 77 shows the yield of the cells and polymer and the analyzed result of the monomer unit. Besides, FIGS. 108 to 114 show the mass spectra, obtained by the GC-MS measurement, of

5 3-hydroxybutyrate methyl ester, 3-hydroxyhexanoate methyl ester, 3-hydroxyoctanoate methyl ester, 3-hydroxydecanoate methyl ester, 3-hydroxydodecanoate methyl ester, 3-hydroxydodecenoate methyl ester and 3-hydroxy-5- phenoxyvalerate (3HPxV) methyl ester,

10 respectively.

From this result, it was revealed that PHA copolymer containing 3-hydroxy-5-phenoxyvaleric acid (3HPxV) unit could be produced using strain YN2 with 5-phenoxyvaleric acid as the substrate.

15 [Example 70]

<Production of PHA Containing HPxV Unit by Using Strain H45 (Sodium Malate Two-Step Culture)>

Strain H45 was inoculated in 200 mL of M9 culture containing 0.5% sodium malate and 0.1% 5-phenoxyvaleric acid (PxVA) and cultured with 125 strokes/min of

20 shaking at 30°C. After 60 hr, the cells were collected by centrifugation, re-suspended into 200 mL of M9 culture medium containing 0.5% sodium maleate and 0.1% PxVA and no nitrogen source ( $\text{NH}_4\text{Cl}$ ) and further cultured

25 with 125 strokes/min of shaking at 30°C. After 24 hr, the cells were collected by centrifugation, washed once with cold methanol, lyophilized and weighed.

This lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 24 hr to extract polymer. After filtered through a 0.45  $\mu$ m pore-size membrane filter, the extract was concentrated using a rotary evaporator and the concentrated solution was re-precipitated in cold methanol, and the precipitate was collected and vacuum-dried to obtain a polymer, then this polymer was weighed.

The molecular weight of the obtained polymer was measured by using gel permeation chromatography (GPC: Toso/HLC-8020; column: Polymer Laboratory/PL gel/MIXED-C/5  $\mu$ m; solvent: chloroform; polystyrene reduced molecular weight).

The unit composition of the obtained polymer was analyzed as follows: To 5 mg of a polymer sample put in a 25 mL volume round bottom flask, 2 mL of chloroform and 2 mL of methanol containing 3% (v/v) sulfuric acid was added, the mixture was subjected to 100°C and 3.5 hr reflux and separated with a further addition of water, then the organic layer was analyzed on a gas chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; column: DB-WAXETR (J & W Co. available); EI method) to identify a methyl esterified substance of the PHA monomer unit. Table 78 shows the yield of the cells and polymer and the analyzed result of the monomer unit. Besides, FIGS. 115 to 120 show the mass spectra, obtained by the GC-MS measurement, of

3-hydroxyhexanoate methyl ester, 3-hydroxyoctanoate  
methyl ester, 3-hydroxydecanoate methyl ester,  
3-hydroxydodecanoate methyl ester, 3-hydroxydodecenoate  
methyl ester and 3-hydroxy-5-phenoxyvalerate (3HPxV)  
5 methyl ester, respectively.

From this result, it was revealed that PHA  
copolymer containing 3-hydroxy-5-phenoxyvaleric acid  
(3HPxV) unit could be produced using strain H45 with  
5-phenoxyvaleric acid as the substrate.

10 [Example 71]

<Production of PHA Containing HPV Unit by Using Strain  
YN2 (Fructose Two-Step Culture)>

Strain YN2 was inoculated in 200 mL of M9 culture  
medium containing 0.5% fructose and 0.1%  
15 5-phenylvaleric acid (PVA) and cultured with 125  
strokes/min of shaking at 30°C. After 120 hr, the  
cells were collected by centrifugation, re-suspended  
into 200 mL of M9 culture medium containing 0.5%  
fructose and 0.1% PxVA and no nitrogen source ( $\text{NH}_4\text{Cl}$ )  
20 and further cultured with 125 strokes/min of shaking at  
30°C. After 50 hr, the cells were collected by  
centrifugation, washed once with cold methanol,  
lyophilized and weighed.

This lyophilized pellet was suspended in 100 mL of  
25 chloroform and stirred at 60°C for 24 hr to extract  
polymer. After filtered through a 0.45  $\mu\text{m}$  pore-size  
membrane filter, the extract was concentrated by using

a rotary evaporator and the concentrated solution was re-precipitated in cold methanol, further the precipitate alone was collected and vacuum-dried to obtain a polymer, then this polymer was weighed.

5           The molecular weight of the obtained polymer was measured using gel permeation chromatography (GPC: Toso/HLC-8020; column: Polymer Laboratory/PL gel/MIXED-C/5  $\mu$ m; solvent: chloroform; polystyrene reduced molecular weight).

10           The unit composition of the obtained polymer was analyzed as follows: To 5 mg of a polymer sample put in a 25 mL volume round bottom flask, 2 mL of chloroform and 2 mL of methanol containing 3% (v/v) sulfuric acid was added, the mixture was subjected to 100°C and 3.5  
15 hr reflux and separated with a further addition of water, then the organic layer was analyzed on a gas chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; column: DB-WAXETR (J & W Co. available); EI method) to identify a methyl esterified substance of  
20 the PHA monomer unit. Table 79 shows the yield of the cells and polymer and the analyzed result of the monomer unit. Besides, FIGS. 121 to 123 show the mass spectra, obtained by the GC-MS measurement of  
25 3-hydroxyoctanoate methyl ester, 3-hydroxydecanoate methyl ester and 3-hydroxy-5-phenylvalerate (3HPV) methyl ester, respectively.

From this result, it was revealed that a PHA

copolymer containing 3-hydroxy-5-phenylvaleric acid (3HPV) unit could be produced using strain YN2 with 5-phenylvaleric acid as the substrate.

[Example 72]

5     <Production of PHA Containing HPV Unit by Using Strain YN2 (Mannose Two-Step Culture)>

Strain YN2 was inoculated in 200 mL of M9 culture medium containing 0.5% mannose and 0.1% 5-phenylvaleric acid (PVA) and cultured with 125 strokes/min of shaking  
10     at 30°C. After 43 hr, the cells were collected by centrifugation, re-suspended into 200 mL of M9 culture medium containing 0.5% mannose and 0.1% PxVA and no nitrogen source ( $\text{NH}_4\text{Cl}$ ) and further cultured with 125  
15     strokes/min of shaking at 30°C. After 91 hr, the cells were collected by centrifugation, washed once with cold methanol, lyophilized and weighed.

This lyophilized pellet was suspended in 100 mL of chloroform and stirred at 60°C for 24 hr to extract polymer. After filtered through a 0.45  $\mu\text{m}$  pore-size  
20     membrane filter, the extract was concentrated using a rotary evaporator and the concentrated solution was re-precipitated in cold methanol, and the precipitate was collected and vacuum-dried to obtain a polymer, then this polymer was weighed.

25     The molecular weight of the obtained polymer was measured by using gel permeation chromatography (GPC: Toso/HLC-8020; column: Polymer Laboratory/PL

gel/MIXED-C/5  $\mu$ m; solvent: chloroform; polyst reduced molecular weight).

The unit composition of the obtained pol  
analyzed as follows: To 5 mg of a polymer sam  
5 a 25 mL volume round bottom flask, 2 mL of ch  
and 2 mL of methanol containing 3% (v/v) sulf  
was added, the mixture was subjected to 100°C  
hr reflux and separated with a further additi  
water, then the organic layer was analyzed on  
10 chromatograph - mass spectrometer (GC-MS, Shir  
QP-5050; column: DB-WAXETR (J & W Co. availabl  
method) to identify a methyl esterified subst  
the PHA monomer unit. Table 80 shows the yie  
cells and polymer and the analyzed result of  
15 monomer unit. Besides, FIGS. 124 and 125 sho  
spectra, obtained by the GC-MS measurement, o  
3-hydroxyoctanoate methyl ester and  
3-hydroxy-5-phenylvalerate (3HPV) methyl este  
respectively.

20 From this result, it was revealed that P  
copolymer containing 3-hydroxy-5-phenylvaleric  
(3HPV) unit could be produced using strain YN  
5-phenylvaleric acid as the substrate.

[Example 73]

25 <Production of PHA Containing HPV Unit by Usin  
YN2 (Sodium Lactate Two-Step Culture)>

Strain YN2 was inoculated in 200 mL of M9



gel/MIXED-C/5  $\mu$ m; solvent: chloroform; polystyrene reduced molecular weight).

5       The unit composition of the obtained polymer was analyzed as follows: To 5 mg of a polymer sample put in a 25 mL volume round bottom flask, 2 mL of chloroform and 2 mL of methanol containing 3% (v/v) sulfuric acid was added, the mixture was subjected to 100°C and 3.5 hr reflux and separated with a further addition of water, then the organic layer was analyzed on a gas chromatograph - mass spectrometer (GC-MS, Shimadzu 10 QP-5050; column: DB-WAXETR (J & W Co. available); EI method) to identify a methyl esterified substance of the PHA monomer unit. Table 80 shows the yield of the cells and polymer and the analyzed result of the 15 monomer unit. Besides, FIGS. 124 and 125 show the mass spectra, obtained by the GC-MS measurement, of 3-hydroxyoctanoate methyl ester and 3-hydroxy-5-phenylvalerate (3HPV) methyl ester, respectively.

20       From this result, it was revealed that PHA copolymer containing 3-hydroxy-5-phenylvaleric acid (3HPV) unit could be produced using strain YN2 with 5-phenylvaleric acid as the substrate.

[Example 73]

25       <Production of PHA Containing HPV Unit by Using Strain YN2 (Sodium Lactate Two-Step Culture)>

Strain YN2 was inoculated in 200 mL of M9 culture

medium containing 0.5% sodium lactate and 0.1%  
5-phenylvaleric acid (PVA) and cultured with 125  
strokes/min of shaking at 30°C. After 46 hr, the cells  
were collected by centrifugation, re-suspended into 200  
5 mL of M9 culture containing 0.5% sodium lactate and  
0.1% PxVA and no nitrogen source ( $\text{NH}_4\text{Cl}$ ) and further  
cultured with 125 strokes/min of shaking at 30°C.  
After 28 hr, the cells were collected by  
centrifugation, washed once with cold methanol,  
10 lyophilized and weighed.

This lyophilized pellet was suspended in 100 mL of  
chloroform and stirred at 60°C for 24 hr to extract  
polymer. After filtered through a 0.45  $\mu\text{m}$  pore-size  
membrane filter, the extract was concentrated by using  
15 a rotary evaporator and the concentrated solution was  
re-precipitated in cold methanol, and the precipitate  
was collected and vacuum-dried to obtain a polymer,  
then this polymer was weighed.

The molecular weight of the obtained polymer was  
20 measured using gel permeation chromatography (GPC:  
Toso/HLC-8020; column: Polymer Laboratory/PL  
gel/MIXED-C/5  $\mu\text{m}$ ; solvent: chloroform; polystyrene  
reduced molecular weight).

The unit composition of the obtained polymer was  
25 analyzed as follows: To 5 mg of a polymer sample put in  
a 25 mL volume round bottom flask, 2 mL of chloroform  
and 2 mL of methanol containing 3% (v/v) sulfuric acid

was added, the mixture was subjected to 100°C and 3.5 hr reflux and separated with a further addition of water, then the organic layer was analyzed on a gas chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; column: DB-WAXETR (J & W Co. available); EI method) to identify a methyl esterified substance of the PHA monomer unit. Table 81 shows the yield of the cells and polymer and the analyzed result of the monomer unit. Besides, FIGS. 126 to 129 show the mass spectra, obtained by the GC-MS measurement, of 3-hydroxybutyrate methyl ester, 3-hydroxyoctanoate methyl ester, 3-hydroxydecanoate methyl ester and 3-hydroxy-5-phenylvalerate (3HPV) methyl ester, respectively.

From this result, it was revealed that PHA copolymer containing 3-hydroxy-5-phenylvaleric acid (3HPV) unit could be produced using strain YN2 with 5-phenylvaleric acid as the substrate.

[Example 74]

<Production of PHA Containing HPxB Unit by Using Strain YN2 (Disodium Malate Two-Step Culture)>

*Pseudomonas cichorii* strain YN2 was inoculated in 200 mL of 5 types of M9 culture media each containing 0.1% 4-phenoxy-n-butyric acid (PxBA) and one of 0.5% of disodium malate semihydrate, L-sodium glutamate monohydrate, D(+)-glucose and n-nonanoic acid and polypeptone (Nihon Seiyaku) respectively, and cultured

with 125 strokes/min of shaking at 30°C. After 48 hr, the cells were collected by centrifugation, washed once with cold methanol and vacuum-dried.

These five pellets were suspended in 20 mL of chloroform separately and stirred at 60°C for 20 hr to extract PHA. After filtered through a 0.45 µm pore-size membrane filter, each extract was concentrated using a rotary evaporator and the concentrated solution was re-precipitated in cold methanol, and each precipitate was collected and vacuum-dried to obtain PHA. After subjected to methanolysis in accordance with the usual way, the obtained PHAs were analyzed using a gas chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; EI method) to identify the methyl esterified substance of the PHA monomer unit. As a result, in case of culturing with disodium malate as the growth carbon source, as shown in Table 82, PHA having a high proportion of 3-hydroxy-4-phenoxy-n-butyric acid (3HPxB) unit, a desired monomer unit derived from 4-phenoxy-n-butyric acid, was obtained at a high yield. Furthermore, Table 83 shows the yield of the cells and polymer and the composition of the polymer in the case of culture using disodium malate.

[Example 75]

<Production of PHA Containing HPxB Unit by Using Strain YN2 (Disodium Malate Two-Step Culture: Mass Culture)>

*Pseudomonas cichorii* strain YN2 was inoculated in 200 mL of M9 culture medium containing 0.5% yeast extract (Oriental Yeast Industries), and cultured with 125 strokes/min of shaking at 30°C as a seed culture.

5 In a 10 L jar fermenter containing 5 L of M9 medium containing 0.5% of disodium malate semihydrate and 0.1% 4-phenoxy-n-butyric acid, 50 mL of seed cells was inoculated and shake cultured at 30°C with 80 stroke/min under aeration of 2.5 L/min. After 39 hr, 10 the cells were collected by centrifugation. This pellet was suspended in 120 mL of an approx. 1.7% sodium hypochlorite solution and shaken at 4°C for 2 hr to extract PHA. PHA was collected by centrifugation and dried to obtain 56 mg of PHA per liter culture 15 medium.

After subjected to methanolysis in accordance with the usual way, the obtained PHA was analyzed using a gas chromatograph - mass spectrometer (GC-MS, Shimadzu QP-5050; EI method) to identify a methyl esterified 20 substance of the PHA monomer unit. As a result, the composition ratio (GC-MS, peak area ratio) of 3-hydroxy-4-phenoxy-n-butyric acid unit, a desired monomer unit derived from 4-phenoxy-n-butyric acid, was 99.7%.

25

# SEQUENCE LISTING

<110> Canon Inc.

<120> Polyhydroxyalkanoate its manufacturing method, and microorganisms  
those are used for the method.

<130> 4351009

<160> 1

<210> 1

<211> 1501

<212> DNA

<213> Pseudomonas jessenii 161 strain.

<400> 1

tgaacgctgg cggcaggcct aacacatgca agtcgagcgg atgacgggag ctigtctctg	60
aattcagcgg cggacgggtg agtaatgcct aggaatctgc ctggtagtgg gggacaacgt	120
ctcgaaaggg acgctaatac cgcatacgtc ctacgggaga aagcagggga ccttcgggcc	180
tigcgtatc agatgagcct aggtcggatt agctagttag tgaggtaatg gctcaccaag	240
gcgacgatcc gtaactggtc tgagaggatg atcagtcaca ctggaactga gacacggtec	300
agactcctac gggaggcagc agtggggaat attggacaat gggcgaaagc ctgatccagc	360
catgccgcgt gtgtgaagaa ggcttcgga ttgtaaagca cttaagttag ggaggaaggg	420
cattaacctt atacgttagt gttttgacgt taccgacaga ataagcaccg gctaactctg	480
tgccagcagc cgcggtaata cagagggtgc aagcgtaaat cggaattact gggcgtaaag	540
cgcgcgtagg tggtttgta agttggatgt gaaagccccg ggctcaacct gggaactgca	600

ttcaaaactg acaagctaga gtatggtaga gggtaggtaga atttcctgtg tagcggtaga	660
atgcgtagat ataggaagga acaccagtag cgaaggcgac cacctggact gatactgaca	720
ctgaggtagc aaagcgtggg gagcaaacag gattagatac cctggtagtc cacgccgtaa	780
acgatgtcaa ctacccgttg ggagccttga gccttagtg gcgcagctaa cgcatlaagt	840
tgaccgcctg gggagtagcg ccgcaagggt aaaactcaaa tgaattgacg gggggccgca	900
caagcggtag agcatgtggt ttaattcgaa gcaacgcgaa gaaccttacc aggccttagc	960
atccaatgaa ctttccagag atggatgggt gccttcggga acattgagac aggtgctgca	1020
tggctgtcgt cagctcgtgt cgtgagatgt tgggttaagt cccgtaacga gcgcaaccct	1080
tgtccttagt taccagcagc taatggtagg cactctaagg agactgccgg tgacaaaccg	1140
gaggaaggtag gggatgacgt caagtcacga tggcccttac ggcctgggct acacacgtgc	1200
tacaatggtc ggtacagagg gttgccaagc cgcgaggtag agctaattcc acaaaaccga	1260
tcgtagtcgg gatcgagtc tgcaactcga ctgcgtgaag tcggaatcgc tagtaatcgc	1320
gaatcagaat gtcgcggtag atacgttccc gggccttgta cacaccgcc gtcacacat	1380
gggagtaggt tgcaccagaa gtagctagtc taaccttcgg gaggacggtt accacgggtg	1440
gattcatgac tggggtgaag tcgtaccaag gtagccgtag gggaacctgc ggctggatca	1500
c	1501

Table 1

Carbon Source (alkanoate)	Weight of dry cell (mg/L)	Weight of dry polymer (mg/L)	Yield (%)
6-phenoxyhexanoic acid	950	100	10.5
8-phenoxyoctanoic acid	820	90	11
11-phenoxyundecanoic acid	150	15	10

Table 2

NA:CHBA	CDW	PDW	Yield	Unit
5:5	756.0	89.1	11.8	NA, CHBA
1:9	132.8	19.3	14.5	NA, CHBA

CDW: Cell (Dry Weight)  
 PDW: Polymer (Dry Weight)  
 Yield: PDW/CDW (%)

Table 3

Chemical Shift/ppm	type	Assignment
1.67	m	c, d
2.39	t	b
2.62	t	e
6.97	t	h, j
7.12	t	g, k
10.7	broad	COOH



Table 4

*P. cichorii* strain H45

Cell (Dry weight)	750mg/L
Polymer (Dry weight)	400mg/L
Polymer (Dry weight)/Cell (Dry weight)	53%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	0%
3-hydroxyvaleric acid	0%
3-hydroxyhexanoic acid	0%
3-hydroxyheptanoic acid	13%
3-hydroxyoctanoic acid	3%
3-hydroxynonanoic acid	37%
3-hydroxydecanoic acid	0%
3-hydroxy-5-(4-fluorophenyl)valeric acid	47%

Table 5

*P. cichorii* strain YN2

Cell (Dry weight)	850mg/L
Polymer (Dry weight)	420mg/L
Polymer (Dry weight)/Cell (Dry weight)	49%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	1%
3-hydroxyvaleric acid	1%
3-hydroxyhexanoic acid	0%
3-hydroxyheptanoic acid	15%
3-hydroxyoctanoic acid	2%
3-hydroxynonanoic acid	68%
3-hydroxydecanoic acid	0%
3-hydroxy-5-(4-fluorophenyl)valeric acid	13%

Table 6

*P. putida* P 91 strain

Cell (Dry weight)	670mg/L
Polymer (Dry weight)	51mg/L
Polymer (Dry weight)/Cell (Dry weight)	8%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	0%
3-hydroxyvaleric acid	1%
3-hydroxyhexanoic acid	0%
3-hydroxyheptanoic acid	11%
3-hydroxyoctanoic acid	2%
3-hydroxynonanoic acid	34%
3-hydroxydecanoic acid	0%
3-hydroxy-5-(4-fluorophenyl)valeric acid	52%

Table 7

*P. jessenii* strain P161

Cell (Dry weight)	1200mg/L
Polymer (Dry weight)	640mg/L
Polymer (Dry weight)/Cell (Dry weight)	53%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	1%
3-hydroxyvaleric acid	1%
3-hydroxyhexanoic acid	0%
3-hydroxyheptanoic acid	17%
3-hydroxyoctanoic acid	3%
3-hydroxynonanoic acid	45%
3-hydroxydecanoic acid	0%
3-hydroxy-5-(4-fluorophenyl)valeric acid	33%

Table 8

Purified PHA

Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	0%
3-hydroxyvaleric acid	0%
3-hydroxyhexanoic acid	0%
3-hydroxyheptanoic acid	0%
3-hydroxyoctanoic acid	0%
3-hydroxynonanoic acid	0%
3-hydroxydecanoic acid	0%
3-hydroxy-5-(4-fluorophenyl)valeric acid	100%

Table 9 (Results of <sup>1</sup>H spectrometry)

Resonance frequency: 400MHz

δ (ppm)	Assignment
0.9 to 1.7	broad peak → impurity
1.8 to 1.9	m:2H, -CH <sub>2</sub> → d
2.4 to 2.6	m:4H, -CH <sub>2</sub> × 2 → b, e
5.2 to 5.3	m:1H, -OCH → c
6.9 to 7.0	t:2H, proton at the ortho position of the F group → h, j
7.1	t:2H, proton at the para position of the F group → g, k
7.3	s:solvent (CDCl <sub>3</sub> )

m:multiplet, t:triplet, s:singlet

Table 10 (Results of  $^{13}\text{C}$  spectrometry )  
Resonance frequency: 100MHz

$\delta$ (ppm)	Assignment
31.0	$-\text{CH}_2 \rightarrow \text{d}$
35.9	$-\text{CH}_2 \rightarrow \text{e}$
39.4	$-\text{CH}_2 \rightarrow \text{b}$
70.5	$-\text{CH} \rightarrow \text{c}$
77.1 to 77.7	Solvent ( $\text{CDCl}_3$ )
115.5, 115.7	$-\text{CH}$ at the ortho position of the F group $\rightarrow \text{h, j}$
130.0	$-\text{CH}$ at the meta position of the F group $\rightarrow \text{g, k}$
136.8	C at the para position of the F group $\rightarrow \text{f}$
160.5, 163.0	$-\text{C}$ at the F substituent position $\rightarrow \text{i}$
169.6	carbonyl group $-\text{C}=\text{O} \rightarrow \text{a}$

Table 11  
(Results of  $^1\text{H}$ -NMR spectrum identification (see Fig. 4))

Chemical Shift /ppm	Integral /H	type	Identification
1.85	4	m	c, d
2.46	2	t	b
3.95	2	t	e
6.83	2	m	h, j
6.97	2	t	g, k
10.15		broad	OH

m:multiple, t:triplet, d:doublet

Table 12 (Various microorganisms and the yields of produced PHA)

	CDW(mg/L)	PDW(mg/L)	PDW/CDW(%)
strain P91 (Example 8)	650	50	7.7
strain YN2 1 (Example 9)	1250	755	60.4
strain P161 (Example 10)	1150	680	59.1
strain H45 (Example 11)	1150	600	52.2
strain YN2 2 (Example 12)	500	240	48.0

Table 13 (Molecular weight of PHA produced by each microorganism)

	Mn ( $\times 10^4$ )	Mw ( $\times 10^5$ )	Mw/Mn
strain P91 (Example 8)	5.1	1.0	2.0
strain YN2 1 (Example 9)	8.8	2.4	2.7
strain P161 (Example 10)	6.8	1.8	2.7
strain H45 (Example 11)	8.8	2.2	2.5
strain YN2 2 (Example 12)	5.7	1.4	2.5

Table 14

Dried cell (mg/L)	Dried polymer (mg/L)	Yield (polymer/cell, %)
850	110	12.9

Table 15

3-hydroxyvaleric acid	1.4%
3-hydroxyheptanoic acid	29.3%
3-hydroxyoctanoic acid	3.2%
3-hydroxynonanoic acid	64.6%
3-hydroxy-5-(4-trifluomethylphenyl) valeric acid	1.5%

Table 16

Dried cell (mg/L)	Dried polymer (mg/L)	Yield (polymer/cell, %)
720	29	4.0

Table 17

3-hydroxyvaleric acid	0.6%
3-hydroxyheptanoic acid	21.5%
3-hydroxyoctanoic acid	4.0%
3-hydroxynonanoic acid	70.5%
3-hydroxydecanoic acid	1.1%
3-hydroxy-5-(4-trifluomethylphenyl) valeric acid	2.3%

Table 18

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	1300	1000
Polymer (Dry Weight) (mg/L)	945	570
Polymer (Dry Weight) / Cell (Dry Weight)	73%	57%
Monomer Unit Composition (area ratio)		
3-hydroxybutyric acid	0%	1%
3-hydroxyvaleric acid	0%	1%
3-hydroxyhexanoic acid	0%	0%
3-hydroxyheptanoic acid	0%	14%
3-hydroxyoctanoic acid	1%	2%
3-hydroxynonanoic acid	0%	70%
3-hydroxydecanoic acid	2%	0%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	0%	0%
3-hydroxy-5-phenylvaleric acid	97%	12%

Table 19

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	750	800
Polymer (Dry Weight) (mg/L)	400	385
Polymer (Dry Weight) / Cell (Dry Weight)	53%	48%
Monomer Unit Composition (area ratio)		
3-hydroxybutyric acid	0%	0%
3-hydroxyvaleric acid	0%	0%
3-hydroxyhexanoic acid	0%	0%
3-hydroxyheptanoic acid	0%	14%
3-hydroxyoctanoic acid	0%	0%
3-hydroxynonanoic acid	0%	76%
3-hydroxydecanoic acid	0%	0%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	0%	0%
3-hydroxy-5-phenylvaleric acid	100%	10%

Table 20

Carbon Source for growth	D-mannose	D-fructose
Cell (Dry Weight) (mg/L)	780	760
Polymer (Dry Weight) (mg/L)	452	418
Polymer (Dry Weight) / Cell (Dry Weight)	58%	55%
Monomer Unit Composition (area ratio)		
3-hydroxybutyric acid	0%	0%
3-hydroxyvaleric acid	0%	0%
3-hydroxyhexanoic acid	0%	0%
3-hydroxyheptanoic acid	0%	0%
3-hydroxyoctanoic acid	2%	1%
3-hydroxynonanoic acid	0%	0%
3-hydroxydecanoic acid	0%	1%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	0%	0%
3-hydroxy-5-phenylvaleric acid	98%	98%

Table 21

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	1150	1180
Polymer (Dry Weight) (mg/L)	830	752
Polymer (Dry Weight) / Cell (Dry Weight)	72%	64%
Monomer Unit Composition (area ratio)		
3-hydroxybutyric acid	0%	2%
3-hydroxyvaleric acid	0%	1%
3-hydroxyhexanoic acid	0%	0%
3-hydroxyheptanoic acid	0%	17%
3-hydroxyoctanoic acid	1%	3%
3-hydroxynonanoic acid	0%	44%
3-hydroxydecanoic acid	3%	0%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	0%	0%
3-hydroxy-5-phenylvaleric acid	96%	33%

Table 22

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	1250	920
Polymer (Dry Weight) (mg/L)	900	543
Polymer (Dry Weight) / Cell (Dry Weight)	72%	59%

Monomer Unit Composition (area ratio)

3-hydroxybutyric acid	0%	0%
3-hydroxyvaleric acid	0%	0%
3-hydroxyhexanoic acid	0%	0%
3-hydroxyheptanoic acid	0%	11%
3-hydroxyoctanoic acid	1%	0%
3-hydroxynonanoic acid	0%	80%
3-hydroxydecanoic acid	2%	0%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	0%	0%
3-hydroxy-5-(4-fluorophenyl)valeric acid	97%	9%

Table 23

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	1100	900
Polymer (Dry Weight) (mg/L)	143	119
Polymer (Dry Weight) / Cell (Dry Weight)	13%	13%

Monomer Unit Composition (area ratio)

3-hydroxybutyric acid	2%	2%
3-hydroxyvaleric acid	0%	1%
3-hydroxyhexanoic acid	1%	0%
3-hydroxyheptanoic acid	0%	18%
3-hydroxyoctanoic acid	1%	3%
3-hydroxynonanoic acid	0%	48%
3-hydroxydecanoic acid	0%	0%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	0%	0%
3-hydroxy-6-phenylhexanoic acid	96%	28%



Table 24

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	685	440
Polymer (Dry Weight) (mg/L)	137	263
Polymer (Dry Weight) / Cell (Dry Weight)	20%	60%

Monomer Unit Composition (area ratio)

3-hydroxybutyric acid	0%	0%
3-hydroxyvaleric acid	0%	1%
3-hydroxyhexanoic acid	0%	1%
3-hydroxyheptanoic acid	0%	30%
3-hydroxyoctanoic acid	3%	4%
3-hydroxynonanoic acid	0%	62%
3-hydroxydecanoic acid	4%	1%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	1%	1%
3-hydroxy-4-phenoxybutyric acid	92%	0%

Table 25

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	450	340
Polymer (Dry Weight) (mg/L)	18	216
Polymer (Dry Weight) / Cell (Dry Weight)	4%	64%

Monomer Unit Composition (area ratio)

3-hydroxybutyric acid	0%	0%
3-hydroxyvaleric acid	0%	1%
3-hydroxyhexanoic acid	1%	0%
3-hydroxyheptanoic acid	0%	28%
3-hydroxyoctanoic acid	5%	4%
3-hydroxynonanoic acid	0%	67%
3-hydroxydecanoic acid	5%	0%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	1%	0%
3-hydroxy-4-phenoxybutyric acid	88%	0%

Table 26

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	600	400
Polymer (Dry Weight) (mg/L)	51	144
Polymer (Dry Weight) / Cell (Dry Weight)	9%	36%
Monomer Unit Composition (area ratio)		
3-hydroxybutyric acid	3%	0%
3-hydroxyvaleric acid	0%	1%
3-hydroxyhexanoic acid	1%	1%
3-hydroxyheptanoic acid	0%	26%
3-hydroxyoctanoic acid	9%	5%
3-hydroxynonanoic acid	0%	63%
3-hydroxydecanoic acid	11%	2%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	0%	0%
3-hydroxy-4-phenoxybutyric acid	76%	2%

Table 27

Carbon Source for growth	D-glucose	n-nonanoic acid
Cell (Dry Weight) (mg/L)	1150	900
Polymer (Dry Weight) (mg/L)	590	420
Polymer (Dry Weight) / Cell (Dry Weight)	51%	47%
Monomer Unit Composition (area ratio)		
3-hydroxybutyric acid	1%	0%
3-hydroxyvaleric acid	0%	0%
3-hydroxyhexanoic acid	0%	0%
3-hydroxyheptanoic acid	0%	13%
3-hydroxyoctanoic acid	0%	5%
3-hydroxynonanoic acid	0%	69%
3-hydroxydecanoic acid	10%	0%
3-hydroxyundecanoic acid	0%	0%
3-hydroxydodecanoic acid	0%	0%
3-hydroxy-5-phenylvaleric acid	89%	13%

Table 28

	CDW (mg/L)	PDW (mg/L)	Yield (%)
(1) YE	1225	488	39.8
(2) BE	600	185	30.8
(3) CA	950	445	46.8
(4) PP	1200	755	62.9

CDW: Cell (Dry Weight) (mg/L)  
 PDW: Polymer (Dry Weight) (mg/L)  
 Yield: PDW/CDW (%)

Table 29

	CDW (mg/L)	PDW (mg/L)	Yield (%)
(1) YE	1100	225	20.5
(2) PP	1200	850	70.8

CDW: Cell (Dry Weight) (mg/L)  
 PDW: Polymer (Dry Weight) (mg/L)  
 Yield: PDW/CDW (%)

Table 30

	CDW (mg/L)	PDW (mg/L)	Yield (%)
(1) YE	1050	205	19.5
(2) PP	1000	345	34.5

CDW: Cell (Dry Weight) (mg/L)  
 PDW: Polymer (Dry Weight) (mg/L)  
 Yield: PDW/CDW (%)

Table 31

	CDW (mg/L)	PDW (mg/L)	Yield (%)
(1) YE	750	220	29.3
(2) SG	700	260	37.1
(3) CA	900	340	37.7
(4) PP	1100	450	40.9

CDW: Cell (Dry Weight) (mg/L)

PDW: Polymer (Dry Weight) (mg/L)

Yield: PDW/CDW (%)

Table 32

	CDW (mg/L)	PDW (mg/L)	Yield (%)
(1) YE	950	325	34.2
(2) SG	750	240	32.0
(3) BE	450	130	28.9
(4) PP	1000	450	45.0

CDW: Cell (Dry Weight) (mg/L)

PDW: Polymer (Dry Weight) (mg/L)

Yield: PDW/CDW (%)

Table 33

Identification results of the  $^1\text{H}$ -NMR spectral patterns (cf. Fig. 14)

Chemical shift (ppm)	Identification
2.80	b1
4.24	d1
5.50	c1
7.00	f1, j1
8.20	g1, l1

Table 34

Cell (Dry Weight)	280 mg/L
Polymer (Dry Weight)	30 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	10.7%
Ratio of monomer unit determined by NMR (mole ratio)	
3HNO <sub>2</sub> PxB contained in PHA	3.3 mol%
Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	9.3%
3-hydroxyhexanoic acid	2.0%
3-hydroxyoctanoic acid	12.2%
3-hydroxydecanoic acid	47.9%
3-hydroxydodecanoic acid	15.7%
3-hydroxydodecenoic acid	12.9%

Table 35

Cell (Dry Weight)	250 mg/L
Polymer (Dry Weight)	30 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	12.0%
Ratio of monomer unit determined by NMR (mole ratio)	
3HNO <sub>2</sub> PxB contained in PHA	4.3 mol%
Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	1.3%
3-hydroxyhexanoic acid	0.0%
3-hydroxyoctanoic acid	12.6%
3-hydroxydecanoic acid	38.8%
3-hydroxydodecanoic acid	22.7%
3-hydroxydodecenoic acid	24.6%

Table 36

Cell (Dry Weight)	785 mg/L
Polymer (Dry Weight)	55 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	7.0%
Ratio of monomer unit determined by NMR (mole ratio)	
3HNO <sub>2</sub> PxB contained in PHA	3.8 mol%
Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	2.9%
3-hydroxyhexanoic acid	1.5%
3-hydroxyoctanoic acid	12.1%
3-hydroxydecanoic acid	40.0%
3-hydroxydodecanoic acid	14.7%
3-hydroxydodecenoic acid	28.8%

Table 37

Identification results of <sup>1</sup>H-NMR  
spectral patterns (cf. Fig. 15)

Chemical shift (ppm)	Identification
2.79	b1
4.18	d1
5.51	c1
6.98	f1, j1
7.58	g1, l1

Table 38

PHA production by *Pseudomonas cichorii* YN2

Cell (Dry Weight)	900 mg/L
Polymer (Dry Weight)	180 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	20.0%
Ratio of monomer unit determined by NMR (mole ratio)	
3HCNPxBA contained in PHA	4.1 mol%
Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	9.3%
3-hydroxyhexanoic acid	2.0%
3-hydroxyoctanoic acid	12.2%
3-hydroxydecanoic acid	47.9%
3-hydroxydodecanoic acid	15.7%
3-hydroxydodecenoic acid	12.9%

Table 39

PHA production by *Pseudomonas cichorii* H45

Cell (Dry Weight)	775 mg/L
Polymer (Dry Weight)	150 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	19.4%
Ratio of monomer unit determined by NMR (mole ratio)	
3HCNPxBA contained in PHA	3.2 mol%
Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	70.5%
3-hydroxyhexanoic acid	1.0%
3-hydroxyoctanoic acid	10.3%
3-hydroxydecanoic acid	13.4%
3-hydroxydodecanoic acid	2.3%
3-hydroxydodecenoic acid	2.6%

Table 40

PHA production by *Pseudomonas cichorii* YN2

Cell (Dry Weight)	930 mg/L
Polymer (Dry Weight)	200 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	21.5%
Ratio of monomer unit determined by NMR (mole ratio)	
3HCNPxBA contained in PHA	4.5 mol%
Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	12.5%
3-hydroxyhexanoic acid	2.0%
3-hydroxyoctanoic acid	12.2%
3-hydroxydecanoic acid	47.3%
3-hydroxydodecanoic acid	15.2%
3-hydroxydodecenoic acid	10.8%

Table 41

PHA production by *Pseudomonas cichorii* H45

Cell (Dry Weight)	750 mg/L
Polymer (Dry Weight)	135 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	18.0%
Ratio of monomer unit determined by NMR (mole ratio)	
3HCNPxBA contained in PHA	4.4 mol%
Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	25.0%
3-hydroxyhexanoic acid	2.1%
3-hydroxyoctanoic acid	21.4%
3-hydroxydecanoic acid	37.3%
3-hydroxydodecanoic acid	6.0%
3-hydroxydodecenoic acid	8.2%



Table 42

PHA production by *Pseudomonas cichorii* YN2

---

Cell (Dry Weight)	1030 mg/L
Polymer (Dry Weight)	130 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	12.6%

---

Ratio of monomer unit determined by NMR (mole ratio)	
3HCNPxBA contained in PHA	7.2 mol%

---

Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	7.3%
3-hydroxyhexanoic acid	1.9%
3-hydroxyoctanoic acid	14.3%
3-hydroxydecanoic acid	48.2%
3-hydroxydodecanoic acid	12.8%
3-hydroxydodecenoic acid	15.5%

---

Table 43

PHA production by *Pseudomonas cichorii* H45

---

Cell (Dry Weight)	695 mg/L
Polymer (Dry Weight)	55 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	7.9%

---

Ratio of monomer unit determined by NMR (mole ratio)	
3HCNPxBA contained in PHA	2.6 mol%

---

Fatty acid-derived monomer unit composition (peak area ratio)	
3-hydroxybutyric acid	2.3%
3-hydroxyhexanoic acid	1.7%
3-hydroxyoctanoic acid	19.5%
3-hydroxydecanoic acid	52.1%
3-hydroxydodecanoic acid	10.3%
3-hydroxydodecenoic acid	14.1%

---

Table 44

$^1\text{H}$ -NMR Spectrum (cf. Fig. 16)

Chemical shift (ppm)	Integral value	type	Identification
2.11	2H	quint	$\text{CH}_2$ c
2.59	2H	t	$\text{CH}_2$ b
3.97	2H	t	$\text{CH}_2$ d
6.82	2H	m	g, i
6.95	2H	m	f, j
8.00 to 13.00	1H	br	OH

Table 45

$^{13}\text{C}$ -NMR Spectrum (cf. Fig. 17)

Chemical shift (ppm)	type	Identification
24.21	s	$\text{CH}_2$ c
30.39	s	$\text{CH}_2$ b
66.96	s	$\text{CH}_2$ d
115.23 & 115.31	d	f, j or g, i
115.56 & 115.79	d	f, j or g, i
154.70 & 154.71	d	e
155.97 & 158.33	d	h
179.40	s	$\text{C}=\text{O}$ a

Table 46

PHA production containing 3HpFPxB unit by strain YN2

Cell (Dry Weight)	885 mg/L
Polymer (Dry Weight)	220 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	24.9 %
Polymer Molecular Weight	Mn = 42,400 Mw = 90,600
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	1.8 %
3-hydroxyhexanoic acid	1.0 %
3-hydroxyoctanoic acid	5.4 %
3-hydroxydecanoic acid	10.4 %
3-hydroxydodecanoic acid	2.9 %
3-hydroxydodecenoic acid	5.9 %
3-hydroxy-4-(4-fluorophenoxy)butyric acid	72.6 %

Table 47

<sup>1</sup>H-NMR Spectrum (cf. Fig. 19)

Chemical shift (ppm)	Identification
2.76	2H, CH <sub>2</sub> b1
3.95 to 4.06	2H, CH <sub>2</sub> d1
5.46	1H, CH c1
6.71 to 6.90	4H, -C <sub>6</sub> H <sub>4</sub> - f1, g1, i1, j1

Table 48

Production of PHA containing 3HpFPxB unit  
by culturing strain H45

Cell (dry weight)	640 mg/L
Polymer (dry weight)	90 mg/L
Polymer (dry weight)/Cell (dry weight)	14.0%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	4.6%
3-hydroxyhexanoic acid	2.0%
3-hydroxyoctanoic acid	15.2%
3-hydroxydecanoic acid	19.8%
3-hydroxydodecanoic acid	4.0%
3-hydroxydodecenoic acid	7.4%
3-hydroxy-4-(4-fluorophenoxy)butyric acid	47.0%

Table 49

Production of PHA containing 3HpFPxB unit  
by culturing strain YN2

Cell (dry weight)	780 mg/L
Polymer (dry weight)	200 mg/L
Polymer (dry weight)/Cell (dry weight)	25.6%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	0.3%
3-hydroxyhexanoic acid	0.8%
3-hydroxyoctanoic acid	3.9%
3-hydroxydecanoic acid	6.7%
3-hydroxydodecanoic acid	2.1%
3-hydroxydodecenoic acid	3.7%
3-hydroxy-4-(4-fluorophenoxy)butyric acid	82.5%

Table 50

Production of PHA containing 3HpFPxB unit  
by culturing strain H45

Cell (dry weight)	590 mg/L
Polymer (dry weight)	45 mg/L
Polymer (dry weight)/Cell (dry weight)	7.6%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	0.2%
3-hydroxyhexanoic acid	1.9%
3-hydroxyoctanoic acid	11.9%
3-hydroxydecanoic acid	12.1%
3-hydroxydodecanoic acid	2.2%
3-hydroxydodecenoic acid	5.0%
3-hydroxy-4-(4-fluorophenoxy)butyric acid	66.7%

Table 51

Production of PHA containing 3HpFPxB unit  
by culturing strain YN2

Cell (dry weight)	960 mg/L
Polymer (dry weight)	155 mg/L
Polymer (dry weight)/Cell (dry weight)	16.1%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	0.5%
3-hydroxyhexanoic acid	1.0%
3-hydroxyoctanoic acid	8.9%
3-hydroxydecanoic acid	23.4%
3-hydroxydodecanoic acid	7.0%
3-hydroxydodecenoic acid	14.2%
3-hydroxy-4-(4-fluorophenoxy)butyric acid	45.0%

Table 52

Production of PHA containing 3HpFPxB unit  
by culturing strain H45

Cell (dry weight)	545 mg/L
Polymer (dry weight)	30 mg/L
Polymer (dry weight)/Cell (dry weight)	5.5%
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	2.3%
3-hydroxyhexanoic acid	1.6%
3-hydroxyoctanoic acid	15.6%
3-hydroxydecanoic acid	36.9%
3-hydroxydodecanoic acid	8.7%
3-hydroxydodecenoic acid	12.9%
3-hydroxy-4-(4-fluorophenoxy)butyric acid	22.0%

Table 53

<sup>1</sup>H-NMR Spectrum (cf. Fig. 20)

Chemical shift (ppm)	Integral value	type	Identification
2.11	2H	d, quint	CH <sub>2</sub> c
2.59	2H	t	CH <sub>2</sub> b
3.97	2H	t	CH <sub>2</sub> d
6.62	3H	m	h, i, j
7.21	1H	m	f
10.62	1H	br	OH

Table 54

$^{13}\text{C}$ -NMR Spectrum (cf. Fig. 21)

Chemical shift (ppm)	type	Identification
24.1	s	$\text{CH}_2$ c
30.34	s	$\text{CH}_2$ b
66.62	s	$\text{CH}_2$ d
101.23 & 102.19	d	f
107.39 & 107.60	d	h
110.08 & 110.11	d	j
130.04 & 130.14	d	i
159.92 & 160.03	d	e
162.29 & 164.73	d	g
179.19	s	$\text{C}=\text{O}$ a

Table 55

PHA production containing 3HmFPxB unit by strain YN2

Cell (Dry Weight)	745 mg/L
Polymer (Dry Weight)	80 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	10.7 %
Polymer Molecular Weight	Mn = 34,500 Mw = 75,200
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	1.2 %
3-hydroxyhexanoic acid	1.0 %
3-hydroxyoctanoic acid	6.5 %
3-hydroxydecanoic acid	9.5 %
3-hydroxydodecanoic acid	3.5 %
3-hydroxydodecenoic acid	5.9 %
3-hydroxy-4-(3-fluorophenoxy)butyric acid	72.5 %

Table 56

<sup>1</sup>H-NMR Spectrum (cf. Fig. 23)

Chemical shift (ppm)	Identification
2.75	2H, CH <sub>2</sub> b1
4.00	2H, CH <sub>2</sub> d1
5.48	1H, CH c1
6.52 to 6.62	3H, h1, i1, j1
7.26	3H, f1

Table 57

PHA production containing 3HmFPxB unit by strain H45

Cell (Dry Weight)	630 mg/L
Polymer (Dry Weight)	45 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	7.1 %
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	3.7 %
3-hydroxyhexanoic acid	2.2 %
3-hydroxyoctanoic acid	21.0 %
3-hydroxydecanoic acid	29.2 %
3-hydroxydodecanoic acid	8.2 %
3-hydroxydodecenoic acid	10.1 %
3-hydroxy-4-(3-fluorophenoxy)butyric acid	25.6 %



Table 58

PHA production containing 3HmFPxB unit by strain H45

Cell (Dry Weight)	515 mg/L
Polymer (Dry Weight)	40 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	7.8 %
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	3.0 %
3-hydroxyhexanoic acid	2.0 %
3-hydroxyoctanoic acid	16.8 %
3-hydroxydecanoic acid	16.8 %
3-hydroxydodecanoic acid	4.7 %
3-hydroxydodecenoic acid	7.4 %
3-hydroxy-4-(3-fluorophenoxy)butyric acid	49.3 %

Table 59

PHA production containing 3HmFPxB unit by strain YN2

Cell (Dry Weight)	900 mg/L
Polymer (Dry Weight)	90 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	10.0 %
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	20.1 %
3-hydroxyhexanoic acid	1.5 %
3-hydroxyoctanoic acid	9.8 %
3-hydroxydecanoic acid	15.5 %
3-hydroxydodecanoic acid	5.5 %
3-hydroxydodecenoic acid	9.7 %
3-hydroxy-4-(3-fluorophenoxy)butyric acid	37.9 %

Table 60

PHA production containing 3HmFPxB unit by strain H45

Cell (Dry Weight)	565 mg/L
Polymer (Dry Weight)	25 mg/L
Polymer (Dry Weight) / Cell (Dry Weight)	4.4 %
Monomer Unit Composition (area ratio)	
3-hydroxybutyric acid	4.2 %
3-hydroxyhexanoic acid	1.9 %
3-hydroxyoctanoic acid	17.8 %
3-hydroxydecanoic acid	38.0 %
3-hydroxydodecanoic acid	9.5 %
3-hydroxydodecenoic acid	13.8 %
3-hydroxy-4-(3-fluorophenoxy)butyric acid	14.8 %

Table 61

Cell (Dry Weight) (mg/L)	665
Polymer (Dry Weight) (mg/L)	105
Number Average Molecular Weight (Mn) $\times 10^4$	1.6
Weight Average Molecular Weight (Mw) $\times 10^4$	3.7
3-hydroxybutyric acid (%)	75.2
3-hydroxy-5-(4-fluorophenoxy)valeric acid (%)	24.8

Table 62

Cell (Dry Weight) (mg/L)	1120
Polymer (Dry Weight) (mg/L)	625
Number Average Molecular Weight (Mn) $\times 10^4$	4.8
Weight Average Molecular Weight (Mw) $\times 10^4$	9.9
3-hydroxybutyric acid (%)	17.4
3-hydroxy-5-(4-fluorophenyl)valeric acid (%)	82.6

Table 63

Cell (Dry Weight) (mg/L)	835
Polymer (Dry Weight) (mg/L)	395
Number Average Molecular Weight (Mn) $\times 10^4$	5.2
Weight Average Molecular Weight (Mw) $\times 10^4$	14.9
3-hydroxyoctanoic acid (%)	10.6
3-hydroxydecanoic acid (%)	9.5
3-hydroxy-5-(4-fluorophenoxy)valeric acid (%)	79.9

Table 64

Cell (Dry Weight) (mg/L)	1450
Polymer (Dry Weight) (mg/L)	1010
Number Average Molecular Weight (Mn) $\times 10^4$	6.0
Weight Average Molecular Weight (Mw) $\times 10^4$	14.8
3-hydroxyoctanoic acid (%)	2.8
3-hydroxydecanoic acid (%)	2.7
3-hydroxy-5-(4-fluorophenoxy)valeric acid (%)	94.5

Table 65

Cell (Dry Weight) (mg/L)	1605
Polymer (Dry Weight) (mg/L)	760
Number Average Molecular Weight (Mn) $\times 10^4$	4.6
Weight Average Molecular Weight (Mw) $\times 10^4$	12.6
3-hydroxyoctanoic acid (%)	0.5
3-hydroxydecanoic acid (%)	0.4
3-hydroxy-5-(4-fluorophenyl)valeric acid (%)	90.0
3-hydroxy-5-(4-fluorophenoxy)valeric acid (%)	9.1

Table 66

Cell (Dry Weight) (mg/L)	1200
Polymer (Dry Weight) (mg/L)	500
Number Average Molecular Weight (Mn) $\times 10^4$	2.2
Weight Average Molecular Weight (Mw) $\times 10^4$	4.9
3-hydroxybutyric acid (%)	3.4
3-hydroxyoctanoic acid (%)	0.3
3-hydroxydecanoic acid (%)	0.5
3-hydroxy-5-phenoxyvaleric acid (%)	34.1
3-hydroxy-7-phenoxyheptanoic (%)	51.1
3-hydroxy-9-phoxynonanoic acid (%)	10.6

Table 67

Cell (Dry Weight) (mg/L)	1305
Polymer (Dry Weight) (mg/L)	765
Number Average Molecular Weight (Mn) $\times 10^4$	5.0
Weight Average Molecular Weight (Mw) $\times 10^4$	11.6
3-hydroxybutyric acid (%)	0.1
3-hydroxyhexanoic acid (%)	0.3
3-hydroxyoctanoic acid (%)	3.0
3-hydroxydecanoic acid (%)	5.2
3-hydroxydodecanoic acid (%)	1.6
3-hydroxydodecenoic acid (%)	2.0
3-hydroxy-5-phenoxyvaleric acid (%)	40.7
3-hydroxy-7-phenoxyheptanoic (%)	40.4
3-hydroxy-9-phoxynonanoic acid (%)	6.7

Table 68

Cell (Dry Weight) (mg/L)	1085
Polymer (Dry Weight) (mg/L)	585
Number Average Molecular Weight (Mn) $\times 10^4$	4.0
Weight Average Molecular Weight (Mw) $\times 10^4$	8.9
3-hydroxybutyric acid (%)	0.4
3-hydroxyhexanoic acid (%)	0.3
3-hydroxyoctanoic acid (%)	3.2
3-hydroxydecanoic acid (%)	5.1
3-hydroxydodecanoic acid (%)	1.1
3-hydroxydodecenoic acid (%)	0.9
3-hydroxy-5-phenoxyvaleric acid (%)	43.0
3-hydroxy-7-phenoxyheptanoic acid (%)	43.9
3-hydroxy-9-phenoxynonanoic acid (%)	2.1

Table 69

Cell (Dry Weight) (mg/L)	1100
Polymer (Dry Weight) (mg/L)	440
Number Average Molecular Weight (Mn) $\times 10^4$	4.0
Weight Average Molecular Weight (Mw) $\times 10^4$	7.4
3-hydroxybutyric acid (%)	16.4
3-hydroxy-4-phenoxybutyric acid (%)	13.4
3-hydroxy-6-phenoxyhexanoic acid (%)	67.9
3-hydroxy-8-phenoxyoctanoic acid (%)	2.3

Table 70

Cell (Dry Weight) (mg/L)	860
Polymer (Dry Weight) (mg/L)	190
Number Average Molecular Weight (Mn) $\times 10^4$	3.4
Weight Average Molecular Weight (Mw) $\times 10^4$	6.8
3-hydroxybutyric acid (%)	0.2
3-hydroxy-4-phenoxybutyric acid (%)	5.1
3-hydroxy-6-phenoxyhexanoic acid (%)	82.9
3-hydroxy-8-phenoxyoctanoic acid (%)	11.8

Table 71

Cell (Dry Weight) (mg/L)	1405
Polymer (Dry Weight) (mg/L)	700
Number Average Molecular Weight (Mn) $\times 10^4$	4.9
Weight Average Molecular Weight (Mw) $\times 10^4$	10.7
3-hydroxybutyric acid (%)	4.8
3-hydroxyoctanoic acid (%)	1.2
3-hydroxydecanoic acid (%)	0.5
3-hydroxy-4-phenoxybutyric acid (%)	7.8
3-hydroxy-6-phenoxyhexanoic acid (%)	74.8
3-hydroxy-8-phenoxyoctanoic acid (%)	10.9

Table 72

Cell (Dry Weight) (mg/L)	1255
Polymer (Dry Weight) (mg/L)	560
Number Average Molecular Weight (Mn) $\times 10^4$	4.8
Weight Average Molecular Weight (Mw) $\times 10^4$	9.7
3-hydroxybutyric acid (%)	0.2
3-hydroxyhexanoic acid (%)	0.1
3-hydroxyoctanoic acid (%)	0.9
3-hydroxydecanoic acid (%)	0.9
3-hydroxydodecanoic acid (%)	0.1
3-hydroxydodecenoic acid (%)	0.2
3-hydroxy-4-phenoxybutyric acid (%)	2.5
3-hydroxy-6-phenoxyhexanoic acid (%)	82.8
3-hydroxy-8-phenoxyoctanoic acid (%)	12.3

Table 73

Cell (Dry Weight) (mg/L)	995
Polymer (Dry Weight) (mg/L)	505
Number Average Molecular Weight (Mn) $\times 10^4$	4.6
Weight Average Molecular Weight (Mw) $\times 10^4$	9.2
3-hydroxybutyric acid (%)	1.4
3-hydroxyoctanoic acid (%)	0.1
3-hydroxydecanoic acid (%)	0.2
3-hydroxy-5-phenoxyvaleric acid (%)	29.7
3-hydroxy-7-phenoxyheptanoic acid (%)	68.6

Table 74

Cell (Dry Weight) (mg/L)	815
Polymer (Dry Weight) (mg/L)	270
Number Average Molecular Weight (Mn) $\times 10^4$	3.3
Weight Average Molecular Weight (Mw) $\times 10^4$	6.4
3-hydroxybutyric acid (%)	1.2
3-hydroxy-5-phenoxyvaleric acid (%)	26.7
3-hydroxy-7-phenoxyheptanoic acid (%)	72.1

Table 75

Cell (Dry Weight) (mg/L)	1520
Polymer (Dry Weight) (mg/L)	860
Number Average Molecular Weight (Mn) $\times 10^4$	6.1
Weight Average Molecular Weight (Mw) $\times 10^4$	13.0
3-hydroxybutyric acid (%)	0.1
3-hydroxyhexanoic acid (%)	0.4
3-hydroxyoctanoic acid (%)	3.5
3-hydroxydecanoic acid (%)	4.1
3-hydroxydodecanoic acid (%)	1.1
3-hydroxydodecenoic acid (%)	3.1
3-hydroxy-5-phenoxyvaleric acid (%)	55.0
3-hydroxy-7-phenoxyheptanoic acid (%)	32.7

Table 76

Cell (Dry Weight) (mg/L)	1305
Polymer (Dry Weight) (mg/L)	685
Number Average Molecular Weight (Mn) $\times 10^4$	4.1
Weight Average Molecular Weight (Mw) $\times 10^4$	8.8
3-hydroxyhexanoic acid (%)	0.1
3-hydroxyoctanoic acid (%)	1.3
3-hydroxydecanoic acid (%)	1.8
3-hydroxydodecanoic acid (%)	0.4
3-hydroxydodecenoic acid (%)	0.6
3-hydroxy-5-phenoxyvaleric acid (%)	36.1
3-hydroxy-7-phenoxyheptanoic acid (%)	59.7

Table 77

Cell (Dry Weight) (mg/L)	890
Polymer (Dry Weight) (mg/L)	420
Number Average Molecular Weight (Mn) $\times 10^4$	9.7
Weight Average Molecular Weight (Mw) $\times 10^4$	29.7
3-hydroxybutyric acid (%)	0.2
3-hydroxyhexanoic acid (%)	0.3
3-hydroxyoctanoic acid (%)	2.4
3-hydroxydecanoic acid (%)	3.3
3-hydroxydodecanoic acid (%)	0.7
3-hydroxydodecenoic acid (%)	1.5
3-hydroxy-5-phenoxyvaleric acid (%)	91.6

Table 78

Cell (Dry Weight) (mg/L)	910
Polymer (Dry Weight) (mg/L)	390
Number Average Molecular Weight (Mn) $\times 10^4$	9.1
Weight Average Molecular Weight (Mw) $\times 10^4$	21.4
3-hydroxyhexanoic acid (%)	0.2
3-hydroxyoctanoic acid (%)	2.2
3-hydroxydecanoic acid (%)	4.9
3-hydroxydodecanoic acid (%)	0.8
3-hydroxydodecenoic acid (%)	1.5
3-hydroxy-5-phenoxyvaleric acid (%)	90.4

Table 79

Cell (Dry Weight) (mg/L)	1400
Polymer (Dry Weight) (mg/L)	935
Number Average Molecular Weight (Mn) $\times 10^4$	6.2
Weight Average Molecular Weight (Mw) $\times 10^4$	14.0
3-hydroxyoctanoic acid (%)	0.6
3-hydroxydecanoic acid (%)	0.7
3-hydroxy-5-phenylvaleric acid (%)	98.7



Table 80

Cell (Dry Weight) (mg/L)	1350
Polymer (Dry Weight) (mg/L)	955
Number Average Molecular Weight (Mn) $\times 10^4$	6.1
Weight Average Molecular Weight (Mw) $\times 10^4$	13.8
3-hydroxyoctanoic acid (%)	1.9
3-hydroxy-5-phenylvaleric acid (%)	98.1

Table 81

Cell (Dry Weight) (mg/L)	2050
Polymer (Dry Weight) (mg/L)	1310
Number Average Molecular Weight (Mn) $\times 10^4$	6.3
Weight Average Molecular Weight (Mw) $\times 10^4$	13.9
3-hydroxybutyric acid (%)	7.5
3-hydroxyoctanoic acid (%)	0.8
3-hydroxydecanoic acid (%)	0.8
3-hydroxy-5-phenylvaleric acid (%)	90.6

Table 82

Production of Polyhydroxyalkanoate by Using strain YN2

	Polymer Yield (mg/L)	3-hydroxy-4- phenoxy-n-butyric acid unit ratio
Disodium malate	20	96.6 %
Sodium L-glutamate	13	8.9 %
D(+)-glucose	8	98.4 %
n-nonanoic acid	440	ND
Polypeptone	17	43.5 %

\*GC-MS, TIC peak area ratio, ND not detected.

Table 83

Production of Polyhydroxyalkanoate by Using strain YN2

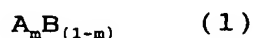
---

Cell (Dry Weight) (mg/L)	290
Polymer (Dry Weight) (mg/L)	20
Monomer Unit Composition (peak area ratio)	
3-hydroxybutyric acid	0.1 %
3-hydroxyhexanoic acid	0.2 %
3-hydroxyoctanoic acid	1.1 %
3-hydroxynonanoic acid	0.1 %
3-hydroxydecanoic acid	0.9 %
3-hydroxydodecanoic acid	0.2 %
3-hydroxydodecenoic acid	0.5 %
3-hydroxy-4-phenoxy-n-butyric acid	96.9 %

---

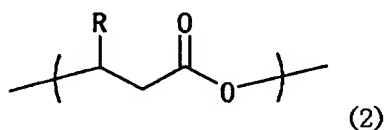
WHAT IS CLAIMED IS:

1. A polyhydroxyalkanoate having a monomer unit composition represented by General Formula (1):

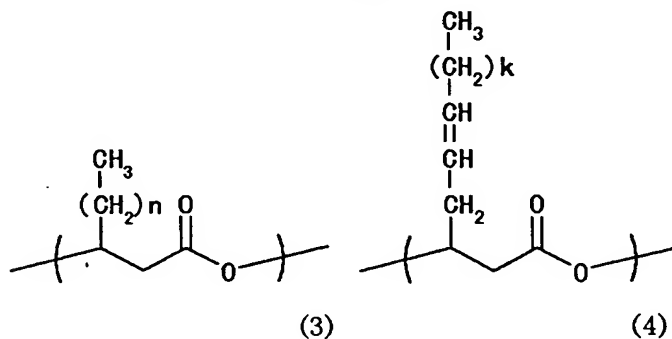


5 wherein A is represented by General Formula (2), B is at least one selected from the group consisting of monomer units represented by General Formula (3) or (4), and m has a value of 0.01 or larger and smaller than 1:

10



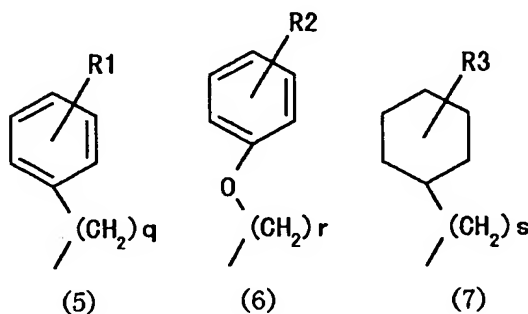
15



wherein

20 n has a value of 0 to 10, k has a value of 3 or 5, and R is at least one group selected from the group consisting of groups represented by General Formulae (5) to (7):

25



in Formula (5)

R1 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and q is an integer selected from 1 to 8;

5 in Formula (6)

R2 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and r is an integer selected from 1 to 8;

in Formula (7)

10 R3 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and s is an integer selected from 1 to 8;

provided that following R are excluded from the choice:

15 when selecting one type of group as R in the general formula (2):

groups of Formula (5) in which R1 is H and q = 2, R1 is H and q = 3, and R1 is -NO<sub>2</sub> and q = 2;

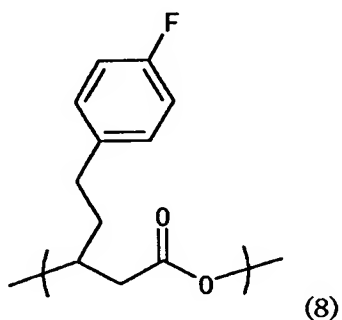
20 groups of Formula (6) in which R2 is a halogen atom and r = 2, provided that two components are selected as B from General Formula (3) or (4), R2 is -CN and r = 3, and R2 is -NO<sub>2</sub> and r = 3; and the groups of Formula (7) in which R3 is H and s = 1, and R3 is H and s = 2; and

25 when selecting two types of groups as R in General Formula (2), the combinations of two types of groups of Formula (6) in which R2 is a halogen atom and r = 2,

and R2 is a halogen atom and  $r = 4$ , provided that one component is selected as B from General Formula (3) or (4).

- 5            2. A polyhydroxyalkanoate comprising  
3-hydroxy-5-(4-fluorophenyl)valeric acid of Formula (8)  
as a monomer unit.

10

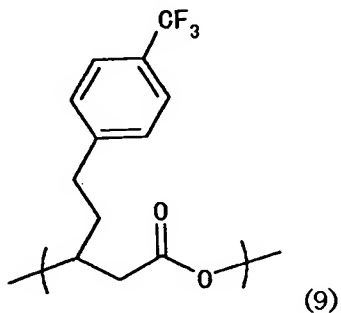


15

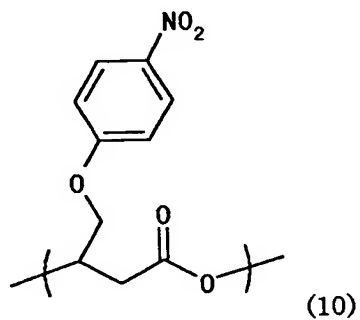
3. A polyhydroxyalkanoate comprising  
3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid of  
Formula (9) as a monomer unit.

20

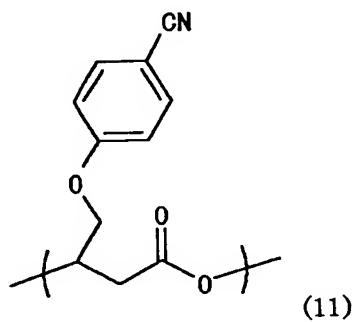
25



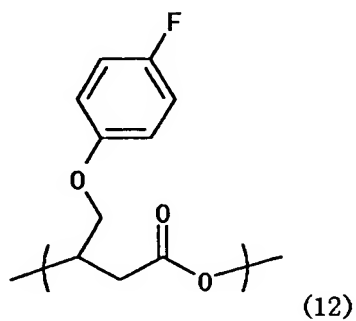
4. A polyhydroxyalkanoate comprising  
3-hydroxy-4-(4-nitrophenoxy)butyric acid of Formula  
(10) as a monomer unit.



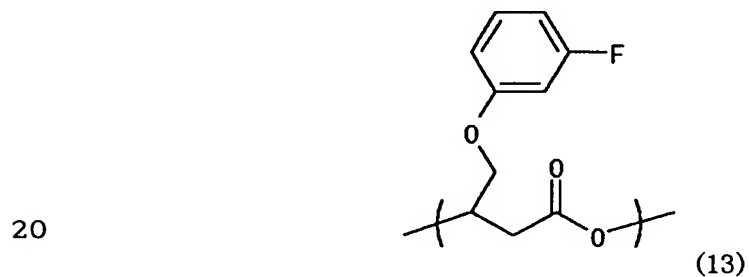
5. A polyhydroxyalkanoate comprising  
3-hydroxy-4-(4-cyanophenoxy)butyric acid of Formula  
15 (11) as a monomer unit.



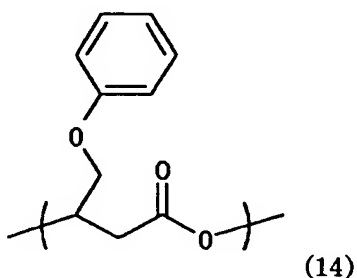
6. A polyhydroxyalkanoate comprising  
3-hydroxy-4-(4-fluorophenoxy)butyric acid of Formula  
(12) as a monomer unit.



7. A polyhydroxyalkanoate comprising  
3-hydroxy-4-(3-fluorophenoxy)butyric acid of Formula  
15 (13) as a monomer unit.

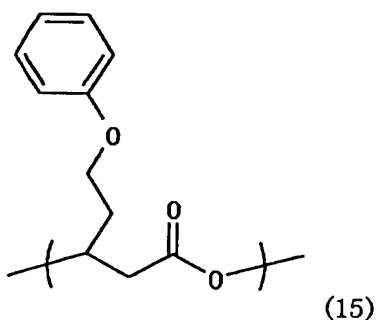


8. The polyhydroxyalkanoate according to claim 1, comprising 3-hydroxy-4-phenoxybutyric acid of Formula (14) as a monomer unit.



10

9. The polyhydroxyalkanoate according to claim 1, comprising 3-hydroxy-5-phenoxyvaleric acid of Formula (15) as a monomer unit.



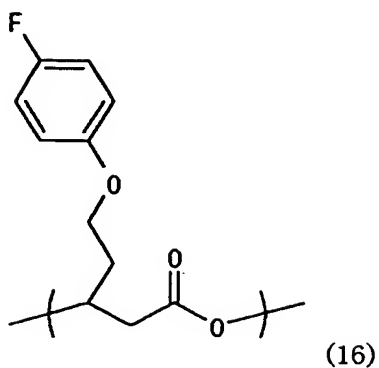
20

10. The polyhydroxyalkanoate according to claim 1, comprising 3-hydroxy-5-(4-fluorophenoxy)valeric acid of Formula (16) as a monomer unit, the monomer unit not being a 3-component-system.

25



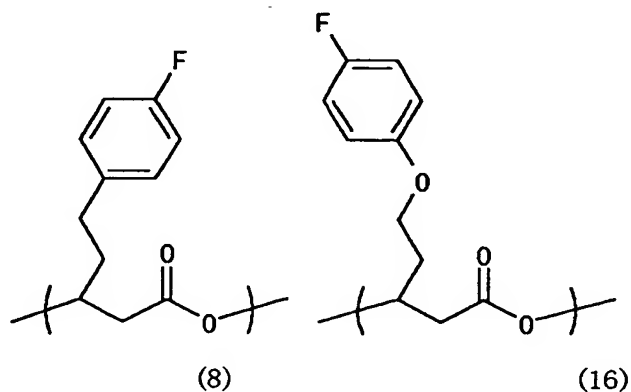
5



10

11. A polyhydroxyalkanoate comprising 3-hydroxy-5-(4-fluorophenyl)valeric acid of Formula (8) and 3-hydroxy-5-(4-fluorophenoxy)valeric acid of Formula (16) as monomer units.

15

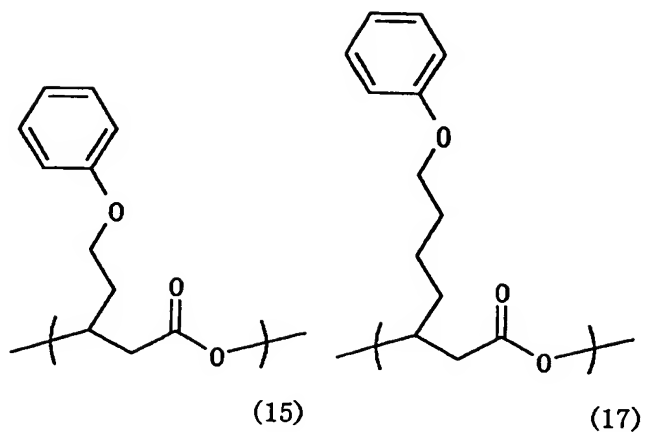


20

12. The polyhydroxyalkanoate according to claim 1, comprising 3-hydroxy-5-phenoxyvaleric acid of Formula (15) and 3-hydroxy-7-phenoxyheptanoic acid of Formula (17) as monomer units.

25

5

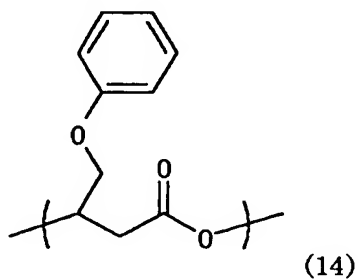


10

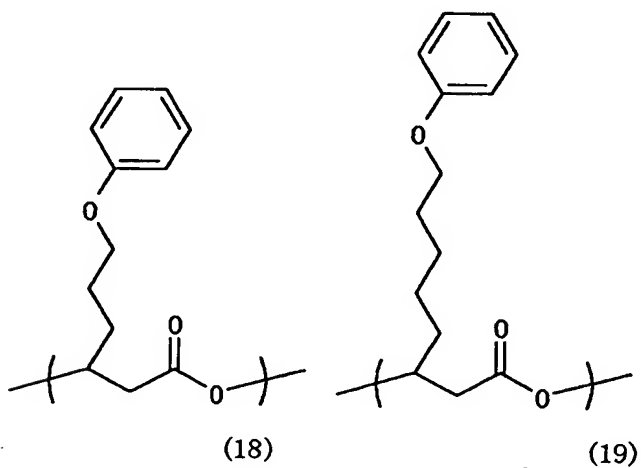
13. The polyhydroxyalkanoate according to claim 1, comprising 3-hydroxy-4-phenoxybutyric acid of Formula (14), 3-hydroxy-6-phenoxyhexanoic acid of Formula (18), and 3-hydroxy-8-phenoxyoctanoic acid of Formula (19) as monomer units.

15

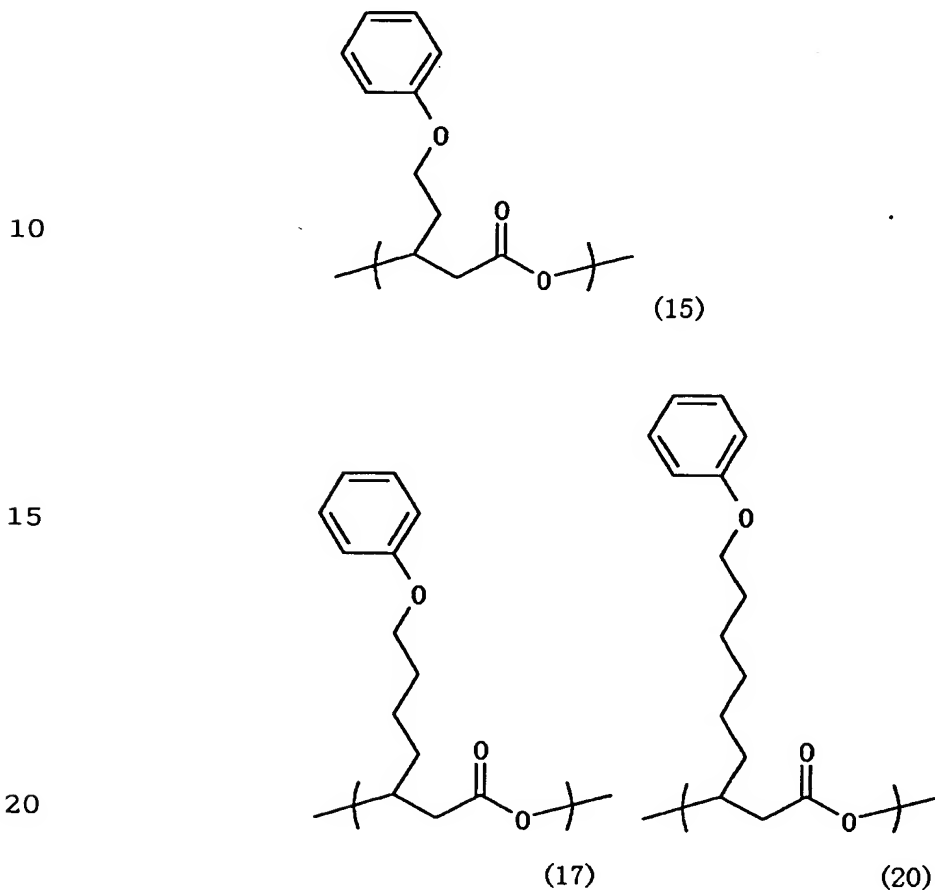
20



25

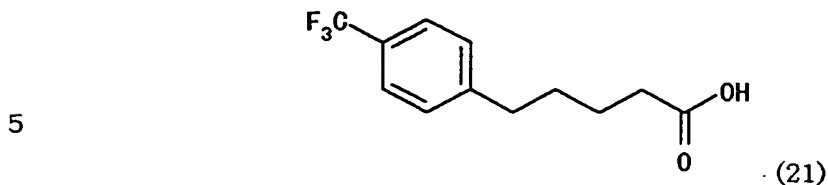


14. The polyhydroxyalkanoate according to claim  
1, comprising 3-hydroxy-5-phenoxyvaleric acid of  
Formula (15), 3-hydroxy-7-phenoxyheptanoic acid of  
Formula (17), and 3-hydroxy-9-phenoxynonanoic acid of  
5 Formula (20) as monomer units.



25 15. The polyhydroxyalkanoate according to any one  
of claims 1 to 14, wherein the number average molecular  
weight is 10000 to 200000.

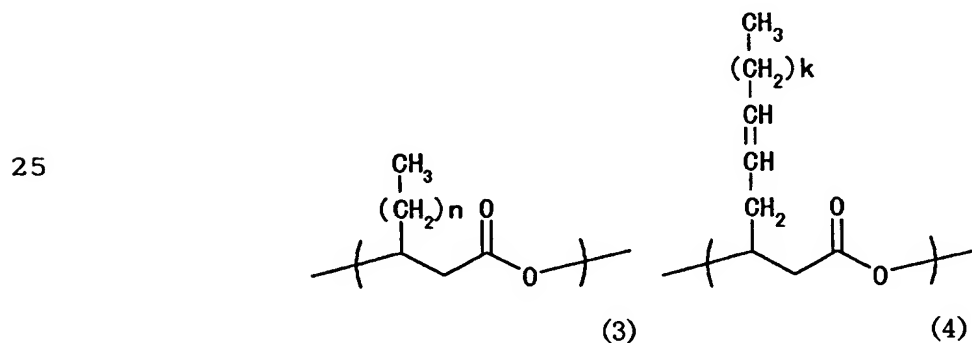
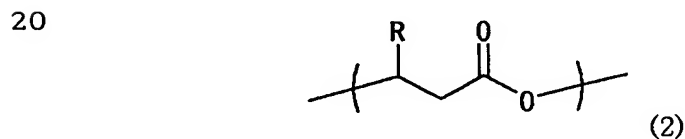
16. 5-(4-trifluoromethylphenyl)valeric acid of  
Formula 21.



17. A process of producing a  
polyhydroxyalkanoate, comprising a step of culturing a  
microorganism capable of synthesizing a  
polyhydroxyalkanoate of which monomer unit is  
represented by Formula (1) from an alkanoate in a  
medium containing the alkanoate:



wherein A is represented by General Formula (2), B  
is at least one selected from the group consisting of  
monomer units represented by General Formula (3) or  
(4), and m is 0.01 or larger and smaller than 1,



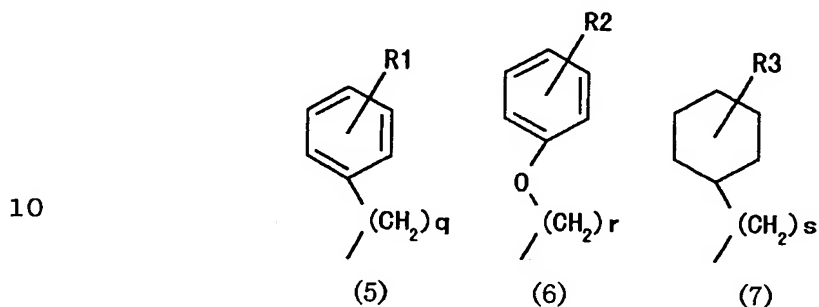
wherein

n is an integer selected from 0 to 10, k is 3 or 5, and

R is at least one group selected from the group

consisting of the groups represented by General

5 Formulae (5) to (7):



in Formula (5)

R1 is a group selected from the group consisting of a

15 hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub>  
and -C<sub>3</sub>F<sub>7</sub>; and q is an integer selected from 1 to 8;

in Formula (6)

R2 is a group selected from the group consisting of a

20 hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub>  
and -C<sub>3</sub>F<sub>7</sub>; and r is an integer selected from 1 to 8;

in Formula (7)

R3 is a group selected from the group consisting of a

hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub>  
and -C<sub>3</sub>F<sub>7</sub>; and s is an integer selected from 1 to 8;

25 provided that following R are excluded from the  
choice:

when selecting one type of group as R in General

Formula (2):

groups of Formula (5) in which R1 is H and q = 2,  
R1 is H and q = 3, and R1 is -NO<sub>2</sub> and q = 2;

the groups of Formula (6) in which R2 is a halogen atom  
5 and r = 2, R2 is -CN and r = 3, and R2 is -NO<sub>2</sub> and r =  
3; and

groups of Formula (7) in which R3 is H and s = 1,  
and R3 is H and s = 2; and

when selecting two types of groups as R in General  
10 Formula (2),

groups of Formula (6) in which R2 is a halogen  
atom and r = 2.

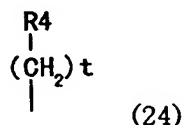
18. A process of producing polyhydroxyalkanoate,  
15 comprising a step of culturing a microorganism capable  
of producing the polyhydroxyalkanoate utilizing  
alkanoate in a medium containing the alkanoate and a  
saccharide.

20 19. The process according to claim 18, wherein  
the alkanoate is represented by Formula (22) and the  
polyhydroxyalkanoate comprises a monomer unit of  
Formula (23):



wherein

R is at least one group selected from the group consisting of groups of Formula (24):

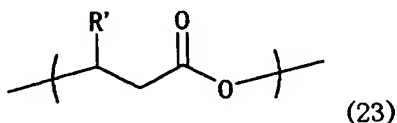


5

wherein

R<sub>4</sub> represents a substituted or unsubstituted phenyl group, a substituted or unsubstituted phenoxy group, or a substituted or unsubstituted cyclohexyl group, and t is an integer of 1 to 8 independently;

10



wherein R' is at least one group selected from the group consisting of the selected R and groups represented by Formula (24) where R<sub>4</sub> is the same as the selected R but t is shorter by multiply of 2 than t of the selected R and not 0.

15

20

20. The process according to claim 18, wherein the microorganism is cultured in one step in a medium containing the alkanoate of Formula (22) and a saccharide.

25

21. The process according to claim 18, wherein the microorganism is cultured in at least two steps: one is in a medium containing the alkanoate of Formula

(22) and a saccharide and the subsequent one is in a medium containing the alkanoate of Formula (22) and a saccharide with nitrogen source limitation.

5           22. The process according to claim 18, wherein the microorganism is cultured by inoculating the microorganism precultured in a medium containing a saccharide.

10           23. The process according to claim 18, wherein the saccharide is at least one selected from the group consisting of glucose, fructose and mannose.

15           24. A process of producing polyhydroxyalkanoate, comprising a step of culturing a microorganism capable of producing a polyhydroxyalkanoate utilizing an alkanoate in a medium containing the alkanoate and a polypeptone.

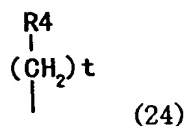
20           25. The process according to claim 24, wherein the alkanoate is represented by Formula (22) and the polyhydroxyalkanoate comprises a monomer unit of Formula (23):



wherein



R is at least one group selected from the group consisting of the groups of Formula (24):

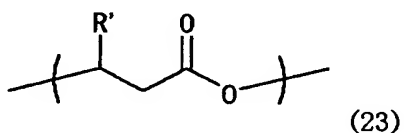


5

wherein

R<sub>4</sub> represents a substituted or unsubstituted phenyl group, a substituted or unsubstituted phenoxy group, or a substituted or unsubstituted cyclohexyl group, and t is an integer of 1 to 8 independently

10



wherein R' is at least one group selected from the group consisting of the selected R and groups represented by Formula (24) where R<sub>4</sub> is the same as the selected R but t is shorter by multiply of 2 than t of the selected R and not 0.

15

20            26. The process according to claim 24, wherein the microorganism is cultured in one step in a medium containing the alkanoate of Formula (22) and polypeptone.

25            27. The process according to claim 24, wherein the microorganism is cultured in at least two steps: one is in a medium containing an alkanoate represented

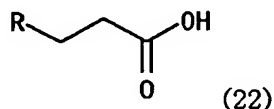
by Formula (22) and polypeptone, and the subsequent one is in a medium containing the alkanoate with nitrogen source limitation.

5           28. A process of producing polyhydroxyalkanoate comprising a step of culturing a microorganism capable of producing a polyhydroxyalkanoate utilizing an alkanoate in a medium containing the alkanoate and an organic acid involved in TCA cycle.

10

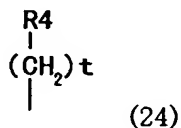
29. The process according to claim 28, wherein the alkanoate is represented by Formula (22) and the polyhydroxyalkanoate comprises a monomer unit represented by Formula (23):

15



wherein

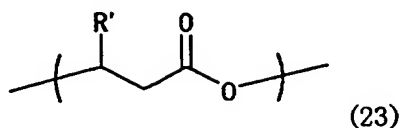
R is at least one group selected from the group  
20 consisting of groups of Formula (24):



wherein

25   R4 represents a substituted or unsubstituted phenyl group, a substituted or unsubstituted phenoxy group, or a substituted or unsubstituted cyclohexyl group, and t

is an integer of 1 to 8 independently;



5            wherein R' is at least one group selected from the  
group consisting of the selected R and groups  
represented by Formula (24) where R<sub>4</sub> is the same as the  
selected R but t is shorter by multiply of 2 than t of  
the selected R and not 0.

10

30. The process according to claim 28, wherein  
the microorganism is cultured in one step in the medium  
containing the alkanoate of Formula (22) and an organic  
acid involved in TCA cycle.

15

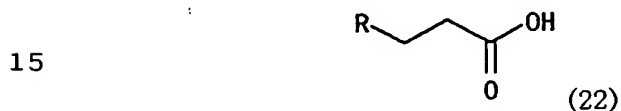
31. The process according to claim 28, wherein  
the microorganism is cultured in at least two steps:  
one is in a medium containing an alkanoate of Formula  
(22) and an organic acid involved in TCA cycle and the  
20 subsequent one is in a medium containing the alkanoate  
of Formula (22) and an organic acid involved in TCA  
cycle with nitrogen source limitation.

32. The process according to claim 28, wherein  
25 the organic acid involved in TCA cycle is at least one  
selected from the group consisting of lactic acid,  
pyruvic acid, citric acid, succinic acid, fumaric acid,

malic acid and salts thereof.

33. A process of producing polyhydroxyalkanoate, wherein the a microorganism is cultured in at least two steps: one is in a medium containing an alkanoate and a polypeptone and the subsequent one is in a medium containing the alkanoate and pyruvic acid or salt thereof with nitrogen source limitation.

34. The process according to claim 33, wherein the alkanoate is represented by Formula (22) and the polyhydroxyalkanoate comprises a monomer unit of Formula (23):



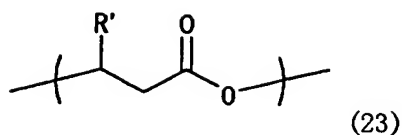
wherein

R is at least one group selected from the group consisting of the groups of Formula (24):



wherein

R4 represents a substituted or unsubstituted phenyl group, a substituted or unsubstituted phenoxy group, or a substituted or unsubstituted cyclohexyl group, and t is an integer of 1 to 8 independently;



wherein R' is at least one group selected from the  
5 group consisting of the selected R and groups  
represented by Formula (24) where R<sub>4</sub> is the same as the  
selected R but t is shorter by multiply of 2 than t of  
the selected R and not 0.

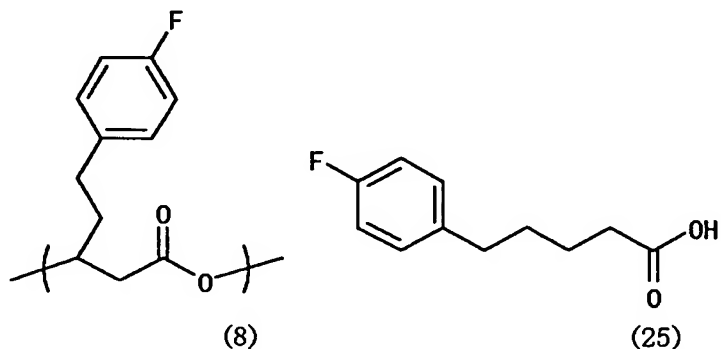
10 35. The process of producing polyhydroxyalkanoate  
according to any one of claims 17, 18, 24, 28 and 33,  
wherein the microorganism belongs to *Pseudomonas* sp. ✓

15 36. The process according to claim 35, wherein  
the microorganism is at least one strain selected from  
the group consisting of *Pseudomonas cichorii* YN2, FERM  
BP-7375; *Pseudomonas cichorii* H45, FERM BP-7374;  
*Pseudomonas putida* P91, FERM BP-7373; and *Pseudomonas*  
*jessenii* P161, FERM BP-7376.

20 37. The process of producing polyhydroxyalkanoate  
comprising a monomer unit of Formula (8) according to  
any one of claims 17, 18, 24, 28 and 33, comprising a  
step of culturing a microorganism capable of producing  
25 polyhydroxyalkanoate comprising a monomer unit of  
3-hydroxy-5-(4-fluorophenyl)valeric acid utilizing  
5-(4-fluorophenyl)valeric acid in a medium containing

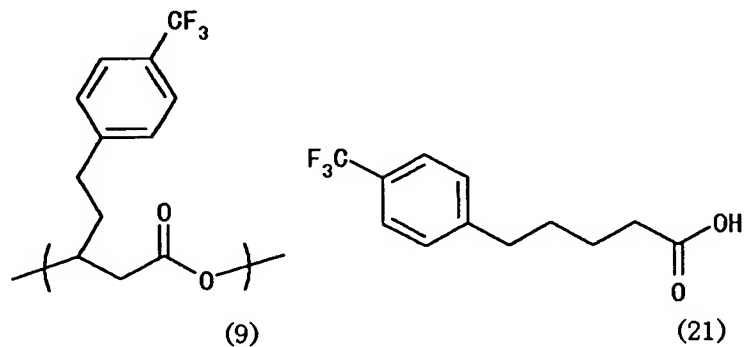
5-(4-fluorophenyl)valeric acid of Formula (25).

5



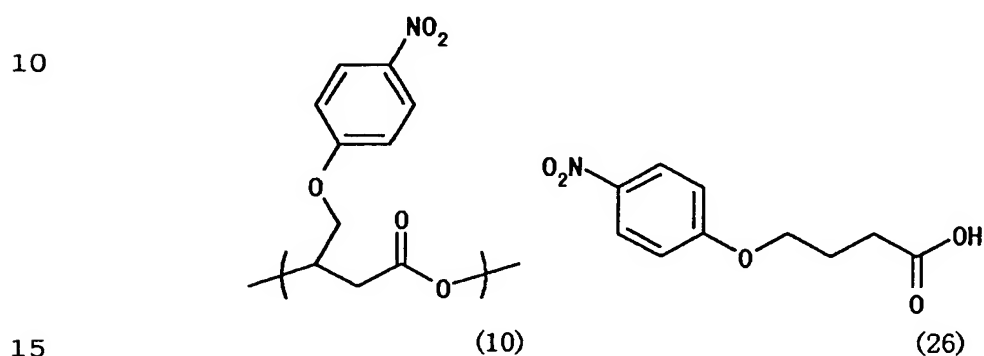
10            38. The process of producing polyhydroxyalkanoate  
comprising a monomer unit of Formula (9) according to  
any one of claims 17, 18, 24, 28 and 33, comprising a  
step of culturing a microorganism capable of producing  
polyhydroxyalkanoate comprising a monomer unit of  
15    3-hydroxy-5-(4-trifluoromethylphenyl)valeric acid  
utilizing 5-(4-trifluoromethylphenyl)valeric acid in a  
medium containing 5-(4-trifluoromethylphenyl)valeric  
acid of Formula (21).

20



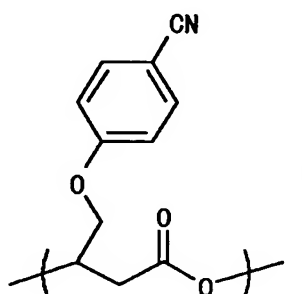
25

39. The process of producing polyhydroxyalkanoate comprising a monomer unit of Formula (10) according to any one of claims 17, 18, 24, 28 and 33, comprising a step of culturing a microorganism capable of producing polyhydroxyalkanoate comprising a monomer unit of 3-hydroxy-4-(4-nitrophenoxy)butyric acid utilizing 4-(4-nitrophenoxy)butyric acid in a medium containing 4-(4-nitrophenoxy)butyric acid of Formula (26).

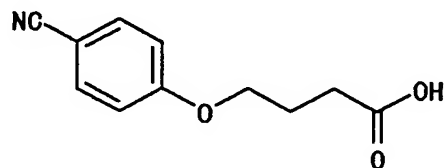


40. The process of producing polyhydroxyalkanoate comprising a monomer unit of Formula (11) according to any one of claims 17, 18, 24, 28 and 33, comprising a step of culturing a microorganism capable of producing polyhydroxyalkanoate comprising a monomer unit of 3-hydroxy-4-(4-cyanophenoxy)butyric acid utilizing 4-(4-cyanophenoxy)butyric acid in a medium containing 4-(4-cyanophenoxy)butyric acid of Formula (27).

5



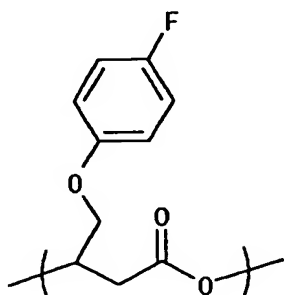
(11)



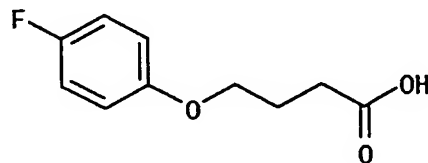
(27)

41. The process of producing polyhydroxyalkanoate  
 10 comprising a monomer unit of Formula (12) according to  
 any one of claims 17, 18, 24, 28 and 33, comprising a  
 step of culturing a microorganism capable of producing  
 polyhydroxyalkanoate comprising a monomer unit of  
 3-hydroxy-4-(4-fluorophenoxy)butyric acid utilizing  
 15 4-(4-fluorophenoxy)butyric acid in a medium containing  
 4-(4-fluorophenoxy)butyric acid of Formula (28).

20



(12)

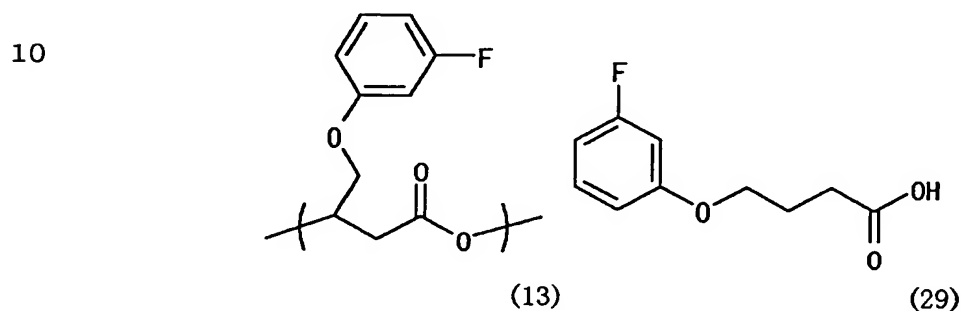


(28)

25



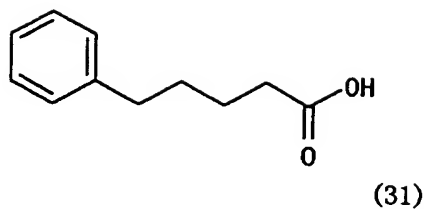
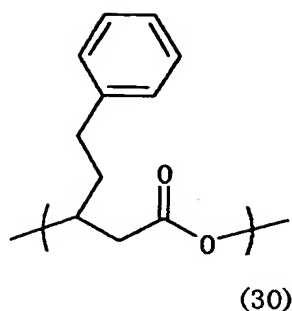
42. The process of producing polyhydroxyalkanoate comprising a monomer unit of Formula (13) according to any one of claims 17, 18, 24, 28 and 33, comprising a step of culturing a microorganism capable of producing polyhydroxyalkanoate comprising a monomer unit of 3-hydroxy-4-(3-fluorophenoxy)butyric acid utilizing 4-(3-fluorophenoxy)butyric acid in a medium containing 4-(3-fluorophenoxy)butyric acid of Formula (29).



43. The process of producing polyhydroxyalkanoate comprising a monomer unit of Formula (30) according to any one of claims 18, 24, 28 and 33, comprising a step of culturing a microorganism capable of producing polyhydroxyalkanoate comprising a monomer unit of 3-hydroxy-5-phenylvaleric acid utilizing 5-phenylvaleric acid in a medium containing 5-phenylvaleric acid of Formula (31).

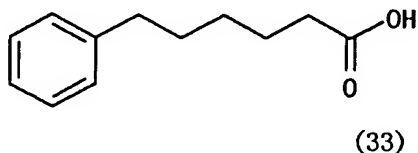
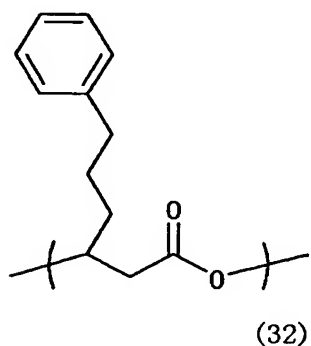
25

5



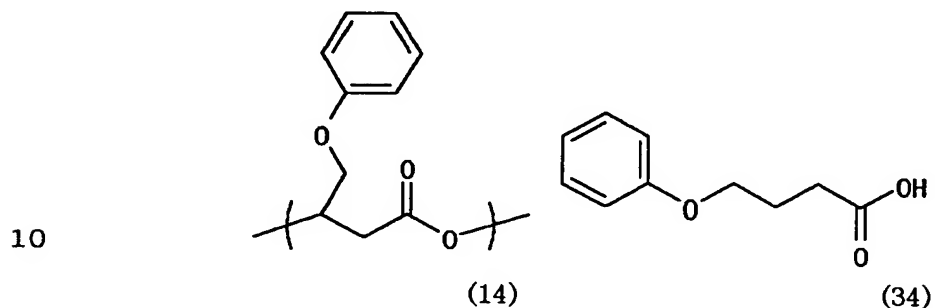
44. The process of producing polyhydroxyalkanoate  
 10 comprising a monomer unit of Formula (32) according to  
 any one of claims 18, 24, 28 and 33, comprising a step  
 of culturing a microorganism capable of producing  
 polyhydroxyalkanoate comprising a monomer unit of  
 3-hydroxy-6-phenylhexanoic acid utilizing  
 15 6-phenylhexanoic acid in a medium containing  
 6-phenylhexanoic acid of Formula (33).

20

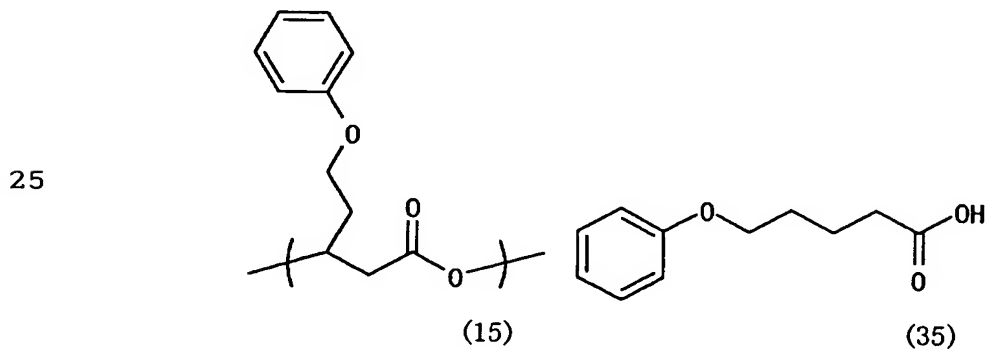


25 45. The process of producing polyhydroxyalkanoate  
 comprising a monomer unit of Formula (14) according to  
 any one of claims 17, 18, 24, 28 and 33, comprising a

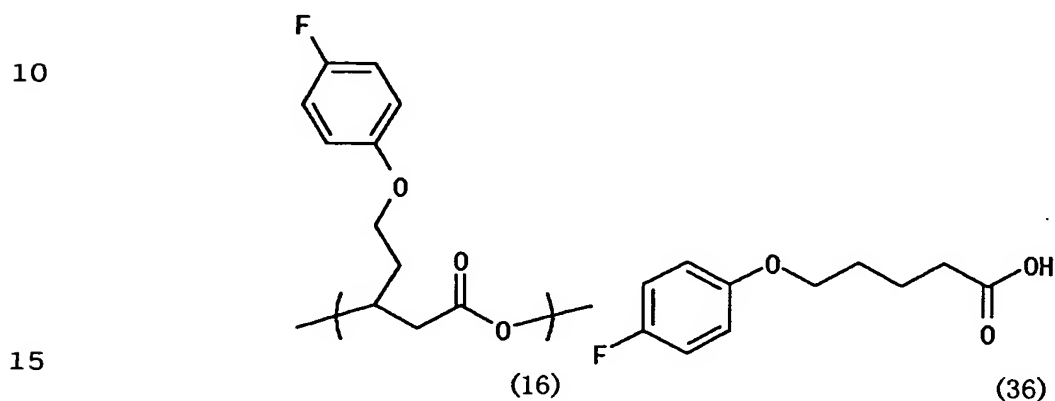
step of culturing a microorganism capable of producing  
a polyhydroxyalkanoate comprising a monomer unit of  
3-hydroxy-4-phenoxybutyric acid utilizing  
4-phenoxybutyric acid in a medium containing  
5 4-phenoxybutyric acid of Formula (34).



46. The process of producing polyhydroxyalkanoate  
15 comprising a monomer unit of Formula (15) according to  
any one of claims 17, 18, 24, 28 and 33, comprising a  
step of culturing a microorganism capable of producing  
polyhydroxyalkanoate comprising a monomer unit of  
3-hydroxy-5-phenoxyvaleric acid utilizing  
20 5-phenoxyvaleric acid in a medium containing  
5-phenoxyvaleric acid of Formula (35).

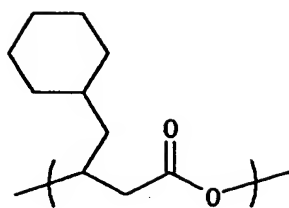


47. The process of producing polyhydroxyalkanoate comprising a monomer unit of Formula (16) according to any one of claims 18, 24, 28 and 33, comprising a step of culturing a microorganism capable of producing  
5 polyhydroxyalkanoate comprising a monomer unit of 3-hydroxy-5-(4-fluorophenoxy)valeric acid utilizing 5-(4-fluorophenoxy)valeric acid in a medium containing 5-(4-fluorophenoxy)valeric acid of Formula (36).

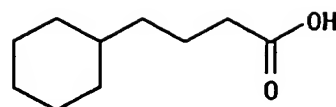


48. The process of producing polyhydroxyalkanoate comprising a monomer unit of Formula (37) according to any one of claims 18, 24, 28 and 33, comprising a step  
20 of culturing a microorganism capable of producing polyhydroxyalkanoate comprising a monomer unit of 3-hydroxy-4-cyclohexylbutyric acid utilizing 4-cyclohexylbutyric acid in a medium containing  
25 4-cyclohexylbutyric acid of Formula (38).

5



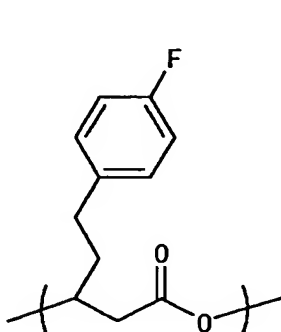
(37)



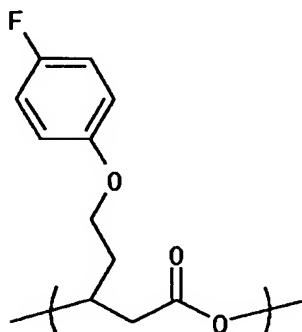
(38)

49. The process of producing polyhydroxyalkanoate comprising monomer units of Formulae (8) and (16) according to any one of claims 18, 24, 28 and 33, comprising a step of culturing a microorganism capable of producing polyhydroxyalkanoate comprising monomer units of 3-hydroxy-5-(4-fluorophenyl)valeric acid and 3-hydroxy-5-(4-fluorophenoxy)valeric acid utilizing 5-(4-fluorophenyl)valeric acid and 5-(4-fluorophenoxy)valeric acid in a medium containing 5-(4-fluorophenyl)valeric acid of Formula (25) and the 5-(4-fluorophenoxy)valeric acid of Formula (36).

20

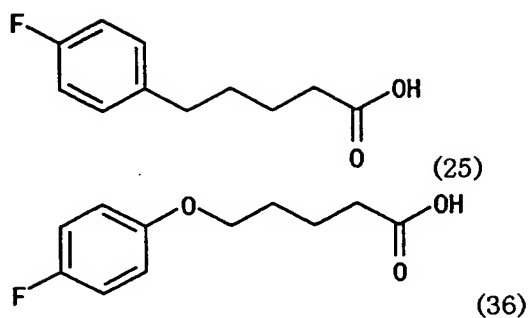


(8)

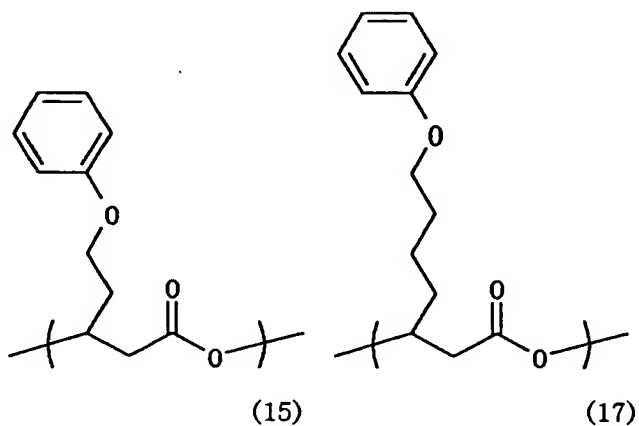


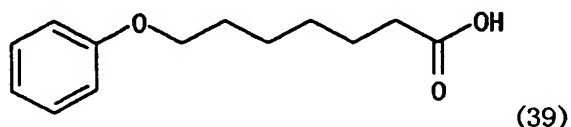
(16)

25



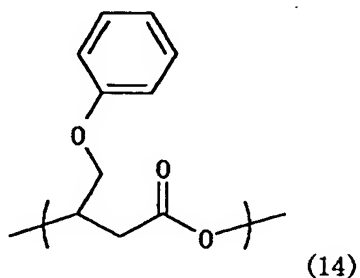
50. The process of producing polyhydroxyalkanoate comprising monomer units of Formulae (15) and (17) according to any one of claims 17, 18, 24, 28 and 33, comprising a step of culturing a microorganism capable of producing polyhydroxyalkanoate comprising monomer units of 3-hydroxy-5-phenoxyvaleric acid and 3-hydroxy-7-phenoxyheptanoic acid utilizing 7-phenoxyheptanoic acid in a medium containing 7-phenoxyheptanoic acid of Formula (39).





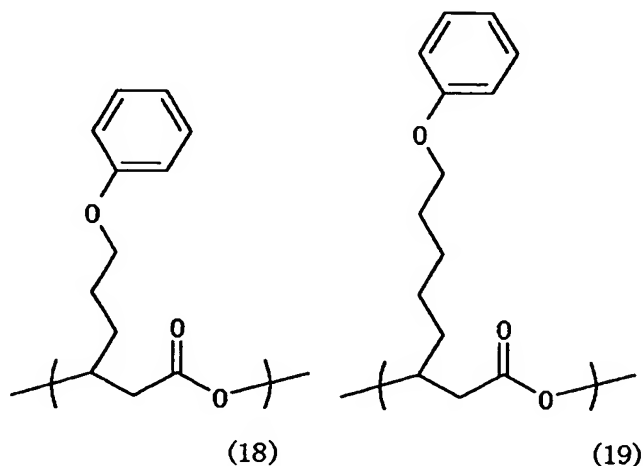
5            51. The process of producing polyhydroxyalkanoate  
 comprising monomer units of Formulae (14), (18) and  
 (19) according to any one of claims 17, 18, 24, 28 and  
 33, comprising a step of culturing a microorganism  
 capable of producing polyhydroxyalkanoate comprising  
 10 monomer units of 3-hydroxy-4-phenoxybutyric acid,  
 3-hydroxy-6-phenoxyhexanoic acid and  
 3-hydroxy-8-phenoxyoctanoic acid utilizing  
 8-phenoxyoctanoic acid in a medium containing  
 8-phenoxyoctanoic acid of Formula (40).

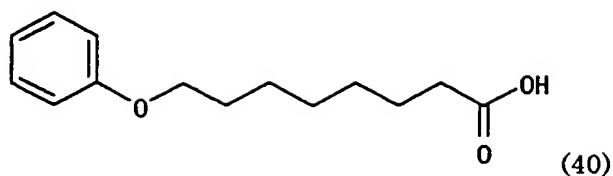
15



20

25

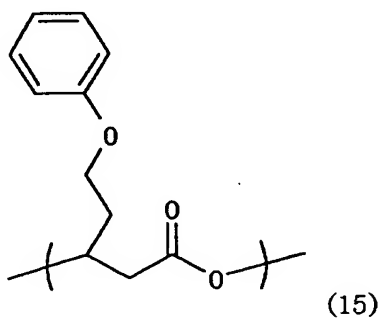




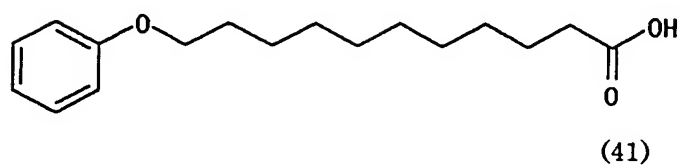
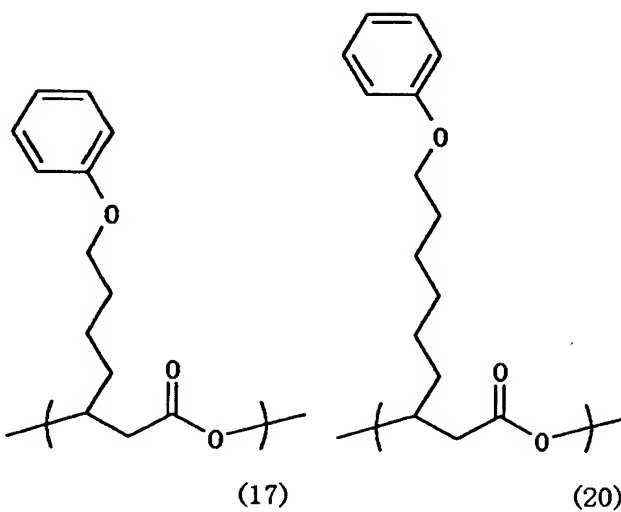
5

52. The process of producing polyhydroxyalkanoate comprising monomer units of Formulae (15), (17) and (20) according to any one of claims 17, 18, 24, 28 and 33, comprising a step of culturing a microorganism capable of producing polyhydroxyalkanoate comprising monomer units of 3-hydroxy-5-phenoxyvaleric acid, 3-hydroxy-7-phenoxyhexanoic acid and 3-hydroxy-9-phoxynonanoic acid utilizing 11-phenoxyundecanoic acid in a medium containing 11-phenoxyundecanoic acid of Formula (41).

20



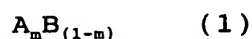




53. *Pseudomonas cichorii* H45, FERM BP-7374.
54. *Pseudomonas cichorii* YN2, FERM BP-7375.
55. *Pseudomonas putida* P91, FERM BP-7373.
56. *Pseudomonas jessenii* P161, FERM BP-7376.

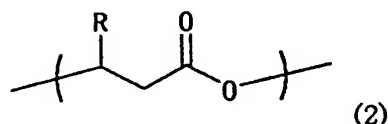
# ABSTRACT OF THE DISCLOSURE

A polyhydroxyalkanoate having a monomer unit composition represented by General Formula (1):

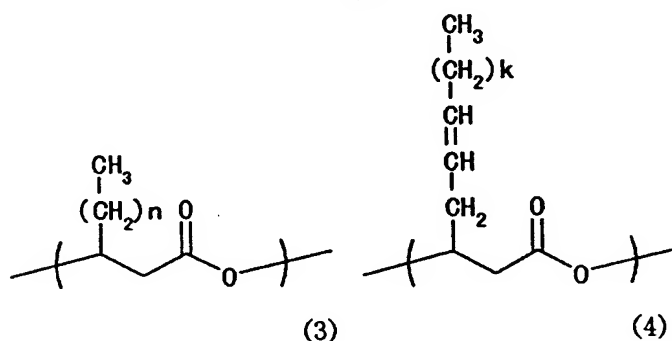


wherein A is represented by General Formula (2), B is at least one selected from the group consisting of monomer units represented by General Formula (3) or (4), and m has a value of 0.01 or larger and smaller than 1:

10



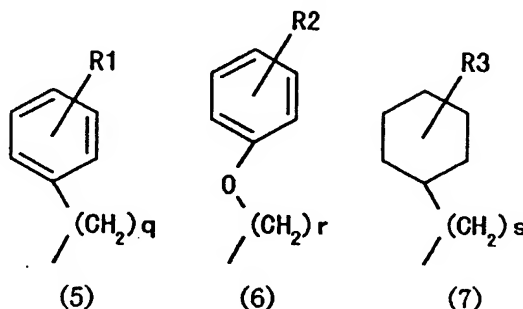
15



wherein

n has a value of 0 to 10, k has a value of 3 or 5, and R is at least one group selected from the group consisting of groups represented by General Formulae (5) to (7):

25



in Formula (5)

R1 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and q is an integer selected from 1 to 8;

5 in Formula (6)

R2 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and r is an integer selected from 1 to 8;

in Formula (7)

10 R3 is a group selected from the group consisting of a hydrogen atom (H), halogen atoms, -CN, -NO<sub>2</sub>, -CF<sub>3</sub>, -C<sub>2</sub>F<sub>5</sub> and -C<sub>3</sub>F<sub>7</sub>; and s is an integer selected from 1 to 8.

The efficient production methods are also provided.